

mcr-3 and mcr-4 Variants in Carbapenemase-Producing Clinical Enterobacteriaceae Do Not Confer Phenotypic Polymyxin Resistance

Teo, Jeanette W. P.; Kalisvar, Marimuthu; Venkatachalam, Indumathi; Ng, Oon Tek; Lin, Raymond T. P.; Octavia, Sophie

2018

Teo, J. W. P., Kalisvar, M., Venkatachalam, I., Ng, O. T., Lin, R. T. P., & Octavia, S. (2018). mcr-3 and mcr-4 Variants in Carbapenemase-Producing Clinical Enterobacteriaceae Do Not Confer Phenotypic Polymyxin Resistance. *Journal of Clinical Microbiology*, 56(3), e01562-17-.

<https://hdl.handle.net/10356/88578>

<https://doi.org/10.1128/JCM.01562-17>

© 2018 American Society for Microbiology. This paper was published in *Journal of Clinical Microbiology* and is made available as an electronic reprint (preprint) with permission of American Society for Microbiology. The published version is available at: [<http://dx.doi.org/10.1128/JCM.01562-17>]. One print or electronic copy may be made for personal use only. Systematic or multiple reproduction, distribution to multiple locations via electronic or other means, duplication of any material in this paper for a fee or for commercial purposes, or modification of the content of the paper is prohibited and is subject to penalties under law.



mcr-3 and *mcr-4* Variants in Carbapenemase-Producing Clinical *Enterobacteriaceae* Do Not Confer Phenotypic Polymyxin Resistance

Jeanette W. P. Teo,^a Marimuthu Kalisvar,^{b,c,d} Indumathi Venkatachalam,^e Oon Tek Ng,^{b,c,f} Raymond T. P. Lin,^{a,g} Sophie Octavia^g

^aNational University Hospital, Department of Laboratory Medicine, Singapore

^bTan Tock Seng Hospital, Department of Infectious Diseases, Singapore

^cNational Centre for Infectious Diseases, Singapore

^dNational University of Singapore, Yong Loo Lin School of Medicine, Singapore

^eSingapore General Hospital, Department of Infectious Diseases, Singapore

^fNanyang Technological University, Lee Kong Chian School of Medicine, Singapore

^gNational Public Health Laboratory, Ministry of Health, Singapore

KEYWORDS bioinformatics, *Enterobacteriaceae*, next-generation sequencing, PCR, antibiotic resistance, plasmid, polymyxin

The worldwide distribution of plasmid-mediated colistin resistance determinants (*mcr-1*, *mcr-2*, *mcr-3*, and *mcr-4*) coupled to the emerging observation that colistin resistance is more prevalent in carbapenem-resistant *Enterobacteriaceae* (CRE) (1, 2) presents a daunting challenge in combatting antimicrobial resistance. Undoubtedly, next-generation sequencing approaches have expedited the discovery of mobile colistin resistance determinants (3). In this study, we undertook the *in silico* screening of 500 phenotypically carbapenem-resistant carbapenemase-producing *Enterobacteriaceae* whole genomes for the presence of the *mcr* gene, using CLC Genomics Workbench (CLC Bio-Qiagen, Aarhus, Denmark). The isolates comprised clinical and screening pure cultures submitted to the national reference laboratory for mandatory CRE surveillance. Locally, the presence of *mcr-1* as well as its cocarriage with KPC-2 had been previously well described (4, 5); hence, we did not look further into the distribution of *mcr-1*. *mcr-2* was not detected among the genomes analyzed. *mcr-3* was identified in one *Escherichia coli* genome (ENT1955) by the use of both read mapping and *de novo* assembly. To date, several *mcr-3* variants have been observed (6, 7). The *mcr-3* gene identified in this study shared 99.94% nucleotide identity with the first *mcr-3* gene discovered by Yin et al. (8) (GenBank accession no. [KY924928.1](https://doi.org/10.1093/nar/kn249)) due to a “C” deletion at nucleotide position 218; the deletion was confirmed by Sanger sequencing. This resulted in a truncated protein of only 88 amino acids (Table 1) that was deemed to be nonfunctional (10). This *mcr-3*-like gene has been deposited in GenBank (see below). The genome of ENT1955 was *de novo* assembled using all the reads, and the draft genome was annotated. No plasmid-related genes were found in the approximately 9-kb contig containing the *mcr-3*-like gene. We also performed read mapping using *E. coli* Y5 (GenBank accession no. [CP013483.1](https://doi.org/10.1093/nar/kn249)) as the reference because it was the most closely related genome available in GenBank. Reads that did not match the *E. coli* Y5 chromosome were separately assembled. Using this approach, the derived *mcr-3*-like contig was almost identical to the one obtained from *de novo* assembly using all reads and, again, no plasmid-related genes were observed. This led us to believe that, in contrast to those reported in several previous studies (8, 11, 12), our *mcr-3*-like gene was not plasmid associated. Interestingly, downstream of the *mcr-3*-like gene, a gene encoding IS2

