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2021

Beloin, C. & McDougald, D. (2021). Speciality grand challenge for “biofilms” . *Frontiers in Cellular and Infection Microbiology*, 11, 632429-.

<https://dx.doi.org/10.3389/fcimb.2021.632429>

<https://hdl.handle.net/10356/151791>

<https://doi.org/10.3389/fcimb.2021.632429>

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Specialty Grand Challenge for “Biofilms”

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Keywords: biofilm infections, multi-species biofilm, infection models, biofilm detection methods, biofilm heterogeneity

INTRODUCTION TO BIOFILMS - PROPERTIES OF BIOFILMS

The idea that most bacteria in nature are attached to surfaces was recognized as early as 1933 when Henrici stated after investigating fresh-water bacteria that “it is quite evident that for the most part water bacteria are not free floating organisms, but grow attached upon submerged surfaces” (Henrici, 1933). Zobell followed up with work that demonstrated seawater biofilms precede the attachment of other macro-fouling organisms (Zobell and Allen, 1935). Thus, began this field of research which was furthered by work on biofilms occurring in nature and the growing realization that a number of infections are biofilm-related (Costerton et al., 1999; Hall-Stoodley et al., 2004).

Biofilm Life Cycle

Biofilms are structurally diverse and dynamic (Hall-Stoodley et al., 2004). Biofilm formation occurs in stages beginning with the attachment of a cell on the surface, followed by division and secretion of extracellular polymeric substances (EPS) and formation of microcolonies (Costerton et al., 1987). There are channels that allow for exchange of nutrients and cellular waste between the microcolonies. The final stage of the biofilm lifecycle is dispersal which is driven by, but not limited to environmental cues and bacterially derived signals (reviewed in (McDougald et al., 2011). These signals include the second messenger cyclic-di-GMP (Jenal et al., 2017), nitric oxide (Barraud et al., 2009) and quorum sensing (Rice et al., 2005).

Increased Resistance to Antimicrobials and Matrix Properties

Biofilms are more resistant to a variety of stresses than their planktonic counterparts. For example, biofilms have been reported to be up to 1000-fold more resistant to antibiotics than planktonic cells (Costerton et al., 1999) as well as showing increased resistance to other environmental stresses and predation by protists (Matz et al., 2005). This increased resistance is due to changes in cellular metabolism of biofilm cells compared to planktonic cells (Lebeaux et al., 2014; Crabbé et al., 2019). It is also due to the physical protection provided by the extracellular matrix (Flemming and Wingender, 2010).

The biofilm matrix is composed of a variety of biopolymers, including polysaccharides, proteins, nucleic acids, lipids, and other biopolymers such as humic substances (Flemming and Wingender, 2010). These components can vary widely between different bacterial species. The polysaccharides and proteins help to protect biofilm cells from antimicrobials by acting as a barrier to their penetration into the biofilm. *Pseudomonas aeruginosa* requires extracellular DNA or eDNA to form a biofilm and for motility within the biofilm (Whitchurch et al., 2002; Gloag et al., 2013). It is this matrix that lends stability to the biofilm making them difficult to remove. The matrix can also bind

OPEN ACCESS

Edited and Reviewed by:

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Specialty section:

This article was submitted to
Biofilms,
a section of the journal
Frontiers in Cellular and
Infection Microbiology

Received: 23 November 2020

Accepted: 28 January 2021

Published: 22 February 2021

Citation:

Beloin C and McDougald D (2021)
Specialty Grand
Challenge for “Biofilms”.
Front. Cell. Infect. Microbiol. 11:632429.
doi: 10.3389/fcimb.2021.632429

extracellular enzymes making them available to biofilm cells and sequester other damaging compounds.

CHALLENGES IN METHODS FOR INVESTIGATING BIOFILMS

From Monospecies to Polymicrobial Biofilms

Since the early observation of biofilms in their natural environments, notably in rivers, researchers have realized the heterogeneity and complexity of biofilm composition and structure. After a long period during which monospecies biofilms have been studied with success we have moved towards the study of polymicrobial biofilms, which are also relevant in multiple biofilm-associated infections in animals and humans and often at the origin of chronic infections (Peters et al., 2012; Wolcott et al., 2013). Among the multitude of *in vitro* and *in vivo* models of biofilm formation and infection models that have been developed (Coenye and Nelis, 2010; Lebeaux et al., 2013; Brackman and Coenye, 2016) some specific models are dedicated to polymicrobial biofilms (Gabriliska and Rumbaugh, 2015; Tay et al., 2016). The relevance of studying polymicrobial biofilms was demonstrated by the importance of the interactions of micro-organisms for the development of biofilms in the context of infection, notably through the cross-talk between species and their impact on mechanisms of antimicrobial recalcitrance (Kragh et al., 2016; Liu et al., 2016; Orazi and O'Toole, 2019; Ibberson and Whiteley, 2020). Further developments of such models are essential to uncover the molecular mechanisms beyond the specific behavior of micro-organisms during such polymicrobial biofilms.

