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Human natural killer cells control *Plasmodium falciparum* infection by eliminating infected red blood cells

Qingfeng Chen, Anburaj Amaladoss, Weijian Ye, Min Liu, Sara Dummler, Fang Kong, Lan Hiong Wong, Hooi Linn Loo, Eva Loh, Shu Qi Tan, Thiam Chye Tan, Kenneth T. E. Chang, Ming Dao, Subra Suresh, Peter R. Preiser, and Jianzhu Chen

Immune deficiency mouse–human chimeras provide a powerful approach to study host-specific pathogens, such as *Plasmodium falciparum* that causes human malaria. Supplementation of immunodeficient mice with human RBCs supports infection by human *Plasmodium* parasites, but these mice lack the human immune system. By combining human RBC supplementation and humanized mice that are optimized for human immune cell reconstitution, we have developed RBC-supplemented, immune cell-optimized humanized (RICH) mice that support multiple cycles of *P. falciparum* infection. Depletion of human natural killer (NK) cells, but not macrophages, in RICH mice results in a significant increase in parasitemia. Further studies in vitro show that NK cells preferentially interact with infected RBCs (iRBCs), resulting in the activation of NK cells and the elimination of iRBCs in a contact-dependent manner. We show that the adhesion molecule lymphocyte-associated antigen 1 is required for NK cell interaction with and elimination of iRBCs. Development of RICH mice and validation of *P. falciparum* infection should facilitate the dissection of human immune responses to malaria parasite infection and the evaluation of therapeutics and vaccines.

Malaria is caused by infection with parasites of the *Plasmodium* species which are transmitted by bites of infected Anopheles mosquitoes. *Plasmodium* species are highly host specific, making it difficult to model human parasite infection in laboratory animals. So far, most in vivo experimental studies of malaria have been carried out with mouse and rat *Plasmodium* strains in rodents. Differences in invasion and disease pathology between human and rodent parasite species have impeded the translation of findings from rodents into humans. The lack of appropriate small animal models also has impeded the evaluation of new drugs and vaccines before clinical trials (1).

To overcome this challenge, one approach is to supplement SCID mice with human RBCs. The resulting mice support a limited blood-stage *P. falciparum* infection (2–4). The need to inject large volumes of human RBCs repeatedly and to treat mice with anti-neutrophil antibody and highly toxic clodronate liposomes to suppress the rapid clearance of the injected human RBCs by macrophages in the recipient mice makes working with this system difficult. More recently NOD-SCID IL2rg−/− (NSG) mice have been shown to support a more efficient *P. falciparum* infection without the treatment of clodronate liposomes or anti-neutrophil antibody (5). Furthermore, a recent report shows the development of liver-stage *P. falciparum* infection in immunocompromised and fumarylacetoacetate hydrolase-deficient (*Fah−/−, Rgs2−/−, I2rg−/−*) (FRG) mice. Backcrossing of FRG mice to the NOD background and supplementing the resulting mice with human RBCs led to reproducible transition from liver-stage infection to blood-stage infection (6). Despite such progress, none of the existing mouse models of human parasite infection has a human immune system.

The immune system plays a critical role in the control of parasite infection. Studies in mice using mouse *Plasmodium* strains have shown that mouse immune cells such as natural killer (NK) cells, T cells, dendritic cells, and B cells all contribute to anti-parasitic immunity (7–10). Notably, depletion of NK cells in a mouse model of *Plasmodium chabaudi* infection results in more severe disease associated with higher parasitemia and mortality (11). In vitro, *P. falciparum*-infected human RBCs are shown to interact with human NK cells, leading to the induction of IFN-γ (12). Compared with *P. falciparum* schizonts, live infected RBCs (iRBCs) induce more rapid activation and more production of IFN-γ by NK cells (13). More recently, it has been shown that, in addition to IFN-γ, activated human NK cells also produce perforin and granzyme against *P. falciparum*-infected RBCs (14). However, because of the lack of appropriate models, little is known about the role of human NK cells in the control of *P. falciparum* infection in vivo. NK cells are cytolytic and can lyse virus-infected cells and tumor cells (15). However, whether...
NK cells also can eliminate parasite-infected RBCs directly has not been demonstrated comprehensively.