Accepted manuscript posted online 13
December 2017

Citation Teo JWP, Kalisvar M, Venkatachalam I, Ng OT, Lin RTP, Octavia S. 2018. *mcr-3* and *mcr-4* variants in carbapenemase-producing clinical *Enterobacteriaceae* do not confer phenotypic polymyxin resistance. *J Clin Microbiol* 56: e01562-17. <https://doi.org/10.1128/JCM.01562-17>.

Editor Nathan A. Ledebauer, Medical College of Wisconsin

Copyright © 2018 American Society for Microbiology. All Rights Reserved.

Address correspondence to Jeanette W. P. Teo, Jeanette_Teo@nuhs.edu.sg.

TABLE 1 Characteristics of KPC-2 carbapenemase-producing *Enterobacteriaceae* cocarrying *mcr* variants

Isolate name	Species	Source ^a	MIC (mg/liter) ^b		Date of isolation	Multilocus sequence type	MCR mutation(s)	Other acquired resistance determinants ^d	Replicon of <i>mcr</i> plasmid	Reference <i>mcr</i> plasmid (GenBank accession no.)
			Colistin	Polymyxin B						
<i>mcr</i> -3-like isolate ENT1955	<i>E. coli</i>	Rectal swab	0.25	0.25	4 April 2015	167	MCR-3-like truncated at amino acid position 88	<i>bla</i> _{KPC-2} , <i>bla</i> _{TEM-1B} , <i>aadA1</i> , <i>erm</i> (B), <i>mph</i> (A), <i>floR</i> , <i>sul3</i> , <i>tet</i> (A)		
<i>mcr</i> -4.2 isolates ENT1164	<i>E. cloacae</i>	Rectal swab	32 ^c	16 ^c	11 May 2014	54	V179G, V236F in comparison to MCR-4	<i>bla</i> _{KPC-2} , <i>bla</i> _{TEM-1A} , <i>bla</i> _{TEM-1B} , <i>aac</i> (6') <i>lb-cr</i> , <i>aacA4</i> , <i>fosA</i> , <i>mph</i> (A), <i>catB8</i> , <i>catA1</i> , <i>sul1</i>	ColE10	<i>Salmonella</i> sp. plasmid pMCR_R3445 (MF543359)
ENT1344	<i>E. cloacae</i>	Rectal swab	16 ^c	16 ^c	28 July 2014	54	V179G, V236F in comparison to MCR-4	<i>bla</i> _{KPC-2} , <i>bla</i> _{TEM-1A} , <i>bla</i> _{TEM-1B} , <i>bla</i> _{SHV-12} , <i>aac</i> (6') <i>lb-cr</i> , <i>aacA4</i> , <i>fosA</i> , <i>mph</i> (A), <i>catA1</i> , <i>catB8</i> , <i>sul1</i> , <i>QnrS1</i>	ColE10	<i>Salmonella</i> sp. plasmid pMCR_R3445 (MF543359)
ENT1606	<i>E. cloacae</i>	Rectal swab	16 ^c	16 ^c	24 October 2014	54	V179G, V236F in comparison to MCR-4	<i>bla</i> _{KPC-2} , <i>bla</i> _{TEM-1A} , <i>aac</i> (6') <i>lb-cr</i> , <i>aacA4</i> , <i>fosA</i> , <i>mph</i> (A), <i>catA1</i> , <i>catB8</i> , <i>sul1</i>	ColE10	<i>Salmonella</i> sp. plasmid pMCR_R3445 (MF543359)