Tackling the Heterogeneity of Biofilms

In addition to the complexity of polymicrobial biofilms, one hallmark of biofilms in general is their heterogeneity in terms of local environmental conditions that translates into a heterogeneity of physiology of biofilm cells. While studies using technologies such as transcriptomics, proteomics and metabolomics have been successfully applied to image the global physiology or responses to stress of biofilms cells (Seneviratne et al., 2020), there is a clear need to think singularity and to develop technologies that would allow for the study of the physiology of individual biofilm cells and to understand the social interactions within biofilms (Rode et al., 2020). These studies can be performed with the improvement of methods such as single-cell RNA transcriptomics (Blattman et al., 2020; Imdahl et al., 2020) and technologies used to isolate cells from biofilms (Ma et al., 2019). Such recently developed technologies have been successfully applied to image the physiology of *S. aureus* persister cells within macrophages and should be applied hopefully to biofilms. Imaging technologies have also recently improved to locate and trace individual cells within biofilms. Multiplex FISH technology has been applied to characterize the biogeography of the oral microbiota (Valm et al., 2012; Valm, 2019) while recent microscopy imaging and analyzing technologies allowing for dynamically tracking single biofilm bacteria (Hartmann et al., 2019), for example upon antibiotic stress in *V. cholerae* (Díaz-Pascual et al., 2019).

Additionally, some very promising tools have been recently adapted to metabolite identification at the single cell level in biofilm, for instance using Raman spectroscopy, Mass spectrometry or electro-chemical chip/fluorophores (Baig et al., 2016; Bellin et al., 2016; Bodelón et al., 2016; Schiessl et al., 2019; Geier et al., 2020; Yang et al., 2020). The upcoming challenge will be, in addition to further development of these technologies, to adapt them to direct observation or characterization of *in situ* biofilms and to models, whether *in vitro*, *ex vivo*, or *in vivo*, that better feature real *in situ* biofilms.

Going Toward a Better Understanding of *In Vivo* Biofilms: Improving Detection and Observation of *In Situ* Biofilms and Developing Novel Models?

Today there is absolutely no doubt that biofilms occur *in vivo*, especially in compromised hosts, with almost 80% of human infections now recognized to be biofilm-related (NIH, 2002). Biofilms form in body compartments as well as attached to cells and indwelling devices (Bjarnsholt et al., 2013). One well-studied example of this is the infection of cystic fibrosis (CF) patients' lungs by *P. aeruginosa*. These biofilms are extremely resistant to antimicrobials for reasons discussed above but also due to their ability to resist host defenses. For example, in both *in vitro* and *in vivo* CF models, *P. aeruginosa* was shown to respond to the presence of host immune cells by upregulating a number of quorum-sensing regulated virulence factors such as rhamnolipids which are known to kill immune cells (Alhede et al., 2009). Other common chronic infections include chronic otitis media and chronic wounds that tend to be caused by multispecies infections (Burmølle et al., 2010). Other major sources of *in vivo* biofilms are indwelling medical devices such as intravenous or urinary catheters, pacemakers, endotracheal tubes and artificial joints (Lebeaux et al., 2014; Stewart and Bjarnsholt, 2020). Microorganisms commonly associated with medical devices are *Staphylococcus epidermidis*, *Staphylococcus aureus*, and *P. aeruginosa* (Hall-Stoodley et al., 2004). These biofilms often result in replacement of the device, which for artificial joints is a significant task. The dispersal of these biofilms has been discussed as a potential treatment, however a large number of planktonic cells entering the blood stream may be more dangerous than other treatments. Numerous infections are also related to biofilms in animals, for example in bovine mastitis, pneumonia, liver abscess, enteritis, urinary tract infection, otitis, wound infection, etc. (Abdullahi et al., 2016) and in plants where bacterial and fungal pathogens colonization can be at the origin of different diseases (Bogino et al., 2013; Castiblanco and Sundin, 2016; Motaung et al., 2020).

