

Draft Genome Sequence of *Klebsiella pneumoniae* Strain KP-1

Kai Wei Kelvin Lee,^{a,b} Krithika Arumugam,^a Rikky Wenang Purbojati,^a Qi Xiang Martin Tay,^{a,b} Rohan Benjamin Hugh Williams,^a Staffan Kjelleberg,^{a,b,c} Scott A. Rice^{a,b,c}

Singapore Centre on Environmental Life Sciences Engineering, Nanyang Technological University, Singapore^a; School of Biological Sciences, Nanyang Technological University, Singapore^b; Centre for Marine Bio-Innovation and School of Biotechnology and Biomolecular Sciences, University of New South Wales, Sydney, Australia^c

***Klebsiella pneumoniae* is ubiquitous in the environment and is a member of a three-species biofilm model. We compared the genome sequence of an environmental isolate, *K. pneumoniae* strain KP-1, to those of two clinical strains (NTUH-K2044 and MGH 78578). KP-1 possesses strain-specific prophage sequences that distinguish it from the clinical strains.**

Received 14 November 2013 Accepted 18 November 2013 Published 19 December 2013

Citation Lee KWK, Arumugam K, Purbojati RW, Tay QXM, Williams RBH, Kjelleberg S, Rice SA. 2013. Draft genome sequence of *Klebsiella pneumoniae* strain KP-1. *Genome Announc.* 1(6):e01082-13. doi:10.1128/genomeA.01082-13.

Copyright © 2013 Lee et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 3.0 Unported license](https://creativecommons.org/licenses/by/3.0/).

Address correspondence to Scott A. Rice, rscott@ntu.edu.sg.

Klebsiella pneumoniae is ubiquitous in the environment, where it is involved in nitrogen fixation (1). It also causes bovine mastitis (2) and nosocomial infections in humans (3). It coexists with *Pseudomonas aeruginosa* and *Pseudomonas protegens* in metalworking fluids and the gut of the silk moth *Bombyx mori* (4, 5). A reproducible, mixed-species biofilm model comprising *K. pneumoniae* KP-1, *P. aeruginosa* PAO1, and *P. protegens* Pf-5 was thus developed to study bacterial interspecies interactions and their effects on biofilm development and fitness (6). While the complete genome sequences of PAO1 and Pf-5 were available to facilitate “omics” studies such as transcriptomics, KP-1 is a newly isolated environmental isolate, and its genome has not been sequenced previously.

The strain was shotgun sequenced on a 454 GS-FLX sequencing platform (Roche, Basel, Switzerland) and an Illumina MiSeq benchtop sequencer (Illumina, CA, USA). The reads from both platforms were trimmed, and *de novo* assembly was performed using Newbler v2.6 (Roche). With 1,673,246 and 1,840,620 reads from the 454 GS-FLX and MiSeq sequencing platforms, respectively, 24 contigs with a total length of 5,131,085 bp and an average GC content of 57.6% were assembled. The open reading frames (ORFs) were predicted using Glimmer v3.02 (7) and annotated by performing BLASTX analysis (*E* value $<10^{-3}$, $>80\%$ identity) against the nonredundant protein sequence database of the National Center for Biotechnology Information. The tRNAs were predicted using tRNAscan-SE v1.3.1 (8), while rRNA sequences were identified by RNAmmer v1.2 (9). A total of 4,587 ORFs were annotated in addition to 73 tRNAs, five 5S rRNAs, and one 23S rRNA. Genomic comparisons to two clinical isolates, *K. pneumoniae* strain NTUH-K2044 (GenBank accession number AP006725) and strain MGH 78578 (GenBank accession number CP000647), by mGenomeSubtractor v1.3 (*H* value <0.42) (10) showed that KP-1 possesses several strain-specific genes that encode phage-related proteins such as tail proteins, capsid proteins, terminases, and integrases. Differences between the prophage sequences of strain NTUH-K2044 and strain MGH 78578 were previously reported and suggested to be markers that could be used to

distinguish the various lineages of *K. pneumoniae* (11). The present study further suggests that prophage sequences could be the ideal evolutionary markers for identifying the different lineages of *K. pneumoniae*.