ENT1017	<i>E. cloacae</i>	Rectal swab	0.25	0.25	2 February 2014	54	V179G, V236F in comparison to MCR-4	<i>bla</i> _{KPC-2} , <i>bla</i> _{TEM-1B} , <i>bla</i> _{SHV-12} , <i>fosA</i> , <i>mph</i> (A), <i>catA1</i> , <i>qnrS1</i>	ColE10	<i>Salmonella</i> sp. plasmid pMCR_R3445 (MF543359)
ENT1018	<i>E. cloacae</i>	Rectal swab	0.125	0.125	2 February 2014	54	V179G, V236F in comparison to MCR-4	<i>bla</i> _{KPC-2} , <i>bla</i> _{TEM-1B} , <i>bla</i> _{SHV-12} , <i>fosA</i> , <i>mph</i> (A), <i>catA1</i> , <i>qnrS1</i>	ColE10	<i>Salmonella</i> sp. plasmid pMCR_R3445 (MF543359)
ENT1019	<i>E. cloacae</i>	Rectal swab	0.25	0.25	2 February 2014	54	V179G, V236F in comparison to MCR-4	<i>bla</i> _{KPC-2} , <i>bla</i> _{TEM-1B} , <i>bla</i> _{SHV-12} , <i>fosA</i> , <i>mph</i> (A), <i>catA1</i> , <i>qnrS1</i>	ColE10	<i>Salmonella</i> sp. plasmid pMCR_R3445 (MF543359)

^aIsolates were from a routine surveillance study suggesting colonization.

^bEuropean Committee on Antimicrobial Susceptibility Testing (EUCAST) breakpoints were used for interpretation of colistin MIC results (susceptible, ≤2 mg/liter; resistant, >2 mg/liter).

^cA single "skip well" was observed during testing, which suggests heteroresistance to polymyxins in these isolates (9).

^dDetermined by ResFinder 3.0 (<https://cge.cbs.dtu.dk/services/ResFinder/>).

TABLE 2 Broth microdilution polymyxin susceptibility testing of cloned *mcr-3*-like and *mcr-4.2* expressed in *E. coli* BL21(DE3) host

<i>E. coli</i> BL21(DE3) strain ^a	MIC (mg/liter)	
	Colistin	Polymyxin B
pET48b— <i>mcr-3</i> -like	1	1
pET48b— <i>mcr-4.2</i>	1	1
pET48b	1	0.5
No vector	0.5	1

^aExpression of *mcr* was induced by the addition of 1 mM isopropyl β -d-1-thiogalactopyranoside (IPTG) to cation-adjusted Mueller-Hinton broth media.

transposase TnpB, matching the gene encoding the transposase carried by *mcr-3*-bearing plasmid pWJ1 (8), was detected. Phenotypically, the isolate was susceptible to both colistin and polymyxin B (Table 1) as determined by the reference broth microdilution method (13).