For decades our vision of how biofilms look has been governed by our knowledge developed from the use of *in vitro* biofilms models. As stated above, the increasing number of biofilms isolated from real infection situations have brought into question this conventional thinking with the recognition that, with some exception, *in vivo* biofilms rather correspond to patches or cell aggregates that rarely exceed 200 μm (Bjarnsholt et al., 2013) with important consequences on how biofilms finally interact with the host (Alhede et al., 2020). These observations have led to several important statements: in most cases, *in vitro*,

and sometimes *in vivo* biofilms models do not relevantly recapitulate biofilms in their real infectious context; the diagnosis of the presence of biofilms linked to infection cannot be made *a posteriori* if we want clinicians to adapt their treatment to the presence of a biofilm. With these statements come potential actions: on one side the necessity to improve detection methods that could, for some of them, allow early detection of biofilms in the host and, for others, give a better characterization of the environment of infectious biofilms. Thus, the need to implement some of the *in situ* information into the current *in vitro* and *in vivo* models, and to develop novel models is paramount. It is clear that to this day some effort has been made in these directions, but we will have to do much more to reach these objectives.

We are still trying to identify biofilm biomarkers that could be used not only for detection but also for vaccine development. Important work has been done in the laboratory of the late Prof. Mark Shirliff in this direction notably with a recent demonstration that *S. aureus* orthopedic implant infections could be detected in human sinovial fluids using an antibody against the manganese transporter MntC (Harro et al., 2019; Harro et al., 2020). Peptide-fluorescently labeled probes have been recently identified and used to specifically label *P. aeruginosa* biofilms *in vivo* (Locke et al., 2020). Additionally, numerous other detection methods are under development and efforts should be made to allow these technologies to translate into the clinic (Achinas et al., 2020; Parlak and Richter-Dahlfors, 2020). Among them, recently there has been the successful use of imaging technologies using bioluminescence (Chauhan et al., 2016; Hoffmann et al., 2019; Maiden et al., 2019; Gordon et al., 2020; Kreth et al., 2020; Redman et al., 2020; Van Dyck et al., 2020) or intravital imaging (Thanabalasuriar et al., 2019; Abdul Hamid et al., 2020; Gries et al., 2020; Tian et al., 2020) to study the effect of antimicrobial agents on biofilms or the behavior of immune cells in contact with biofilms in various *in vivo* models of biofilm-related infections.

The concept of the micro-environment of biofilm cells during infection has been well conceptualized in a recent perspective article on chronic wounds where several defined zones can be identified and on which information should be gathered in order to access to a global view of the biofilm infectious micro-environment (Kirketerp-Møller et al., 2020). Ideally, non-destructive methods to sample and analyze this micro-environment using micro-dialysis, micro-probes or electrochemical probes should be adapted to *in vivo* measurement as recently explained in (Røder et al., 2020), however, some of these methods also have applications for biofilm detection.

Improving the models for a better understanding of biofilm formation and biofilm cells behavior is also a major objective that should be reached by researchers in the field. Initiatives to evaluate the available models are welcome and should provide a better view

of the current more relevant models with some clues on how to improve them (Cornforth et al., 2020). With the increased pressure to restrict the use of animals in research there is also a dire need to identify surrogate models. While there is some utility to pursue the development of non-mammalian models of biofilm-infection such as *Drosophila melanogaster*, *Galleria mellonella* or *Danio rerio* (Zebrafish) (Lebeaux et al., 2013), identification of *ex vivo* models that recreate the infectious environment should be encouraged. Among recently developed *ex vivo* models are, for example, an *ex vivo* pig lung biofilm model used for understanding antibiotic tolerance of *P. aeruginosa* biofilms (Hassan et al., 2020) or an *ex vivo* murine skin biopsy model to characterize *B. burgdorferi* biofilms, the etiological agent of Lyme disease (Torres et al., 2020). Last but not least with the development of reconstituted organs such as organoids or organ-on-a-chip, one could anticipate that adaptations of these models to the understanding of the physiopathology of biofilm infections, interactions of biofilms with the immune response or biofilm behaviors in the presence of antimicrobials would provide important information relevant to biofilms in infectious contexts (Jimi et al., 2017; Choi et al., 2020; Yuan et al., 2020).

Major Challenges

As described above, the field of biofilms and interactions with higher organisms has clear challenges for future research. 1) The development of new and improved multi-species biofilm models will allow us to investigate interactions that occur in these biofilms, as most *in vivo* biofilms are multispecies biofilms. This is clearly the next step in understanding biofilms in nature and in disease contexts, and this should come with the use of technologies that would allow probing the infectious micro-environment of biofilms. 2) The development of more relevant models, whether *in vitro*, *ex vivo*, or *in vivo*, to elucidate interactions of biofilms with the host in more detail. These models will be necessary not only for understanding biofilm host interactions but also for the investigation of eradication methods or compounds in a more real setting. 3) The development of imaging and detection technologies that could be used to better visualize and detect biofilms *in situ* and should allow when necessary to access the physiology on individual cells within biofilms. As new methods are developed, using these for biofilm research will generate new knowledge and lead to new areas of biofilm research.