In our study of humanized mice, we previously had developed a simple and effective method of enhancing human cell reconstitution by hydrodynamic expression of human cytokines. Expression of human IL-15 and Flt-3/Flik-2 ligand (Flt-3L) enhances the reconstitution of human NK cells, monocytes, and macrophages (16). In this study, we have constructed humanized mice that have an optimized human immune cell reconstitution as well as high levels of human RBCs through supplementation. We show that such humanized mice support an efficient infection by *P. falciparum*. Depletion of human NK cells, but not macrophages, in these mice results in a significant increase in parasitemia. Our additional studies in vitro show that NK cells interact preferentially with iRBCs and become activated, resulting in the elimination of iRBCs in a contact-dependent manner. We further show that the cell adhesion molecules lymphocyte-associated antigen 1 (LFA-1) and to some extent DNAX accessory molecule 1 (DNAM-1) mediate NK cell interaction with and elimination of iRBCs. Development of humanized mice with robust reconstitution of human immune cells and human RBCs and validation of the model for *P. falciparum* infection should facilitate the dissection of human immune responses to malaria parasite infection and the evaluation of therapeutics and vaccines.

**Results**

**RBC-Supplemented, Immune Cell-Optimized Humanized Mice Support Robust *P. falciparum* Infection.** To construct humanized mice that have an optimized human immune cell reconstitution as well as high levels of human RBCs, we expressed human IL-15 and Flt-3L in humanized mice to enhance the reconstitution of human NK cells, monocytes, and macrophages (16) and supplemented the cytokine-treated mice with human RBCs. Specifically, humanized mice with 40% or more human leukocyte reconstitution in peripheral blood mononuclear cells (PBMCs) were injected with plasmids encoding human IL-15 and Flt3L. One day after plasmid injection, mice were given a daily injection of 1 mg of human RBCs. After supplementing RBC for 7 d, mice were bled and analyzed for human RBC and human immune cell reconstitution by flow cytometry. A significant fraction (25.1 ± 9.8%, n = 10) of RBCs stained positive for human CD235ab (Fig. 1A), and a large fraction (42.1 ± 11.7%, n = 10) of PBMCs stained positive for human CD45 (Fig. 1B). Among various human immune cell types, significant levels of CD56+ NK cells (14.2 ± 3.6%), CD14+ monocytes/macrophages (4.5 ± 1.3%), CD3+ T cells (33.2 ± 12.2%), and CD19+ B cells (51.6 ± 15.4%) were present (Fig. 1C). Among NK cells, both CD56+CD16− and CD56+CD16+ populations were detected, and very few cells were positive for CD69 (Fig. S1). However, NK cells were activated by IL-15 in vitro, suggesting that NK cells in the blood of cytokine-treated mice were not activated. These results show that RBC supplementation into cytokine-treated humanized mice leads to high levels of both human RBC and immune cell reconstitution. We refer to these RBC-supplemented, immune cell-optimized humanized mice as “RICH” mice.

To determine the efficiency of parasite infection, RICH mice were injected i.v. with 5 × 109 synchronized ring-stage parasites from the 3D7 strain of *P. falciparum* 7 d after RBC supplementation. The infected mice were given human RBC supplementation daily throughout the experiment. Blood samples were taken every 48 h to measure parasitemia by microscopy. At all time points, significant levels of iRBCs were detected, with the majority of the parasites being at the ring stage. Over a 240-h period, blood parasitemia increased steadily from 0.01 ± 0.001% at 48 h to 0.13 ± 0.02% at 240 h (Fig. 1D; n = 10). (The parasitemia level would be four times the whole-blood value when normalized to human RBC frequency in the whole blood). These results show that RICH mice can support a significant level of *P. falciparum* infection in the presence of an optimized human immune system and provide an in vivo system for evaluating the role of specific human immune cell types in controlling parasite infection.