There were no exact matches to *mcr-4* in the screened genomes. Instead, the *mcr-4*-like gene was found in six *Enterobacter cloacae* isolates (Table 1), which had 100% nucleotide identity to a putative sulfatase from *Shewanella frigidimarina*. Compared to *mcr-4* (GenBank accession no. [MF543359](#)), this *mcr-4*-like gene carried 2 missense mutations resulting in codon changes at positions V179G and V236F. The *mcr-4*-like gene was located on a 7.7-kb contig, and a BLAST search showed 95% coverage and 99% similarity to the *mcr-4* gene in plasmid pMCR_R3445 (GenBank accession no. [MF543359](#)). Thus, we named this *mcr-4*-like gene "*mcr-4.2*" (see below).

Interestingly, the genomes of all of the *mcr-4.2*-positive isolates also possessed another putative phosphoethanolamine transferase (EPT) (GenBank accession no. [WP_012477388](#)). This putative EPT was not encoded on the plasmid. An alignment of the MCR-4.2 amino acid sequence and the putative EPT against MCR-1 and MCR-2 showed conservation in active site residues (data not shown) (10). However, it appears that the presence of *mcr-4.2* and the predicted novel EPT genes did not confer overt phenotypic resistance to polymyxins (Table 1). The finding of heterogeneously elevated drug MICs for *E. cloacae* isolates is likely attributable to heteroresistance causing a "skip well" phenomenon observed during the broth microdilution susceptibility testing (Table 1) (9).

Expression vector pET-48b(+) (Novagen, WI, USA) was used for functional cloning of the full coding sequences of the *mcr-3*-like and *mcr-4.2* genes but without the fusion tags. There was essentially no difference in the polymyxin MICs for the cloned *mcr-3*-like and *mcr-4.2* genes, which suggests that they did not confer phenotypic resistance (Table 2). Therefore, we concluded that the *mcr-3*-like gene carrying a nonsense mutation was nonfunctional and that *mcr-4.2* alone was unlikely to be the major mechanism of resistance to polymyxins.

In summary, we describe the discovery of new *mcr*-like elements, although phenotypic susceptibility testing indicates that the presence of these genes alone was unlikely to contribute to colistin resistance.

Accession number(s). The *mcr-3*-like and *mcr-4.2* genes identified in this study have been deposited in GenBank under accession no. [MG026622](#) and [MG026621](#), respectively.

ACKNOWLEDGMENTS

We thank the National Public Health Laboratory (NPHL), Singapore, for providing carbapenemase-producing *Enterobacteriaceae* isolates and, in particular, Felicia Ong for performing the carbapenemase screening assays and Siti Zulaina for preparing plate cultures. We thank Bernadette Cheng for performing the expression cloning. We also thank the Carbapenemase-producing *Enterobacteriaceae* in Singapore (CaPES) study group. CaPES study group members are Asok Kurup, Benjamin Cherng, Choong Weng Lam, Deepak Rama Narayana, De Partha Pratim, Hsu Li Yang, Indumathi Venkatachalam, Jeanette Teo, Kalisvar Marimuthu, Koh Tse Hsien,

Micelle Ang, Nancy Tee, Nares Smitasin, Ng Oon Tek, Ooi Say Tat, Prabha Unny Krishnan, Raymond Fong, Raymond Lin Tzer Pin, Surinder Kaur Pada, Tan Thean Yen, and Thoon Koh Cheng.