AUTHOR CONTRIBUTIONS

CB and DM wrote the manuscript. All authors contributed to the article and approved the submitted version.

REFERENCES

- Abdul Hamid, A. I., Nakusi, L., Givskov, M., Chang, Y. T., Marquès, C., and Geirard, P. (2020). A mouse ear skin model to study the dynamics of innate immune responses against *Staphylococcus aureus* biofilms. *BMC Microbiol.* 20, 22. doi: 10.1186/s12866-019-1635-z
- Abdullahi, U. F., Igwenagu, E., Mu'azu, A., Aliyu, S., and Umar, M. I. (2016). Intrigues of biofilm: A perspective in veterinary medicine. *Vet. World* 9, 12–18. doi: 10.14202/vetworld.2016.12-18
- Achinas, S., Yska, S. K., Charalampogiannis, N., Krooneman, J., and Euverink, G. J. W. (2020). A Technological Understanding of Biofilm Detection Techniques: A Review. *Mater. (Basel)* 13(14):3417. doi: 10.3390/ma13143147

- Alhede, M., Bjarnsholt, T., Jensen, P., Phipps, R. K., Moser, C., Christophersen, L., et al. (2009). *Pseudomonas aeruginosa* recognizes and responds aggressively to the presence of polymorphonuclear leukocytes. *Microbiol. (Reading)* 155, 3500–3508. doi: 10.1099/mic.0.031443-0
- Alhede, M., Alhede, M., Qvortrup, K., Kragh, K. N., Jensen, P., Stewart, P. S., et al. (2020). The origin of extracellular DNA in bacterial biofilm infections in vivo. *Pathog. Dis.* 78(2):ftaa018. doi: 10.1093/femspd/ftaa018
- Baig, N., Poliseti, S., Morales-Soto, N., Dunham, S. J. B., Sweedler, J. V., Shrout, J. D., et al. (2016). Label-free molecular imaging of bacterial communities of the opportunistic pathogen *Pseudomonas aeruginosa*. *Proc. SPIE Int. Soc. Opt. Eng.* 9930:993004. doi: 10.1117/12.2236695
- Barraud, N., Storey, M. V., Moore, Z. P., Webb, J. S., Rice, S. A., and Kjelleberg, S. (2009). Nitric oxide-mediated dispersal in single- and multi-species biofilms of clinically and industrially relevant microorganisms. *Microb. Biotechnol.* 2, 370–378. doi: 10.1111/j.1751-7915.2009.00098.x
- Bellin, D. L., Sakhtab, H., Zhang, Y., Price-Whelan, A., Dietrich, L. E., and Shepard, K. L. (2016). Electrochemical camera chip for simultaneous imaging of multiple metabolites in biofilms. *Nat. Commun.* 7, 10535. doi: 10.1038/ncomms10535
- Bjarnsholt, T., Alhede, M., Alhede, M., Eickhardt-Sørensen, S. R., Moser, C., Kühl, M., et al. (2013). The in vivo biofilm. *Trends Microbiol.* 21, 466–474. doi: 10.1016/j.tim.2013.06.002
- Blattman, S. B., Jiang, W., Oikonomou, P., and Tavazoie, S. (2020). Prokaryotic single-cell RNA sequencing by in situ combinatorial indexing. *Nat. Microbiol.* 5, 1192–1201. doi: 10.1038/s41564-020-0729-6
- Bodelón, G., Montes-García, V., López-Puente, V., Hill, E. H., Hamon, C., Sanz-Ortiz, M. N., et al. (2016). Detection and imaging of quorum sensing in *Pseudomonas aeruginosa* biofilm communities by surface-enhanced resonance Raman scattering. *Nat. Mater.* 15, 1203–1211. doi: 10.1038/nmat4720
- Bogino, P. C., Oliva Mde, L., Sorroche, F. G., and Giordano, W. (2013). The role of bacterial biofilms and surface components in plant-bacterial associations. *Int. J. Mol. Sci.* 14, 15838–15859. doi: 10.3390/ijms140815838
- Brackman, G., and Coenye, T. (2016). In Vitro and In Vivo Biofilm Wound Models and Their Application. *Adv. Exp. Med. Biol.* 897, 15–32. doi: 10.1007/5584_2015_5002
- Burmölle, M., Thomsen, T. R., Fazli, M., Dige, I., Christensen, L., Homøe, P., et al. (2010). Biofilms in chronic infections - a matter of opportunity - monospecies biofilms in multispecies infections. *FEMS Immunol. Med. Microbiol.* 59, 324–336. doi: 10.1111/j.1574-695X.2010.00714.x