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**Fig. 1.** RICH mice support a robust *P. falciparum* infection. Humanized mice were injected hydrodynamically with plasmids encoding human IL-15 (50 μg) and Flt3L (10 μg) on day zero. Beginning 1 d after plasmid injection, mice were injected i.p. daily with 1 ml human RBCs. After 7 d of RBC supplementation, mice were infected i.v. with 5 × 106 ring-stage 3D7 parasites and given daily RBC supplementation throughout the experiment (n = 10). (A−C) Human RBC and immune cell reconstitution before parasite infection (7 d after plasmid injection). Shown are representative CD235ab vs. forward scatter (FSC) profiles of whole blood (A), human CD45 (hCD45) vs. mouse CD45.1 (mCD45) staining profiles of PBMCs (B), and human CD56 vs. CD14 and CD3 staining profiles gating on human CD45+ cells (C). The numbers indicate the percentage of cells in the gated regions. The mean ± SEM (n = 10) are shown in Results. (D) Quantification of parasitemia in RICH mice at different time points. Data are shown as percentage (mean ± SEM) of iRBCs among total RBCs and are a compilation from two independent experiments with five mice in each experiment.

NK Cells Play a Critical Role in Controlling Parasite Infection.** We next investigated whether human immune cells respond to *P. falciparum* infection in RICH mice. To determine the role of human NK cells and macrophages, RICH mice were injected with anti-human CD56 and anti-human CD14 antibody to deplete NK cells and macrophages, respectively. As shown in Fig. 2A, the percentages of CD56+ NK cells (13.5 ± 2.1%, n = 6) and CD14+ monocytes/macrophages (4.3 ± 1.1%, n = 6) in PBMCs of RICH mice were not affected significantly by PBS injection. Twenty-four hours after a single injection of anti-human CD56 (clone HCD56), the level of NK cells was reduced by ~25-fold to ~0.4%, as revealed by staining with an anti-CD56 antibody (clone MEM188) that recognizes a different epitope on CD56. This result was confirmed further by staining for the NK cell marker Nkp46 (Fig. S2). Similarly, anti-human CD14 (clone M5E2) treatment reduced the level of monocytes/macrophages by ~40-fold to ~0.1%, as revealed by staining with an anti-CD14 antibody (clone HCD14) that recognizes a different epitope on CD14. This result was confirmed further by staining for CD11b and HLA-DR (Fig. S2). The depletion was sustained for at least 3 d. These mice were injected i.v. with 5 × 106 purified 3D7 ring-stage parasites 24 h after antibody treatment. Two days later, the level of parasitemia in the blood was quantified by microscopy. In the PBS-treated mice, parasitemia was 0.01 ± 0.002% (n = 6), the same as in untreated RICH mice (compare Figs. 2B and 1D). In contrast, parasitemia increased about sevenfold to 0.074 ± 0.02% (n = 6) in NK cell-depleted mice. Depletion of human monocytes/macrophages did not lead to any significant change in parasitemia. These results show that human NK cells, but not macrophages, play a significant role in the immediate control of *P. falciparum* infection in RICH mice.

We measured the level of human IFN-γ and IL-6 in the sera of infected mice 2 d after infection. In PBS-treated RICH mice, the levels of IFN-γ and IL-6 were 189.1 ± 20.2 pg/mL and 130.2 ± 13.6 pg/mL (n = 6), respectively (Fig. 2C and D). In NK cell-depleted RICH mice, the levels of IFN-γ and IL-6 were reduced.
Human NK Cells Respond to and Eliminate iRBCs in Vitro. To investigate how human NK cells control parasite infection, we established an in vitro culture system using purified NK cells, RBCs, and parasites. Human NK cells were purified from adult PBMCs and cultured with human RBCs and 3D7 schizonts at a ratio of 1:25:1. Video microscopy revealed that NK cells migrated and interacted with RBCs randomly. However, when NK cells encountered iRBCs, they formed stable contacts as soon as 2 h after incubation (Fig. S3). Quantification showed that, on average, NK cell contact was 11-fold longer with iRBCs than with uninfected RBCs (4,075 ± 6), indicating preferential interaction between NK cells and iRBCs. Correspondingly, NK cells were activated to express CD69 in cultures in the presence of parasites as early as 4 h but were not activated to express CD69 in cultures in the absence of parasites (Fig. 3B). The fraction of NK cells that were induced to express CD69 reached ~24% after 24 h of culture, suggesting that NK cells are activated by iRBCs in vitro. Further analysis revealed that, although the levels of Nkp46, CD2, CD8, NKGD2, and NKGD2a tended to be higher on CD69+ CD69- cells than on CD69-CD69- NK cells, the difference was not significant.

