

Real-time colorimetric hydration sensor for sport activities

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1 **REAL-TIME COLORIMETRIC SENSOR OF DEHYDRATION AND**
2 **OVER-HYDRATION FOR SPORT ACTIVITIES**

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4 **Yubin Zhou^a, Htet Han^a, Hnin Pwint Phyuson Naw^a, Alice Venecia Lammy^a, Chee**
5 **Hong Goh^a, Souhir Boujday^{b,c,*}, Terry W.J. Steele^{a,*}**

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7 *a School of Materials Science & Engineering, College of Engineering, Nanyang*
8 *Technological University, 50 Nanyang Avenue, Singapore 639798, Singapore*

9 *b Sorbonne Universit s, UPMC, Univ Paris 6, UMR CNRS 7197, Laboratoire de R activit *
10 *de Surface, F75005 Paris, France*

11 *c CNRS, UMR 7197, Laboratoire de R activit de Surface, F75005 Paris, France*

12 ** Corresponding Author*

13 *Souhir Boujday: souhir.boujday@upmc.fr, Terry W.J. Steele: wjsteele@ntu.edu.sg,*

14 *Tel.:65-65927594, fax: 65-67909081.*

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18 **Abstract**

19 Dehydration and over-hydration are a serious health concern in endurance sport activities,
20 especially in tropical climates. Current methods for detection are inconvenient and not widely
21 applied. Point-of-care hydration monitoring during a sport event is an ideal preventative
22 therapy, as it allows personalize real-time feedback towards preventing hydration related
23 injuries.

24 Gold nanoparticles allow colorimetric detection, whose color changes due to changes in
25 colloidal stability. The red-shift in the plasmonic band induces a color change from red to
26 purple upon aggregation that can be observed by the naked eye, which gives promising

27 applications in point-of-care preventative therapies. We developed a gold nanoparticle-based
28 colorimetric sensor for rapid detection of dehydration and over-hydration by monitoring
29 'proof-of-concept' artificial sweat ion concentrations. The colorimetric sensor was designed
30 to distinguish between dehydration and over-hydration through easily observed color changes.
31 In addition, the sensor exhibited a fast response time within minutes. The designed sensor is a
32 promising tool for point-of-care and real-time detection of dehydration and over-hydration.

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34 *Keywords:* gold nanoparticles, colorimetric detection, **sensor**, sport activity, dehydration,
35 over-hydration

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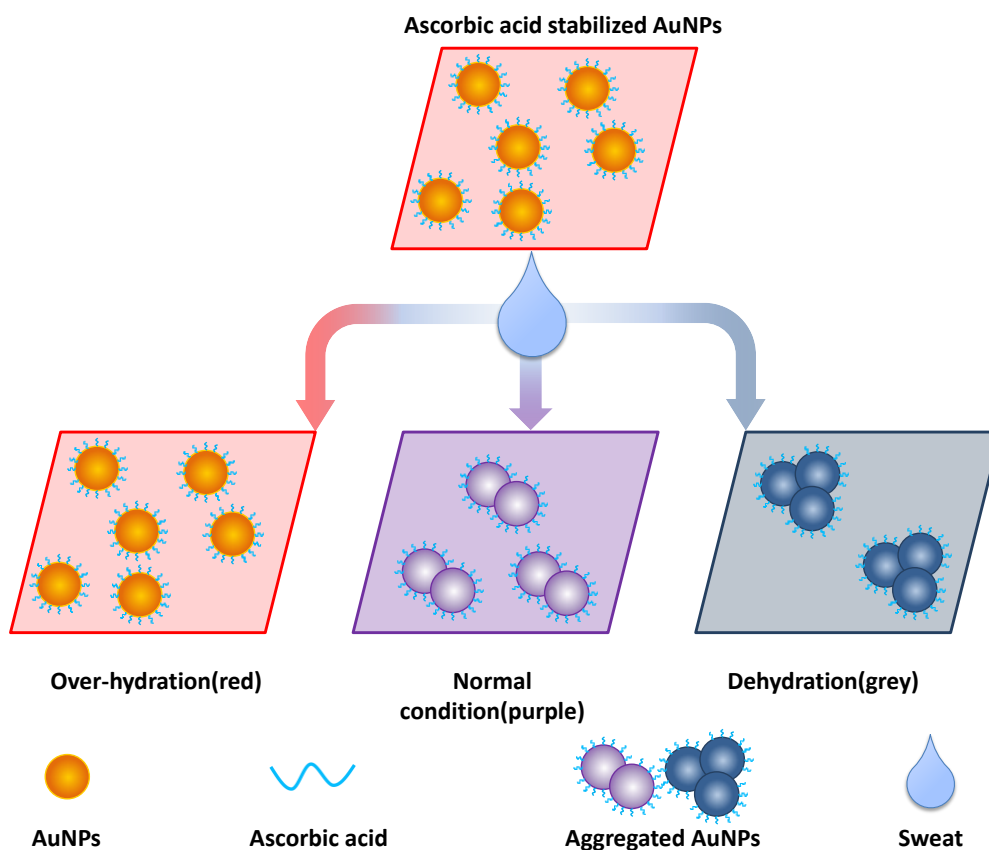
37 **1. Introduction**