Nucleotide sequence accession numbers. This whole-genome shotgun project has been deposited in DDBJ/EMBL/GenBank under the accession number [AVNZ00000000](https://www.ncbi.nlm.nih.gov/nuccore/AVNZ00000000). The version described in this paper is version AVNZ01000000.

ACKNOWLEDGMENTS

Kai Wei Kelvin Lee was supported by the National Research Foundation Singapore under the National Research Foundation (NRF) Environmental and Water Technologies (EWT) Ph.D. Scholarship Programme administered by the Environment and Water Industry Programme Office (EWI). We acknowledge financial support from the Singapore Centre on Environmental Life Sciences Engineering (SCELSE), whose research is supported by the National Research Foundation Singapore, Ministry of Education, Nanyang Technological University and National University of Singapore, under its Research Centre of Excellence Program.

REFERENCES

1. Mahl MC, Wilson PW, Fife MA, Ewing WH. 1965. Nitrogen fixation by members of the tribe *Klebsiellae*. *J. Bacteriol.* 89:1482–1487.
2. Braman SK, Eberhart RJ, Asbury MA, Hermann GJ. 1973. Capsular types of *Klebsiella pneumoniae* associated with bovine mastitis. *J. Am. Vet. Med. Assoc.* 162:109–111.
3. Podschun R, Ullmann U. 1998. *Klebsiella* spp. as nosocomial pathogens: epidemiology, taxonomy, typing methods, and pathogenicity factors. *Clin. Microbiol. Rev.* 11:589–603.
4. Chazal PM. 1995. Pollution of modern metalworking fluids containing biocides by pathogenic bacteria in France. Reexamination of chemical treatments accuracy. *Eur. J. Epidemiol.* 11:1–7.
5. Anand AA, Vennison SJ, Sankar SG, Prabhui DI, Vasan PT, Raghuraman T, Geoffrey CJ, Vendan SE. 2010. Isolation and characterization of bacteria from the gut of *Bombyx mori* that degrade cellulose, xylan, pectin and starch and their impact on digestion. *J. Insect Sci.* 10:107.
6. Lee KWK, Periasamy S, Mukherjee M, Xie C, Kjelleberg S, Rice SA. 24 October 2013. Biofilm development and enhanced stress resistance of a model, mixed-species community biofilm. *ISME J.* doi:10.1038/ismej.2013.194.
7. Delcher AL, Bratke KA, Powers EC, Salzberg SL. 2007. Identifying

- bacterial genes and endosymbiont DNA with Glimmer. *Bioinformatics* 23:673–679.
8. Lowe TM, Eddy SR. 1997. tRNAscan-SE: a program for improved detection of transfer RNA genes in genomic sequence. *Nucleic Acids Res.* 25: 955–964.
 9. Lagesen K, Hallin P, Rødland EA, Staerfeldt HH, Rognes T, Ussery DW. 2007. RNAmmer: consistent and rapid annotation of ribosomal RNA genes. *Nucleic Acids Res.* 35:3100–3108.
 10. Shao Y, He X, Harrison EM, Tai C, Ou HY, Rajakumar K, Deng Z. 2010. mGenomeSubtractor: a web-based tool for parallel in silico subtractive hybridization analysis of multiple bacterial genomes. *Nucleic Acids Res.* 38:W194–W200.
 11. Wu KM, Li LH, Yan JJ, Tsao N, Liao TL, Tsai HC, Fung CP, Chen HJ, Liu YM, Wang JT, Fang CT, Chang SC, Shu HY, Liu TT, Chen YT, Shiao YR, Lauderdale TL, Su IJ, Kirby R, Tsai SF. 2009. Genome sequencing and comparative analysis of *Klebsiella pneumoniae* NTUH-K2044, a strain causing liver abscess and meningitis. *J. Bacteriol.* 191: 4492–4501.