We quantified parasitemia in the culture at 48 and 96 h by microscopy. The difference in parasitemia in cultures with (1.3 ± 0.2%) and without (0.9 ± 0.1%) NK cells already was significant (~twofold) at 48 h, and this difference became even more dramatic (6.0 ± 0.2% vs. 1.6 ± 0.1%, ~3.5-fold) at 96 h. These results were confirmed further by flow cytometry. Cultured cells were stained with the DNA-staining dye Hoechst and CD56 at 48 and 96 h. The iRBCs were positive for Hoechst but negative for CD56, but NK cells were positive for both (Fig. 3C). After culture for 48 h, 1.8 ± 0.1% of RBCs stained positive for Hoechst in the absence of NK cells, indicating infection by parasites. In the presence of NK cells, the percentage was reduced to 0.8 ± 0.1% (twofold). This reduction was more dramatic at 96 h (6.1 ± 0.2% vs. 1.6 ± 0.1%, ~3.8-fold), as was consistent with data from microscopy. Human NK cells purified from humanized mice also exhibited similar antiparasite effects in culture (Fig. 3D).

To gain direct evidence of NK cell killing of iRBCs, we visualized NK cell-iRBC coculture by video microscopy. After 12 h of culture, iRBCs that formed stable contact with NK cells became flattened, indicating leakage of cell content and loss of cell volume, whereas iRBCs without NK cell contact did not (Fig. S4). Furthermore, we tested whether inhibition of perforin activity by concanamycin A (CMA) interferes with elimination of iRBCs by NK cells. As shown in Fig. 3E, in the absence of NK cells, the percentage of iRBCs was ~8%. As expected, this percentage was reduced (to ~4%) in the presence of NK cells. The percentage was elevated (to ~7%) when CMA was added into
We further determined the requirement for cell-cell contact in the presence or absence of human NK cells in the same well or in a separate compartment in the transwell. NK cells were assayed for CD69 expression at 4, 12, and 24 h, and parasitemia was measured at 96 h by flow cytometry. (A) Comparison of CD69 expression by CD69 + NK cells in the same well or in a separate compartment in the transwell at the indicated time points of culture. The numbers indicate percentages of CD69 + cells.

**Fig. 4.** Cell-cell contact is required for NK cell activation and killing of iRBCs. (A and B) Human RBCs were infected with purified 3D7 schizonts in the absence or presence of human NK cells in the same well or in a separate compartment in the transwell. NK cells were assayed for CD69 expression at 4, 12, and 24 h, and parasitemia was measured at 96 h by flow cytometry. (A) Comparison of CD69 expression by CD69 + NK cells in the same well or in a separate compartment in the transwell at the indicated time points of culture. The numbers indicate percentages of CD69 + cells. (B) Representative CD56 vs. Hoechst staining profiles of cultured cells at 96 h. (C) Transwell assay of the killing of iRBCs by NK cells. NK cells were cultured with RBCs and 3D7 schizonts in one chamber (w/o NK). The other chamber contained only RBCs and parasites (w/o NK). Parasitemia in both chambers was quantified by flow cytometry at 48 and 96 h. Shown are representative CD56 vs. Hoechst-staining profiles of cells from the two chambers. The numbers in B and C indicate the percentage of iRBCs among total RBCs.

In contrast, the percentages of iRBCs were ~6% and 4%, respectively, when LFA-1 and DNAM-1 were blocked, suggesting a complete or partial blocking of iRBC killing by NK cells.

To gain insight into how LFA-1 is involved in the elimination of iRBCs by NK cells, we investigated the effect of an LFA-1-blocking antibody on the interaction of NK cells with iRBCs and the activation of NK cells. The length of time that NK cells interacted with iRBCs was reduced from an average of 4,236 ± 718 s in the absence of the LFA-1-blocking antibody to 428 ± 109 s in the presence of the blocking antibody (Fig. 5B). The latter time was virtually the same as the length of time that NK cells interacted with noninfected RBCs (Fig. 3A). Furthermore, in the presence of the LFA-1-blocking antibody, only 13% of NK cells expressed CD69 (Fig. 5C), a fourfold reduction compared with the expression of CD69 in the absence of the LFA-1-blocking antibody (52%). These results suggest that LFA-1 mediates the elimination of iRBCs by NK cells by promoting the interaction NK cell with iRBCs and the activation of NK cells.