- Castiblanco, L. F., and Sundin, G. W. (2016). New insights on molecular regulation of biofilm formation in plant-associated bacteria. *J. Integr. Plant Biol.* 58, 362–372. doi: 10.1111/jipb.12428
- Chauhan, A., Ghigo, J. M., and Beloin, C. (2016). Study of in vivo catheter biofilm infections using pediatric central venous catheter implanted in rat. *Nat. Protoc.* 11, 525–541. doi: 10.1038/nprot.2016.033
- Choi, K. G., Wu, B. C., Lee, A. H., Baquir, B., and Hancock, R. E. W. (2020). Utilizing Organoid and Air-Liquid Interface Models as a Screening Method in the Development of New Host Defense Peptides. *Front. Cell Infect. Microbiol.* 10, 228. doi: 10.3389/fcimb.2020.00228
- Coenye, T., and Nelis, H. J. (2010). In vitro and in vivo model systems to study microbial biofilm formation. *J. Microbiol. Methods* 83, 89–105. doi: 10.1016/j.mimet.2010.08.018
- Cornforth, D. M., Diggle, F. L., Melvin, J. A., Bomberger, J. M., and Whiteley, M. (2020). Quantitative Framework for Model Evaluation in Microbiology Research Using *Pseudomonas aeruginosa* and Cystic Fibrosis Infection as a Test Case. *mBio* 11(1):e03042-19. doi: 10.1128/mBio.03042-19
- Costerton, J. W., Cheng, K. J., Geesey, G. G., Ladd, T. I., Nickel, J. C., Dasgupta, M., et al. (1987). Bacterial biofilms in nature and disease. *Annu. Rev. Microbiol.* 41, 435–464. doi: 10.1146/annurev.mi.41.100187.002251
- Costerton, J. W., Stewart, P. S., and Greenberg, E. P. (1999). Bacterial biofilms: a common cause of persistent infections. *Science* 284, 1318–1322. doi: 10.1126/science.284.5418.1318
- Crabbé, A., Jensen, P., Bjarnsholt, T., and Coenye, T. (2019). Antimicrobial Tolerance and Metabolic Adaptations in Microbial Biofilms. *Trends Microbiol.* 27, 850–863. doi: 10.1016/j.tim.2019.05.003
- Díaz-Pascual, F., Hartmann, R., Lempp, M., Vidakovic, L., Song, B., Jeckel, H., et al. (2019). Breakdown of *Vibrio cholerae* biofilm architecture induced by antibiotics disrupts community barrier function. *Nat. Microbiol.* 4, 2136–2145. doi: 10.1038/s41564-019-0579-2
- Flemming, H. C., and Wingender, J. (2010). The biofilm matrix. *Nat. Rev. Microbiol.* 8, 623–633. doi: 10.1038/nrmicro2415
- Gabrilska, R. A., and Rumbaugh, K. P. (2015). Biofilm models of polymicrobial infection. *Future Microbiol.* 10, 1997–2015. doi: 10.2217/fmb.15.109
- Geier, B., Sogin, E. M., Michellod, D., Janda, M., Kompauer, M., Spengler, B., et al. (2020). Spatial metabolomics of in situ host-microbe interactions at the micrometre scale. *Nat. Microbiol.* 5, 498–510. doi: 10.1038/s41564-019-0664-6
- Gloag, E. S., Turnbull, L., Huang, A., Vallotton, P., Wang, H., Nolan, L. M., et al. (2013). Self-organization of bacterial biofilms is facilitated by extracellular DNA. *Proc. Natl. Acad. Sci. U.S.A.* 110, 11541–11546. doi: 10.1073/pnas.1218898110
- Gordon, O., Miller, R. J., Thompson, J. M., Ordóñez, A. A., Klunk, M. H., Dikeman, D. A., et al. (2020). Rabbit model of *Staphylococcus aureus* implant-associated spinal infection. *Dis. Model Mech.* 13(7):dmm045385. doi: 10.1242/dmm.045385
- Gries, C. M., Rivas, Z., Chen, J., and Lo, D. D. (2020). Intravital Multiphoton Examination of Implant-Associated *Staphylococcus aureus* Biofilm Infection. *Front. Cell Infect. Microbiol.* 10, 574092. doi: 10.3389/fcimb.2020.574092