38 Endurance sport activities are often associated with fatigue, muscle cramps, and hydration
39 imbalance. Among which dehydration is the most common due to the water loss in the form
40 of sweat to keep the body cool during intensive sport activities. The human sweat contains not
41 only water but also other mineral salts, lactic acid, glucose and urea [1]. Depending on the
42 rate of rehydration of the individuals, hydration imbalances will be developed leading to two
43 extreme conditions called dehydration and over-hydration [2]. Detection of hydration status
44 requires complicated procedures to analyze body fluids such as sweat, blood, urine and saliva
45 [3, 4] by analyzing body mass changes, urinary and hematological indices and salivary
46 parameters [5]. Therefore, a non-invasive and easy-to-use technique is still in need to give
47 real-time feedback during continuous strenuous exercise. Hydration status imbalance can be
48 represented by the variation of electrolyte concentration in the sweat. Electrolytes in the sweat
49 contain salts and dissolved ions such as sodium cations, which form the majority of the
50 available ions other than potassium and calcium [1]. Hence, sweat sodium cations were

51 chosen to determine the hydration status with normal physiological sweat concentration at 40
52 mM, over-hydration at 26.5 mM and dehydration at 47.9 mM [6, 7].

53 Gold nanoparticles (AuNPs) have been widely studied and applied in many sensing
54 application of inorganic ions [8] and biological analytes such as amino acids, peptides,
55 proteins, and DNA fragments [9-13]. The optical properties of AuNPs enable their use for
56 colorimetric transduction of molecular interactions simply by standard UV-Vis spectrometer
57 or even by the naked eye [14-15]. Indeed, the small size of these particles compared to that of
58 the visible wavelength gives rise to a strong extinction band in the visible wavelength regime
59 upon collective electron oscillations [16]. This band is the localized surface plasmon
60 resonance (LSPR) and its shape and position are closely dependent from local changes on
61 particle surfaces and from their aggregation/dispersion state. It was reported that the addition
62 of NaCl could induce the aggregation of the AuNPs, resulting in a color change because of
63 the alteration in LSPR [17]. We use this property here to monitor the hydration state in sweat
64 through the detection of standard values of salt concentration. The logic-gate based sensor
65 was developed by tailoring the formulation of AuNPs. The designed model is able to give
66 resolution in the concentration range of human body fluid. Figure 1 displays the working
67 principal of the sensor. This design is applied to detect 26.5, 40 and 47.9 mM of NaCl in
68 artificial sweat that correspond to over hydration, normal hydration and dehydration,
69 respectively. These values are based on the data of average population, but the designed
70 sensor can be customized for other ranges. The objective is displaying three distinguished
71 colors for these three concentration levels within few minutes, which can be discerned by the
72 naked eye. To the best of our knowledge, it is the first time applying AuNPs to sweat sensing
73 for hydration imbalance detection. Therefore, this user friendly and responsive sensor is
74 promising towards real time detection of ion concentrations in excreted body fluids.

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Figure 1. Schematic illustration of the colorimetric sensor.