**Discussion**

By supplementing immunodeficient mice with human RBCs, recent studies (2–6) have established various mouse models that support infection by human *Plasmodium* parasites. However,
a common deficiency of these models is the lack of a human immune system. A mouse model that has a human immune system and that supports infection by human Plasmodium parasites is highly desirable, because it would facilitate both dissection of human immune responses to parasite infection and the evaluation of the efficacy of vaccines and therapeutics that require the presence of a human immune system. Toward this goal, we supplemented humanized mice that are optimized for human immune cell reconstitution with human RBCs. Humanized mice were injected with IL-15– and Flt3L-expressing plasmids to enhance human immune cell reconstitution and then were supplemented daily with human RBCs. One week after cytokine expression and RBC supplementation, human immune cell reconstitution reached the peak level, and the percentage of human RBCs reached ~25%. When these cell–RICH mice were infected with the 3D7 strain of Plasmodium falciparum, parasitemia increased steadily over time from 0.01% at 48 h postinfection to 0.13% at 240 h postinfection. In NSG mice with human RBC supplementation, parasitemia can reach as high as 14% at 240 h postinfection (5). In RICH mice, the parasitemia is 100-fold lower (~0.13%) at the same point in time but is closer to the levels found in most malaria patients (18). The difference between RBC-supplemented NSG mice and RICH mice is the presence of a human immune system in the latter mice, so the differences in parasitemia in the two mice model suggest that the human immune system has an important role in controlling parasite infection in RICH mice (see below). Because RBCs do not express major histocompatibility complex molecules (19), the supplemented human RBCs and the reconstituted human immune cells in humanized mice can be from different donors, thus making it easier to construct RICH mice for Plasmodium infection.

Establishment of the RICH mouse model that supports robust parasite infection in the presence of the human immune system enabled us to investigate the role of human NK cells and macrophages in the control of parasite infection. Depletion of both human and mouse macrophages yielded a much smaller fold increase in blood parasitemia at 48 h postinfection. Because humanized mice have only human and no mouse NK cells, this result suggests that human NK cells play a critical role in the immediate control of parasite infection. In humanized mice, both human and mouse macrophages are present. The lack of any significant effect following depletion of human macrophages could mean that human macrophages are not required for controlling the parasite load at an early stage of infection or that the effect of human macrophage depletion is masked by the presence of mouse macrophages. It is notable that depletion of either human NK cells or macrophages resulted in similar reductions of human IFN-γ and IL-6 in the sera, suggesting that these cytokines do not mediate the antiparasite effect of NK cells. This interpretation was supported further by in vitro studies showing that NK cells exert their function through direct contact with iRBCs and that neutralization of IFN-γ does not diminish the NK cell effect (Fig. 5). Studies suggest that IFN-γ is produced by NK cells in malaria patients (20) and during in vitro culture with iRBCs (15). However, the role of IFN-γ in the antimalaria response is poorly understood. Our results exclude a significant direct antimalarial effect of IFN-γ or any autocrine effect on NK cells. It remains possible, however, that IFN-γ may be important for the downstream activation of human adaptive immune responses. The RICH mice described here provide an in vivo platform for evaluating human adaptive responses, especially T cells, against malaria parasites.

We elucidated how NK cell control parasite infection by further in vitro studies. Although human NK cells from PBMCs have been shown to respond to iRBCs by producing the proinflammatory cytokines IFN-γ, IL-12, and IL-18 as well as the cytotoxic molecules perforin and granzyme (12–14, 21, 22), most previous in vitro studies relied on culturing iRBCs with total PBMCs, which are a mixture of various immune cell populations. In our study, we used purified human NK cells from PBMCs to obtain direct evidence that human NK cells are able to migrate, bind, and eliminate iRBCs in the absence of other immune cell types. We show that, although NK cells appear to scan RBCs randomly in the culture, upon encounter they interact with iRBCs much longer than with uninfected RBCs. The prolonged interaction leads to the activation of NK cells, as indicated by expression of CD69. Once NK cells are activated, they eliminate iRBCs in a contact-dependent manner. By video microscopy, we observed flattening of iRBCs after interaction with NK cells but did not observe any sign of phagocytosis of iRBCs by NK cells. The flattening of iRBCs likely results from the leakage of cell content and loss of cell volume—and hence the killing—of iRBCs. Supporting this interpretation, the inhibition of perforin by CMA significantly reduces the elimination of iRBCs, a result that is consistent with previous observations that perforin and granzymes are induced when NK cells interact with iRBCs (14). Although data from previous research and our present study suggest that NK cells inhibit malaria parasite infection by killing the iRBCs, it remains possible that other mechanisms also may be involved. Although further studies are required to elucidate the detailed mechanisms, our present findings show that both the activation of NK cells by iRBCs and then the elimination of iRBCs by NK cells are contact dependent.