- Hall-Stoodley, L., Costerton, J. W., and Stoodley, P. (2004). Bacterial biofilms: from the natural environment to infectious diseases. *Nat. Rev. Microbiol.* 2, 95–108. doi: 10.1038/nrmicro821
- Harro, J. M., Achermann, Y., Freiberg, J. A., Allison, D. L., Brao, K. J., Marinos, D. P., et al. (2019). Clearance of *Staphylococcus aureus* from In Vivo Models of Chronic Infection by Immunization Requires Both Planktonic and Biofilm Antigens. *Infect. Immun.* 88(1):e00586-19. doi: 10.1128/IAI.00586-19
- Harro, J. M., Shirliff, M. E., Arnold, W., Kofonow, J. M., Dammling, C., Achermann, Y., et al. (2020). Development of a Novel and Rapid Antibody-Based Diagnostic for Chronic *Staphylococcus aureus* Infections Based on Biofilm Antigens. *J. Clin. Microbiol.* 58(5):e01414-19. doi: 10.1128/JCM.01414-19
- Hartmann, R., Singh, P. K., Pearce, P., Mok, R., Song, B., Díaz-Pascual, F., et al. (2019). Emergence of three-dimensional order and structure in growing biofilms. *Nat. Phys.* 15, 251–256. doi: 10.1038/s41567-018-0356-9
- Hassan, M. M., Harrington, N. E., Sweeney, E., and Harrison, F. (2020). Predicting Antibiotic-Associated Virulence of *Pseudomonas aeruginosa* Using an ex vivo Lung Biofilm Model. *Front. Microbiol.* 11, 568510. doi: 10.3389/fmicb.2020.568510
- Henrici, A. T. (1933). Studies of Freshwater Bacteria: I. A Direct Microscopic Technique. *J. Bacteriol.* 25, 277–287. doi: 10.1128/JB.25.3.277-287.1933
- Hoffmann, J. P., Friedman, J. K., Wang, Y., Mclachlan, J. B., Sammarco, M. C., Morici, L. A., et al. (2019). In situ Treatment With Novel Microbiocide Inhibits Methicillin Resistant *Staphylococcus aureus* in a Murine Wound Infection Model. *Front. Microbiol.* 10, 3106. doi: 10.3389/fmicb.2019.03106
- Ibberson, C. B., and Whiteley, M. (2020). The social life of microbes in chronic infection. *Curr. Opin. Microbiol.* 53, 44–50. doi: 10.1016/j.mib.2020.02.003
- Imdahl, F., Vafadarnejad, E., Homberger, C., Saliba, A. E., and Vogel, J. (2020). Single-cell RNA-sequencing reports growth-condition-specific global transcriptomes of individual bacteria. *Nat. Microbiol.* 5, 1202–1206. doi: 10.1038/s41564-020-0774-1
- Jenal, U., Reinders, A., and Lori, C. (2017). Cyclic di-GMP: second messenger extraordinaire. *Nat. Rev. Microbiol.* 15, 271–284. doi: 10.1038/nrmicro.2016.190
- Jimi, S., Miyazaki, M., Takata, T., Ohjimi, H., Akita, S., and Hara, S. (2017). Increased drug resistance of methicillin-resistant *Staphylococcus aureus* biofilms formed on a mouse dermal chip model. *J. Med. Microbiol.* 66, 542–550. doi: 10.1099/jmm.0.000461
- Kirketerp-Møller, K., Stewart, P. S., and Bjarnsholt, T. (2020). The zone model: A conceptual model for understanding the microenvironment of chronic wound infection. *Wound Repair Regen.* 28, 593–599. doi: 10.1111/wrr.12841
- Kragh, K. N., Hutchison, J. B., Melaugh, G., Rodesney, C., Roberts, A. E., Irie, Y., et al. (2016). Role of Multicellular Aggregates in Biofilm Formation. *mBio* 7, e00237. doi: 10.1128/mBio.00237-16
- Kreth, J., Abdelrahman, Y. M., and Merritt, J. (2020). Multiplex Imaging of Polymicrobial Communities-Murine Models to Study Oral Microbiome Interactions. *Methods Mol. Biol.* 2081, 107–126. doi: 10.1007/978-1-4939-9940-8_8
- Lebeau, D., Chauhan, A., Rendueles, O., and Beloin, C. (2013). From in vitro to in vivo Models of Bacterial Biofilm-Related Infections. *Pathogens* 2, 288–356. doi: 10.3390/pathogens2020288