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80 2. Experimental Work

81 All chemicals were purchased from Sigma-Aldrich Singapore. The procedure to synthesize

82 AuNPs by reduction of ascorbic acid was modified from the literature [18-20]. In all the

83 reactions, gold nanoparticle solutions were used at 0.35 g L^{-1} and the volume ratio of NaCl

84 solution or artificial sweat to AuNPs solution was 5:1 unless otherwise stated. Briefly, the

85 synthesis was started with 100 uL of 100 g L^{-1} chloroauric acid ($\text{HAuCl}_4 \cdot 3\text{H}_2\text{O}$), mixed with

86 25 mL of deionized water and 1.5 mL of 21.2 g L^{-1} sodium carbonate solution. The reaction

87 was started by adding 2 mL of 7 g L^{-1} ascorbic acid under vigorous stirring at 1000 rpm on

88 the magnetic stirrer for 15 minutes. The solution mixed was chilled with ice at about $0\text{-}2 \text{ }^\circ\text{C}$.

89 The concentration of the obtained AuNPs solution is 0.35 g L^{-1} . Shimadzu UV-visible

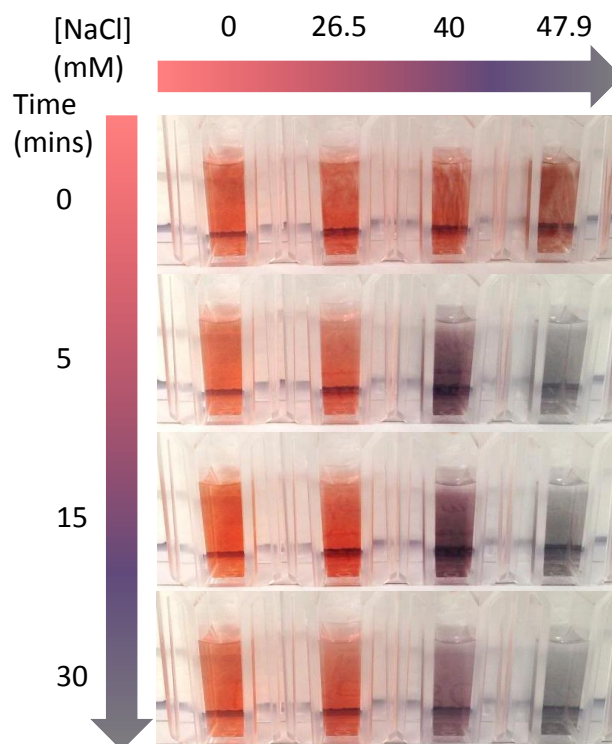
90 spectrophotometer was used to measure the absorption spectra of the AuNPs solutions
91 whereas Malvern Zeta Sizer, described previously [21], was used to measure the size and zeta
92 potential of the AuNPs. NaCl solutions in concentrations of 40, 26.5 and 47.9 mM were use
93 as indicators of normal physiological condition, over-hydration and dehydration. The final
94 optimized formulation was applied to artificial sweat of pH 6.7 according to EN1811 standard
95 [22].

96

97 3. Results & Discussion

98 **3.1 Colorimetric sensing by using Ascorbic Capped AuNPs:** Solutions with different
99 concentrations of NaCl (0, 26.5, 40 and 47.9 mM) were added to the red ascorbic-capped Au
100 nanoparticles. The observed color changes at different incubation times (0 to 30 min) are
101 shown in Figure 2.

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105 **Figure 2.** The different color of AuNPs solution in the presence of different concentrations of NaCl.