We further evaluated the classical NK cell activation and adhesion receptors in interaction with and elimination of iRBCs. Blocking NKG2D, Nkp30, Nkp44, Nkp46, CD2, DNAM-1, and 2B4 did not diminish the killing of iRBCs by NK cells, suggesting that these receptors are not involved in the recognition or killing of iRBCs by NK cells. We found that anti–LFA-1 completely and anti–DNAM-1 partially inhibited the killing of iRBCs by NK cells. These results are consistent with the observation that DNAM-1 is physically and functionally associated with LFA-1 (23). The observed difference between blocking LFA-1 and DNAM-1 suggests that LFA-1 plays a more dominant role. Besides its function in cell adhesion, LFA-1 also is known to contribute to NK cell cytotoxicity (24). We found that blocking of LFA-1 almost completely abolished the preferential interaction of NK cells with iRBCs and significantly blocked the activation of NK cells, suggesting that LFA-1 is required for the interaction between NK cells and iRBCs. The known ligand of LFA-1 is ICAM-4, which is expressed by RBCs (25). Because NK cells preferentially interact with iRBCs rather than with uninfected RBCs, NK cells may recognize other molecules, including parasite proteins, on iRBCs. Elucidation of the molecular nature of these interactions is critical to understand the underlying mechanism of the NK cell response to P. falciparum-infected RBCs.

By supporting a robust parasite infection in the presence of human immune system, RICH mice likely will find wide application in dissecting human immune responses to parasite infection in a physiological setting. It has been demonstrated that a human immune system and hepatocytes can be reconstituted in the same humanized mice (26). Similarly, we have shown previously that engraftment of CD34+ cells from fetter liver into NSG mice leads to reconstitution of human immune cells as well as human hepatocytes in the mouse liver (27). By supplementing human RBCs in such humanized mice, it may be possible to extend this model for both the blood and liver stage of Plasmodium infection in the presence of a human immune system.

Materials and Methods

Mice. NSG mice were purchased from the Jackson Laboratories and maintained under specific pathogen-free conditions in the animal facilities at National University of Singapore (NUS). Humanized mice were constructed as follows: Newborn pups (within 48 h of birth) were irradiated with 100 cGy using a gamma radiation source and were injected intracardially with CD34+ cells from fetal liver (2 × 10^6 cells per recipient). Mice were analyzed for human leukocyte reconstitution at age 10–12 wk by staining for human CD45 and mouse CD45. To make RICH mice, mice with 40% or more human leukocytes in circulation were injected intravenously with plasmids encoding human IL-15 and Flt3L. Beginning 1 d after plasmid injection, mice were i.p. injected daily with human RBC (resuspended in RPMI medium containing 50% [vol/vol] heat-inactivated human serum [Invitrogen]).
Seven days later, the levels of human RBCs in humanized mice were quantified by staining peripheral blood cells with anti-human CD253ab antibody (BioLegend). Mice with a minimum of 20% human RBC reconstitution were used for subsequent experimentation. To deplete human NK cells and macrophages, anti-human CD56 (clone HCD56; BioLegend) and anti-human CD14 (clone M5E2; BioLegend) antibodies were injected i.v. into RICH mice (50 μg per mouse) 24 h before parasite infection. All studies involving mice were approved by the Institutional Animal Care and Use Committee of NUS and MIT.