- Lebeaux, D., Ghigo, J. M., and Beloin, C. (2014). Biofilm-related infections: bridging the gap between clinical management and fundamental aspects of recalcitrance toward antibiotics. *Microbiol. Mol. Biol. Rev.* 78, 510–543. doi: 10.1128/MMBR.00013-14
- Liu, W., Röder, H. L., Madsen, J. S., Bjarnsholt, T., Sørensen, S. J., and Burmölle, M. (2016). Interspecific Bacterial Interactions are Reflected in Multispecies Biofilm Spatial Organization. *Front. Microbiol.* 7, 1366. doi: 10.3389/fmicb.2016.01366
- Locke, L. W., Shankaran, K., Gong, L., Stoodley, P., Vozar, S. L., Cole, S. L., et al. (2020). Evaluation of Peptide-Based Probes toward In Vivo Diagnostic Imaging of Bacterial Biofilm-Associated Infections. *ACS Infect. Dis.* 6, 2086–2098. doi: 10.1021/acinfecdis.0c00125
- Ma, Q., Bücking, H., Gonzalez Hernandez, J. L., and Subramanian, S. (2019). Single-Cell RNA Sequencing of Plant-Associated Bacterial Communities. *Front. Microbiol.* 10, 2452. doi: 10.3389/fmicb.2019.02452
- Maiden, M. M., Zachos, M. P., and Waters, C. M. (2019). Hydrogels Embedded With Melittin and Tobramycin Are Effective Against *Pseudomonas aeruginosa* Biofilms in an Animal Wound Model. *Front. Microbiol.* 10, 1348. doi: 10.3389/fmicb.2019.01348
- Matz, C., McDougald, D., Moreno, A. M., Yung, P. Y., Yildiz, F. H., and Kjelleberg, S. (2005). Biofilm formation and phenotypic variation enhance predation-driven persistence of *Vibrio cholerae*. *Proc. Natl. Acad. Sci. U.S.A.* 102, 16819–16824. doi: 10.1073/pnas.0505350102
- McDougald, D., Rice, S. A., Barraud, N., Steinberg, P. D., and Kjelleberg, S. (2011). Should we stay or should we go: mechanisms and ecological consequences for biofilm dispersal. *Nat. Rev. Microbiol.* 10, 39–50. doi: 10.1038/nrmicro2695
- Motaung, T. E., Peremore, C., Wingfield, B., and Steenkamp, E. (2020). Plant-associated fungal biofilms - knowns and unknowns. *FEMS Microbiol. Ecol.* 96 (12):fiae224. doi: 10.1093/femsec/fiae224
- NIH (2002). RESEARCH ON MICROBIAL BIOFILMS. Office of Extramural Research (OER), National Institutes of Health (NIH), Department of Health and Human Services (HHS Available at: <http://grants.nih.gov/grants/guide/pa-files/PA-03-047.html>).
- Orazi, G., and O'Toole, G. A. (2019). "It Takes a Village": Mechanisms Underlying Antimicrobial Recalcitrance of Polymicrobial Biofilms. *J. Bacteriol.* 202 (1):10.1128/JB.00530-19.
- Parlak, O., and Richter-Dahlfors, A. (2020). Bacterial Sensing and Biofilm Monitoring for Infection Diagnostics. *Macromol. Biosci.* 20, e2000129. doi: 10.1002/mabi.202000129
- Peters, B. M., Jabra-Rizk, M. A., O'may, G. A., Costerton, J. W., and Shirliff, M. E. (2012). Polymicrobial interactions: impact on pathogenesis and human disease. *Clin. Microbiol. Rev.* 25, 193–213. doi: 10.1128/CMR.00013-11
- Redman, W. K., Welch, G. S., and Rumbaugh, K. P. (2020). Differential Efficacy of Glycoside Hydrolases to Disperse Biofilms. *Front. Cell Infect. Microbiol.* 10, 379. doi: 10.3389/fcimb.2020.00379
- Rice, S. A., McDougald, D., Kumar, N., and Kjelleberg, S. (2005). The use of quorum-sensing blockers as therapeutic agents for the control of biofilm-associated infections. *Curr. Opin. Investig. Drugs* 6, 178–184.
- Rode, D. K. H., Singh, P. K., and Drescher, K. (2020). Multicellular and unicellular responses of microbial biofilms to stress. *Biol. Chem.* 401, 1365–1374. doi: 10.1515/hsz-2020-0213
- Röder, H. L., Olsen, N. M. C., Whiteley, M., and Burmölle, M. (2020). Unravelling interspecies interactions across heterogeneities in complex biofilm communities. *Environ. Microbiol.* 22, 5–16. doi: 10.1111/1462-2920.14834