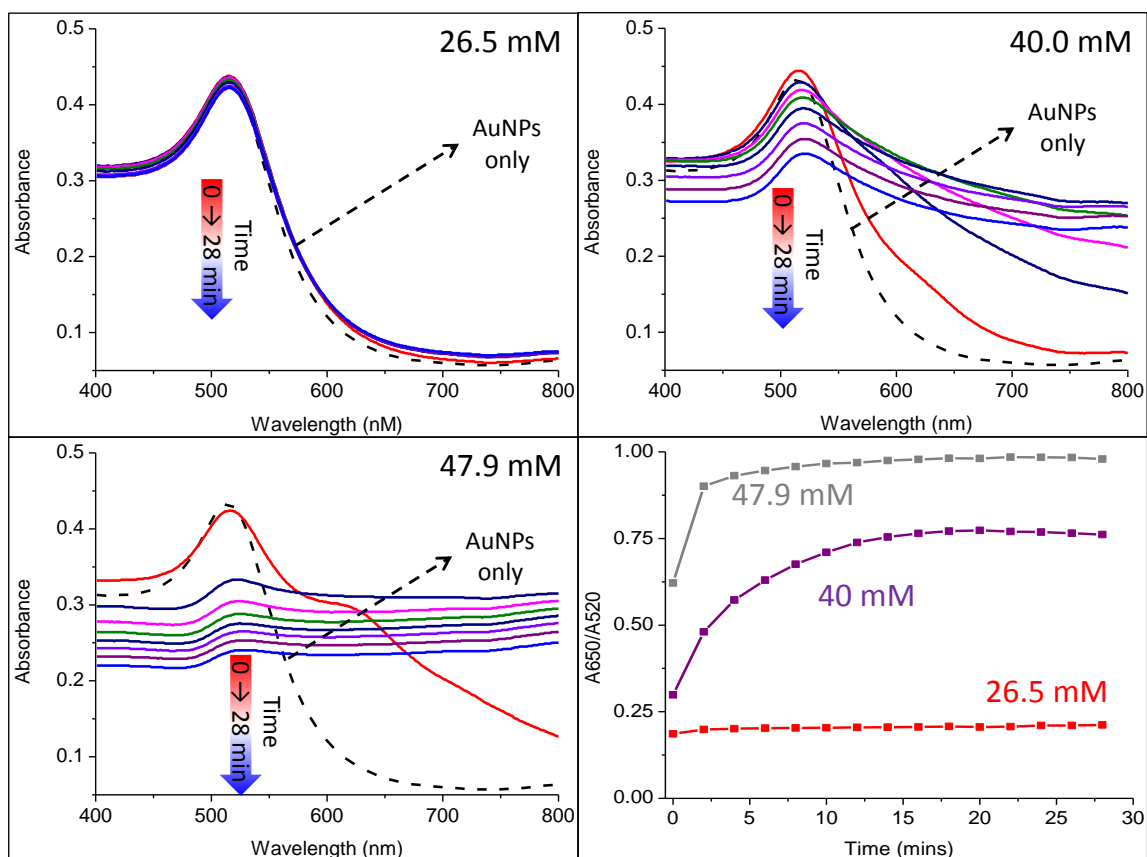
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107 Figure 2 shows that in absence of NaCl, the AuNPs solutions with deionized water (negative
108 control) undergo no color change during the experiment. The AuNPs solution with NaCl 26.5
109 mM also did not display any detectable color change. Starting from 40 mM NaCl, AuNPs
110 solutions changed color from red to purple within 5 minutes. Color was stable during the
111 following 30 min indicating that the detection is achieved at this early stage. The color of
112 AuNPs solution with NaCl 47.9 mM transformed rapidly red to grey color solution at 5
113 minutes, with no color change thereafter. The results show that different colors, easily
114 identified by the naked eye could be obtained rapidly at NaCl concentration of 26.5, 40 and
115 47.9 mM which indicates the sensor can distinguish the hydration imbalance situations.

116

117 **3.2 UV-Vis measurement:** The color change of the AuNPs solutions was verified by
118 measuring the absorption spectra in the visible range by the UV-Vis spectrometer. Figure 3
119 shows the UV spectra of AuNPs in the presence of NaCl at a concentration of 26.5, 40 and
120 47.9 mM respectively as a function of time.

121 For 26.5 mM NaCl UV-Vis spectra weren't modified compared to the original AuNPs
122 solutions. Starting from 40 mM NaCl in the mixture, the intensity of the Plasmon band at 520
123 nm decreases while a broad band in the around 650 nm appears indicating the aggregation of
124 gold nanoparticles.



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127 **Figure 3.** UV spectra of AuNPs in the presence of different concentration of NaCl as a function of time.