REFERENCES

- Giani T, Antonelli A, Caltagirone M, Mauri C, Nicchi J, Arena F, Nucleo E, Bracco S, Pantosti A; AMCLI-CoSA survey participants, Luzzaro F, Pagani L, Rossolini GM. 2017. Evolving beta-lactamase epidemiology in Enterobacteriaceae from Italian nationwide surveillance, October 2013: KPC-carbapenemase spreading among outpatients. *Euro Surveill* 22:30583. <https://doi.org/10.2807/1560-7917.ES.2017.22.31.30583>.
- Huang TD, Bogaerts P, Berhin C, Hoebeke M, Bauraing C, Glupczynski Y. 2017. Increasing proportion of carbapenemase-producing Enterobacteriaceae and emergence of a MCR-1 producer through a multicentric study among hospital-based and private laboratories in Belgium from September to November. *Euro Surveill* 22:30530. <https://doi.org/10.2807/1560-7917.ES.2017.22.19.30530>.
- Kluytmans J. 2017. Plasmid-encoded colistin resistance: mcr-one, two, three and counting. *Euro Surveill* 22:30588. <https://doi.org/10.2807/1560-7917.ES.2017.22.31.30588>.
- Teo JQM, Ong RTH, Xia E, Koh TH, Khor CC, Lee SJY, Lim TP, Kwa AL. 2016. mcr-1 in multidrug-resistant blaKPC-2-producing clinical Enterobacteriaceae isolates in Singapore. *Antimicrob Agents Chemother* 60:6435–6437. <https://doi.org/10.1128/AAC.00804-16>.
- Teo JW, Chew KL, Lin RT. 2016. Transmissible colistin resistance encoded by mcr-1 detected in clinical Enterobacteriaceae isolates in Singapore. *Emerg Microbes Infect* 5:e87. <https://doi.org/10.1038/emi.2016.85>.
- Roer L, Hansen F, Stegger M, Sönksen UW, Hasman H, Hammerum AM. 2017. Novel mcr-3 variant, encoding mobile colistin resistance, in an ST131 *Escherichia coli* isolate from bloodstream infection, Denmark, 2014. *Euro Surveill* 22:22846. <https://doi.org/10.2807/1560-7917.ES.2017.22.31.30584>.
- Ling Z, Yin W, Li H, Zhang Q, Wang X, Wang Z, Ke Y, Wang Y, Shen J. 24 October 2017. Chromosome-mediated mcr-3 variants in *Aeromonas veronii* from chicken meat. *Antimicrob Agents Chemother* <https://doi.org/10.1128/AAC.01272-17>.
- Yin W, Li H, Shen Y, Liu Z, Wang S, Shen Z, Zhang R, Walsh TR, Shen J, Wang Y. 2017. Novel plasmid-mediated colistin resistance gene mcr-3 in *Escherichia coli*. *mBio* 8:e00543-17. <https://doi.org/10.1128/mBio.00543-17>.
- Landman D, Salamera J, Quale J. 2013. Irreproducible and uninterpretable polymyxin B MICs for *Enterobacter cloacae* and *Enterobacter aerogenes*. *J Clin Microbiol* 51:4106–4111. <https://doi.org/10.1128/JCM.02129-13>.
- Hu M, Guo J, Cheng Q, Yang Z, Chan EWC, Chen S, Hao Q. 2016. Crystal structure of *Escherichia coli* originated MCR-1, a phosphoethanolamine transferase for colistin resistance. *Sci Rep* 6:38793. <https://doi.org/10.1038/srep38793>.
- Hernández M, Iglesias MR, Rodríguez-Lázaro D, Gallardo A, Quijada N, Miguela-Villoldo P, Campos MJ, Piriz S, López-Orozco G, de Frutos C, Sáez JL, Ugarte-Ruiz M, Domínguez L, Quesada A. 2017. Co-occurrence of colistin-resistance genes mcr-1 and mcr-3 among multidrug-resistant *Escherichia coli* isolated from cattle, Spain, September 2015. *Euro Surveill* 22:30586. <https://doi.org/10.2807/1560-7917.ES.2017.22.31.30586>.
- Litrup E, Kiil K, Hammerum AM, Roer L, Nielsen EM, Torpdahl M. 2017. Plasmid-borne colistin resistance gene mcr-3 in *Salmonella* isolates from human infections, Denmark, 2009–17. *Euro Surveill* 22:30587. <https://doi.org/10.2807/1560-7917.ES.2017.22.31.30587>.
- Clinical and Laboratory Standards Institute. 2015. M07-A10. Methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically; approved standard: ninth edition. Clinical and Laboratory Standards Institute, Wayne, PA.