

DIY spectrophotometry: Rapid, low-cost RGB approach for vis-spectroscopy, OD600, and fluorescence quantification in profiling nosocomial antibiotic resistance (MRSA)

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DIY spectrophotometry: Rapid, Low-cost RGB approach for Vis-spectroscopy, OD₆₀₀, and Fluorescence Quantification in profiling nosocomial antibiotic resistance (MRSA)

INTRODUCTION

PROBLEM: Existing spectrophotometric assays are prohibitively expensive and incompatible with resource-limited settings (i.e. developing countries, citizen science and field testing), thereby necessitating a viable, economical alternative capable of yielding accurate and reproducible results.

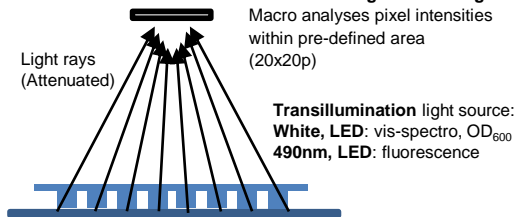
The increasing prevalence of multi-drug resistance has made the rapid & accurate determination of antibiotic susceptibility an imperative. In particular, Methicillin-resistant *S. aureus* (MRSA), a nosocomial pathogen, is responsible for significant inpatient morbidity and mortality [3.2 cases/1000 inpatient days] [1].

AIM: Herein, an affordable, high-throughput, semi-automated method of determining *S. aureus* antibiotic susceptibility via RGB analysis is described.

OD₆₀₀ (MIC) values from broth microdilution, and end-point fluorescence values from strain-typing (PCR) were quantified, yielding absorbance readings comparable to commercial spectrophotometers (R² correlation coefficient = 0.9931, 0.9539 respectively)

METHOD

SETUP: Image capture



RGB Weighting:

- Existing RGB protocols approximate MPIs to a single channel → variable chromophore sensitivity, limited accuracy and reproducibility.
- A novel solution of it is proposed herein: Each RGB channel is weighed (%) based on min/max channel-specific MPI values to yield weighted MPIs → greater R² correlation coefficients by effectively 'diluting' anomalous readings in any given channel & improves sensitivity to chromophores not overtly R/G/B.

$$\text{Weightage Factor}_c = \frac{|\text{Max}_{MPI} - \text{Min}_{MPI}|_c}{\sum_{i=c(R,G,B)}^{all\ c} |\text{Max}_{MPI} - \text{Min}_{MPI}|_i}$$

$$\text{Weighted MPI} = \sum_{i=c(R,G,B)}^{all\ c} \text{MPI}_i \times \text{Weightage factor}_i$$

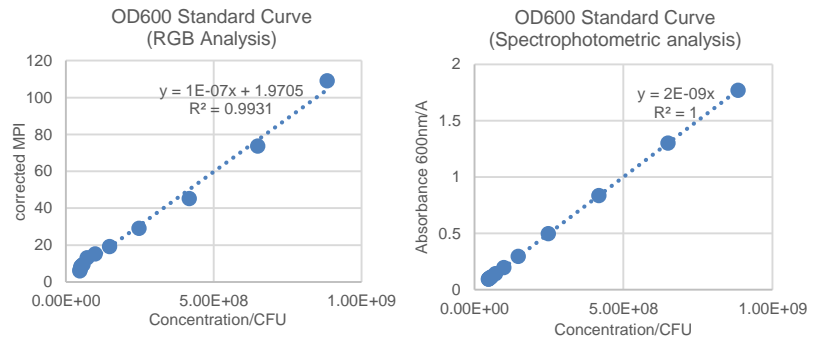
Blank Compensation for Vignetting, Variable Transillumination pathways: Correction factors facilitate the empirical compensation of visual aberrations.

$$\text{Correction Factor (well}_n) = \frac{\text{Max MPI}_{(across\ all\ wells)}}{\text{MPI}_n}$$

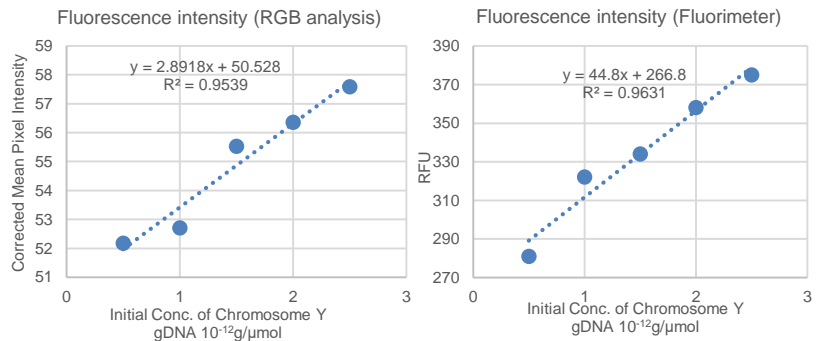
$$\text{Corrected MPI} = \text{Weighted MPI} \times \text{Correction Factor}$$

RESULTS

PHENOTYPIC AST: OD₆₀₀ (Min. Inhibitory Concentration) – Comparison of OD₆₀₀ *S. aureus* broth microdilutions processed using RGB analysis vs commercial spectrophotometry.



GENOTYPIC AST: End-point fluorescence Intensity – Comparison of end-point fluorescence of *y*-chromosome gDNA PCR amplicons (43 cycles). 485nm excitation and 525nm emission wavelengths used for commercial fluorimeter analysis.



DISCUSSION

The accuracy and reproducibility of RGB analysis is comparable to commercial spectrophotometry [R² of 0.9931 vs 1 for OD₆₀₀, R² of 0.9539 vs 0.9631 for fluorescence intensity]. Discrepancies observed at weaker (attenuated) light intensities is attributable to the superimposition of gamma-correction [(I_{out} = I_{in}^{1/2.2})] causing non-linear amplification of RGB values. This disproportionately affects extremely low and high intensity MPI values, amplifying and attenuating them respectively.

The quantifiable range of OD₆₀₀ values corresponds the detection range of **0.0975A – 1.2995A**; for fluorescence, this is **281-375 RFU**; this substantive, reproducible intrapolated range of values would enable the low-cost, rapid and reliable determination of *S. aureus* resistance profiles through both phenotype and genotype:

- High-throughput antibiotic (Methicillin) susceptibility tests in broth microdilutions
- End-point fluorescence of MRSA qPCR (mecA/ BBP2 gene expression)

FUTURE WORK: Integrated Attachment + App on platform (iOS)

> Portable, lightweight, passive attachment (incorporating diffraction grating/wavelength selection) + iSpectro app

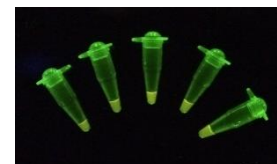


Figure 1: Fluorescent samples of human gDNA under 485nm transillumination