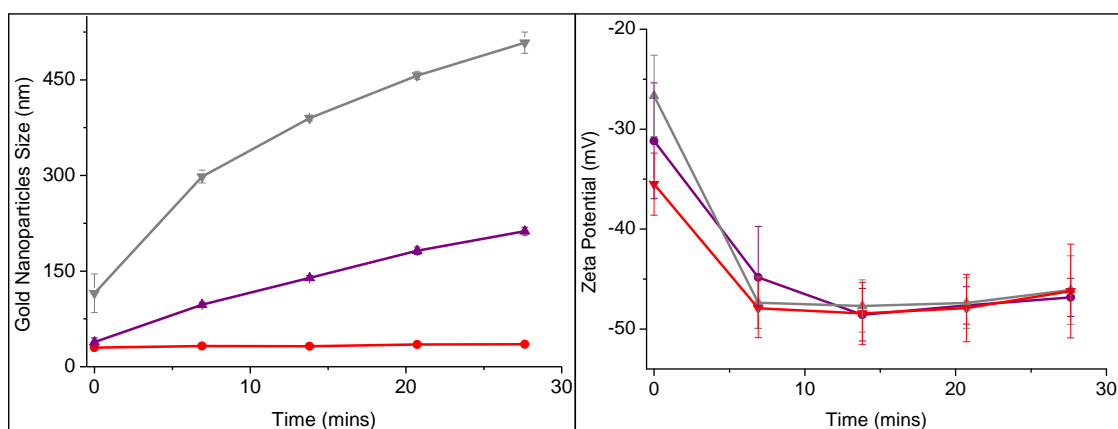
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129 The ratio of absorbance at 650 to 520 nm (A_{650}/A_{520}) is a good indicator of the aggregation
 130 state of gold nanoparticle [23], this ratio in the presence of different concentrations of NaCl at
 131 different times is also analyzed in Figure 3. As the concentrations of NaCl increased from
 132 26.5 to 40 and 47.9 mM, the A_{650}/A_{520} ratio increased indicating the decrease of dispersed
 133 particles and the increase of AuNPs aggregates. This aggregation induces the color change of
 134 the mixture. In addition, A_{650}/A_{520} ratio increases for NaCl concentration of 40 and 47.9
 135 mM, indicating a higher aggregation state for high salt concentration, consistently with the
 136 observed color changes. Finally, the change in A_{650}/A_{520} ratio takes place only during the
 137 first 5 to 10 minutes suggesting that the equilibrium is rapidly reached and the color change is
 138 stable during at least 30 min.

139

140 **3.3 Zeta Sizer Measurement:** The sizes of AuNPs in the experiments were measured with
141 Malvern Zeta Sizer. Figure 4 represents the change in size and zeta potential of the AuNPs
142 solutions in the presence of NaCl solution as time proceeded. Upon the addition of the NaCl
143 solution, the zeta potential of the AuNPs solution was unusually modified. The kinetic
144 followed the previous observation by naked eye and UV-Vis, i.e. a rapid change and a
145 stabilization after 10 min.

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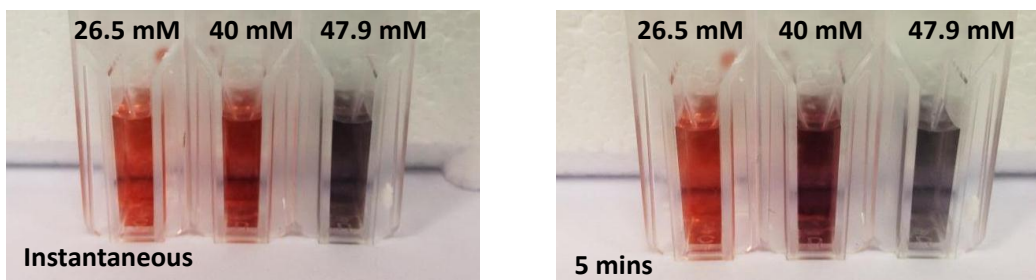
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149 **Figure 4.** Size and Zeta Potential change of AuNPs as a function of time in the presence of different
150 concentrations of NaCl: 26.5 mM (red) 40 mM (purple), 47.9 mM (grey).

151 The hydrodynamic radius of the AuNPs prior to NaCl addition was less than 30 nm. For the
152 lowest NaCl concentration, 26.5 mM, this size wasn't modified during the measurement time,
153 consistently with previously evidenced absence of aggregation. For higher salt concentrations,
154 the measured radius increased without reaching a plateau. This discrepancy may simply result
155 from the difficulty to measure accurately aggregates size. Nevertheless, the observed changes
156 are in agreement with UV-Vis observation and evidence the dependence of colloids
157 aggregation step on the tested range of salt concentration.