- Schiessl, K. T., Hu, F., Jo, J., Nazia, S. Z., Wang, B., Price-Whelan, A., et al. (2019). Phenazine production promotes antibiotic tolerance and metabolic heterogeneity in *Pseudomonas aeruginosa* biofilms. *Nat. Commun.* 10, 762. doi: 10.1038/s41467-019-08733-w
- Seneviratne, C. J., Suriyanarayanan, T., Widyarman, A. S., Lee, L. S., Lau, M., Ching, J., et al. (2020). Multi-omics tools for studying microbial biofilms: current perspectives and future directions. *Crit. Rev. Microbiol.* 46, 759–778. doi: 10.1080/1040841X.2020.1828817
- Stewart, P. S., and Bjarnsholt, T. (2020). Risk factors for chronic biofilm-related infection associated with implanted medical devices. *Clin. Microbiol. Infect.* 26, 1034–1038. doi: 10.1016/j.cmi.2020.02.027
- Tay, W. H., Chong, K. K., and Kline, K. A. (2016). Polymicrobial-Host Interactions during Infection. *J. Mol. Biol.* 428, 3355–3371. doi: 10.1016/j.jmb.2016.05.006
- Thanabalasuriar, A., Scott, B. N. V., Peiseler, M., Willson, M. E., Zeng, Z., Warrener, P., et al. (2019). Neutrophil Extracellular Traps Confine *Pseudomonas aeruginosa* Ocular Biofilms and Restrict Brain Invasion. *Cell Host Microbe* 25, 526–536.e524. doi: 10.1016/j.chom.2019.02.007
- Tian, S., Su, L., Liu, Y., Cao, J., Yang, G., Ren, Y., et al. (2020). Self-targeting, zwitterionic micellar dispersants enhance antibiotic killing of infectious biofilms—An intravital imaging study in mice. *Sci. Adv.* 6, eabb1112. doi: 10.1126/sciadv.abb1112
- Torres, J. P., Senejani, A. G., Gaur, G., Oldakowski, M., Murali, K., and Sapi, E. (2020). Ex Vivo Murine Skin Model for *B. burgdorferi* Biofilm. *Antibiot. (Basel)* 9(9):528. doi: 10.3390/antibiotics9090528
- Valm, A. M., Mark Welch, J. L., and Borisy, G. G. (2012). CLASI-FISH: principles of combinatorial labeling and spectral imaging. *Syst. Appl. Microbiol.* 35, 496–502. doi: 10.1016/j.syapm.2012.03.004
- Valm, A. M. (2019). The Structure of Dental Plaque Microbial Communities in the Transition from Health to Dental Caries and Periodontal Disease. *J. Mol. Biol.* 431, 2957–2969. doi: 10.1016/j.jmb.2019.05.016
- Van Dyck, K., Van Dijck, P., and Vande Velde, G. (2020). Bioluminescence Imaging to Study Mature Biofilm Formation by *Candida* spp. and Antifungal Activity In Vitro and In Vivo. *Methods Mol. Biol.* 2081, 127–143. doi: 10.1007/978-1-4939-9940-8_9
- Whitchurch, C. B., Tolker-Nielsen, T., Ragas, P. C., and Mattick, J. S. (2002). Extracellular DNA required for bacterial biofilm formation. *Science* 295, 1487. doi: 10.1126/science.295.5559.1487
- Wolcott, R., Costerton, J. W., Raoult, D., and Cutler, S. J. (2013). The polymicrobial nature of biofilm infection. *Clin. Microbiol. Infect.* 19, 107–112. doi: 10.1111/j.1469-0691.2012.04001.x
- Yang, C. Y., Bialecka-Fornal, M., Weatherwax, C., Larkin, J. W., Prindle, A., Liu, J., et al. (2020). Encoding Membrane-Potential-Based Memory within a Microbial Community. *Cell Syst.* 10, 417–423.e413. doi: 10.1016/j.cels.2020.04.002
- Yuan, L., De Haan, P., Peterson, B. W., De Jong, E. D., Verpoorte, E., Van Der Mei, H. C., et al. (2020). Visualization of Bacterial Colonization and Cellular Layers in a Gut-on-a-Chip System Using Optical Coherence Tomography. *Microsc. Microanal.* 1–9. doi: 10.1017/S143192762002454X
- Zobell, C. E., and Allen, E. C. (1935). The Significance of Marine Bacteria in the Fouling of Submerged Surfaces. *J. Bacteriol.* 29, 239–251. doi: 10.1128/JB.29.3.239-251.1935

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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