**P. falciparum Culture and Infection.** *P. falciparum* strain 3D7 was obtained from Malaria Research and Reference Reagent Resource and maintained in leukocyte-free human erythrocytes in malaria culture medium (CMC) (10.43 g RPMI 1640 powder, 25 mL 1 M Hepes buffer, 2 g NaHCO₃, 5 g Albumax, 25 mg gentamicin in 1 L milli-Q water) as described by Trager and Jensen (28). The parasites were treated with trypsin/chymotrypsin for 1 h at 37 °C with shaking before purification of schizonts to prevent the reinvasion of residual RBCs (29). The late-stage schizonts were purified by Percoll gradient centrifugation according to the protocol of Fernandez et al. (30).

For in vivo infection, synchronized ring-stage parasites were prepared by sorbitol synchronization. Briefly, 10–12 h postinvasion cultures were spun down at 600 × g for 5 min and were incubated in 5% (vol/vol) sorbitol for 10 min at room temperature. Cells were washed three times in MCM medium.

When parasitemia reached ~10%, the blood was harvested, treated with trypsin/chymotrypsin, and then injected i.v. into humanized mice. Every RICH mouse was injected with 5 × 10⁹ ring-stage parasites.

To set up in vitro NK cells, RBCs, and parasite coculture, human NK cells were purified from PBMCs using the EasySep Human CD56 Positive Selection Kit (Stem Cell Technologies). Human NK cells (with a purity of >95%) were mixed with purified late-stage schizonts at a ratio of 10:1, and then additional human RBCs were added to make a starting parasitemia of 0.2% (NK: schizonts/RBC = 10:1250). For transwell experiments, RBCs and parasites were seeded in the bottom wells of a 24-well plate (Millipore), and NK cells were seeded in the transwell at the same ratio. Cells were cultured at 37 °C, and samples were taken at various time points for flow cytometry analysis of NK cell activation and parasite infection of RBCs. Blocking antibodies against human NKp30 (P30-15), NKp44 (P44-8), NKp46 (9E2), NKG2D (1D11), CD2cid (RPA-2.10), NKp46 (944-8), NKp44 (962), NKpCD2 (1D11), CD2 (RPA-2.10), 2B4 (C1.7), DNAM-1 (11A8), CD11a (H111), and CD18 (T51/18) were purchased from BioLegend and used at a concentration of 1 μg/mL. CM A was purchased from Sigma and used in a culture at a final concentration of 1 nM.

**Antibodies, Flow Cytometry, and ELISA.** The following antibodies were used for flow cytometry: anti-human CD45 (2D1) from BD Biosciences and anti-human CD14 (HC4D14), CD56 (MEM-188), CD19 (HIB19), CD3 (HT3a), CD253ab (HIR2), CD69 (FN50), and anti-mouse CD45.1 (A20) from BioLegend. Cell suspensions of PBMCs and RBCs were stained with appropriate antibodies in 100 μL PBS containing 0.2% BSA and 0.05% sodium azide for 30 min on ice. Stained cells were analyzed by flow cytometry using LSR II, and data were analyzed by FACS Diva (BD Biosciences) or FlowJo (TreeStar Inc.). To measure human cytokines by ELISA, blood from humanized mice was centrifuged at 1,500 × g for 5 min at 4 °C. Serum was used to quantify human cytokines IFN-γ and IL-6 using ELISA kits (BioLegend) following the manufacturer’s protocol.

**Imaging.** NK cells (1 × 10⁷) were incubated with 1 × 10⁵ 3D7 parasite-infected RBCs (10% parasitemia, synchronized) in MCM medium on a culture slide made by mounting a polydimethylsiloxane chamber on a Menzel-Glaser 22 × 60 mm #1 glass coverslip. The culture slide was placed in a Tokai Hit INU Live Cell Microscope Chamber in which the temperature was maintained at 37 °C in an atmosphere of 5% CO₂ and 3% O₂. Live-cell images were acquired on an inverted Olympus IX71 fitted with an Olympus Planapo 60x/1.4 oil lens using a Hamamatsu ORCA-ER (C4742-80–12AG) CCD camera.

**Statistical Analysis.** Data are presented as mean ± SEM. Differences between groups were analyzed via Student t test. A P value <0.05 was considered statistically significant. All calculations were performed using the Origin 8.0 software package.

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