158

159 **3.4 Detection of Artificial Sweat:** The designed sensor was validated in artificial sweat
160 samples. Artificial sweat was synthesized according to EN1811 standard. Figure 5
161 corresponds to the distinguished colors obtained by AuNPs solutions with the addition of
162 EN1811 standard artificial sweat. The corresponding UV spectra are shown in the
163 Supplementary Information (Figure S1) while the standard deviations (%) of the 26.5 mM, 40
164 mM and 47.9 mM samples are 1.8 %, 3.0 % and 1.0 %, respectively. Three distinct colors are
165 obtained upon adding 26.5 mM, 40 mM and 47.9 mM of NaCl. We tested different
166 concentrations of AuNPs and changed the ratio AuNP/NaCl. The optimal performances were
167 obtained using 0.35 g L⁻¹ AuNP solution with 5:1 volume ratio of artificial sweat to AuNPs
168 solution. The results from visual experiment, UV-spectrum and Zeta-potential show the
169 functionality of this colorimetric sensor with the advantages of simplicity and accuracy. The
170 advantage of using ascorbic acid as the capping agent is that it is less stable compared to
171 citrate and other capping and thus optimization of tuning the size of AuNPs with NaCl
172 solution and artificial sweat is more sensitive to NaCl concentrations. Non-invasive methods
173 such as epidermal tattoo potentiometric sodium sensors [24] or the tattoo lactate sensors are
174 current available biosensors. Other sweat sensors such as wearable sweat rate sensors [25] are
175 again dependent of the electronic circuits [26]. Nevertheless, to the best of our knowledge,
176 there is still no colorimetric sensor that monitors sweat sodium concentration for sports
177 activities that applies color change method which is very convenient for the user. We are
178 currently conducting experiments on selection of substrates and deposition methods.
179 Adsorbent pads made from cellulose extraction thimbles, polycaprolactone, cellulose
180 membrane, hydrophilic filter paper and blotting pad will be tried while deposition methods
181 such freeze-drying and vacuum drying will be applied to develop our sensor into solid form.
182



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Figure 5. The different colors of AuNPs with artificial sweat.

186

187 **4. Conclusion**

188

189 In this study, a colorimetric sensor for the detection of hydration level in buffer and artificial
190 sweat was designed and developed. Detection of sodium cations at concentrations indicating
191 over-hydration, normal hydration and dehydration state was achieved using ascorbic acid
192 capped AuNPs. These particles aggregation/dispersion state occurred in the desired range of
193 salt. Colorimetric changes from red to purple to grey with increasing salt concentrations were
194 easily observable with UV-vis spectrometer and naked eyes. This sensor responds according
195 the actual human body conditions during over hydration and dehydration typical of endurance
196 events. Unlike the common assessment methods for sweat, this design can provide real time
197 feedback. It reduces all the intricate methods of monitoring hydration condition by a visual
198 indication of color change. This novel sensor has been shown to be promising for detection of
199 hydration levels with many desirable properties, including simple detection display, high
200 stability, and fast response time. Finally, this model of sensor may be developed to other
201 kinds of detections for small changes of biological substances in excreted fluids.

202

203 **Acknowledgement**

204

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209 Challenge.

210

211 **References**

212

- 213 [1] Harvey CJ, LeBouf RF, Stefaniak AB. Stefaniak. Formulation and stability of a novel artificial human
214 sweat under conditions of storage and use. *Toxicol In Vitro* 2010;24:1790-6.
- 215 [2] Sawka MN, et al. American College of Sports Medicine position stand. Exercise and fluid replacement.
216 *Med Sci Sports Exerc* 2007;39:377-90.
- 217 [3] Fiona E. Pelly GJS, Tanya M. King. Assessment of Hydration of Athletes. *Nutritional Assessment of*
218 *Athletes, Second Edition* 2010:341-74.
- 219 [4] Kerkick C. Fluid Needs of Athletes. *Nutritional Guidelines for Athletic Performance* 2012:175-94.
- 220 [5] Armstrong LE. Assessing Hydration Status: The Elusive Gold Standard. *J Am Coll Nutr* 2007;26:575S-84S.
- 221 [6] Taylor N, Machado-Moreira C. Regional variations in transepidermal water loss, eccrine sweat gland
222 density, sweat secretion rates and electrolyte composition in resting and exercising humans. *Extrem Physiol*
223 *Med* 2013;2:1-30.
- 224 [7] Toker J. The Math of salt loss.
225 http://www.slowtwitch.com/Training/General_Physiology/The_Math_of_salt_loss_1093.html. 2009.
- 226 [8] Lin Z, et al. Visual detection of copper(ii) based on the aggregation of gold nano-particles via click
227 chemistry. *Anal Methods* 2012;4:612-5.
- 228 [9] Ngo YH, et al. Gold nanoparticles paper as a SERS bio-diagnostic platform. *J Colloid Interf Sci*
229 2013;409:59-65.
- 230 [10] Lin Y-W, Liu C-W, Chang H-T. DNA functionalized gold nanoparticles for bioanalysis. *Anal Methods*
231 2009;1:14-24.
- 232 [11] Elghanian R, et al. Selective Colorimetric Detection of Polynucleotides Based on the Distance-Dependent
233 Optical Properties of Gold Nanoparticles. *Science* 1997;277:1078-81.

- 234 [12] Morel A-L, Volmant R-M, M éhivier C, Krafft J-M, Boujday S, Pradier C-M. Optimized immobilization
235 of gold nanoparticles on planar surfaces through alkyldithiols and their use to build 3D biosensors. *Colloids
236 and Surfaces B: Biointerfaces*. 2010;81:304-12.
- 237 [13] Morel A-L, Boujday S, M éhivier C, Krafft J-M, Pradier C-M. Biosensors elaborated on gold nanoparticles,
238 a PM-IRRAS characterisation of the IgG binding efficiency. *Talanta*. 2011;85:35-42.
- 239 [14] Link S, El-Sayed MA. Size and Temperature Dependence of the Plasmon Absorption of Colloidal Gold
240 Nanoparticles. *J Phys Chem B* 1999;103:4212-7.
- 241 [15] Liu X, et al. Biofunctionalized Gold Nanoparticles for Colorimetric Sensing of Botulinum Neurotoxin A
242 Light Chain. *Anal Chem* 2014;86:2345-52.
- 243 [16] Sardar R, et al. Gold Nanoparticles: Past, Present, and Future†. *Langmuir* 2009;25:13840-51.
- 244 [17] Wang G, Sun W. Optical Limiting of Gold Nanoparticle Aggregates Induced by Electrolytes. *The Journal
245 of Physical Chemistry B*. 2006;110:20901-5.
- 246 [18] Caillard L, et al. Gold Nanoparticles on Oxide-Free Silicon–Molecule Interface for Single Electron
247 Transport. *Langmuir* 2013;29:5066-73.
- 248 [19] Tabrizi A, Ayhan F, Ayhan H. Gold nanoparticle synthesis and characterisation. *Hacettepe J Biol & Chem*
249 2009;37:217-26.
- 250 [20] Salcedo ARM, Sevilla III FB. Citrate-Capped Gold Nanoparticles as Colorimetric Reagent for Copper (II)
251 Ions. *Philipp Sci Lett* 2013; 6: 90-96.
- 252 [21] Steele TWJ, et al. Factors influencing polycation/siRNA colloidal stability toward aerosol lung delivery.
253 *Eur J Pharm Biopharm* 2012;80:14-24.
- 254 [22] Midander K, et al. Nickel release from nickel particles in artificial sweat. *Contact Dermatitis* 2007;56:325-
255 30.
- 256 [23] He L, et al. Colorimetric Sensing of Tetracyclines in Milk Based on the Assembly of Cationic Conjugated
257 Polymer-Aggregated Gold Nanoparticles. *Food Anal Methods* 2013;6:1704-11.
- 258 [24] Bandothkar AJ, et al. Epidermal tattoo potentiometric sodium sensors with wireless signal transduction for
259 continuous non-invasive sweat monitoring. *Biosens Bioelectron* 2014;54:603-9.
- 260 [25] Salvo P, et al. A Wearable Sensor for Measuring Sweat Rate. *IEEE Sens J* 2010;10:1557-8.
- 261 [26] Pietro S. Sweat Rate Wearable Sensors. *Biological and Medical Sensor Technologies* 2012:243-262.

262 **Figure captions:**

263 **Figure 1.** Schematic illustration of the colorimetric sensor.

264 **Figure 2.** The different color of AuNPs solution in the presence of different concentrations of NaCl.

265 **Figure 3.** UV spectra of AuNPs in the presence of different concentration of NaCl as a function of
266 time.

267 **Figure 4.** Size and Zeta Potential change of AuNPs as a function of time in the presence of different
268 concentrations of NaCl: 26.5 mM (red) 40 mM (purple), 47.9 mM (grey).

269 **Figure 5.** The different colors of AuNPs with artificial sweat.

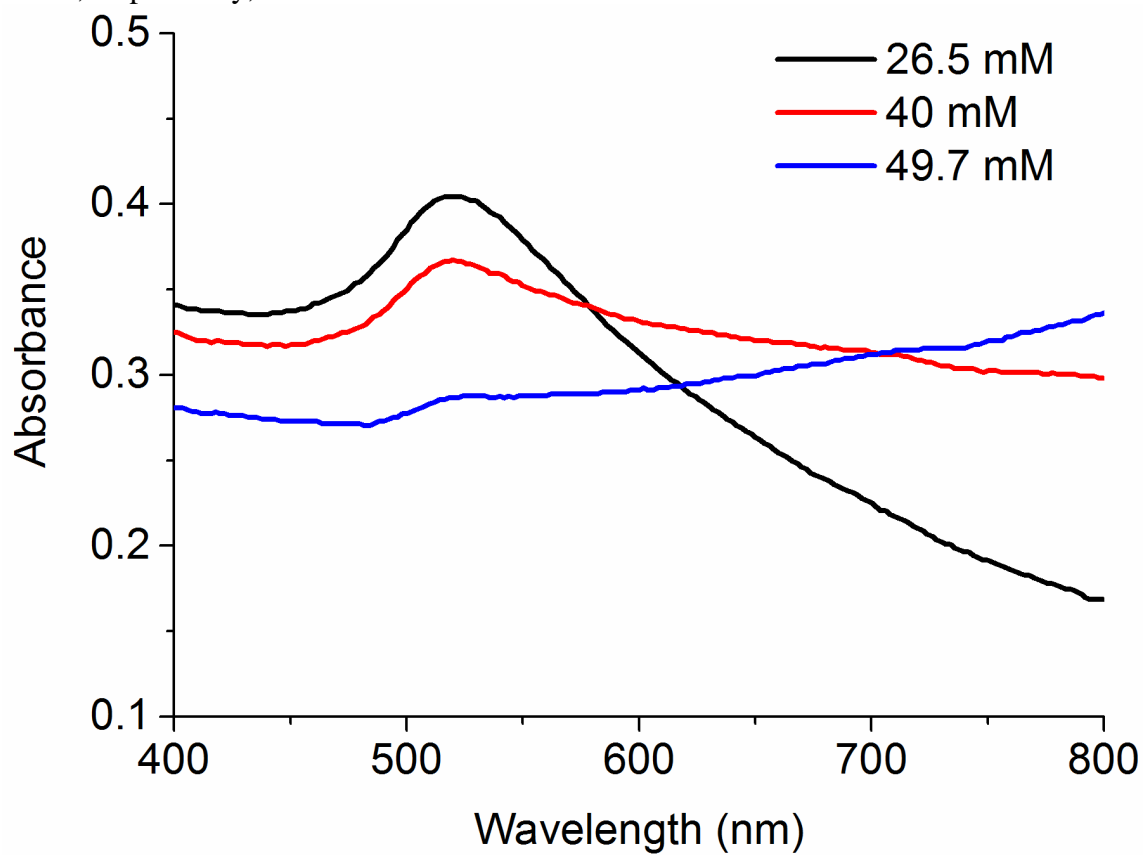
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274 **Supplementary Figure S1 UV spectra of AuNPs in the presence of artificial sweat.**
275 Standard deviations (%) of the 26.5 mM, 40 mM and 47.9 mM samples are 1.8 %, 3.0 % and
276 1.0 %, respectively, n=3.



277