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<td>Author(s)</td>
<td>Goh, K. B.; Li, Hua; Lam, Khin Yong</td>
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Development of a multiphysics model to characterize the responsive behavior of urea-sensitive hydrogel as biosensor

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Abstract: A remarkable feature of biomaterials is their ability to deform in response to certain external bio-stimuli. Here, a novel biochemo-electro-mechanical model is developed for the numerical characterization of the urea-sensitive hydrogel in response to the external stimulus of urea. The urea sensitivity of the hydrogel is usually characterized by the states of ionization and denaturation of the immobilized urease, as such the model includes the effect of the fixed charge groups and temperature coupled with pH on the activity of the urease. Therefore, a novel rate of reaction equation is proposed to characterize the hydrolysis of urea that accounts for both the ionization and denaturation states of the urease subject to the environmental conditions. After examination with the published experimental data, it is thus confirmed that the model can characterize well the responsive behavior of the urea-sensitive hydrogel subject to the urea stimulus, including the distribution patterns of the electrical potential and pH of the hydrogel. The results point to an innovative means for generating electrical power via the enzyme-induced pH and electrical potential gradients, when the hydrogel comes in contact with the urea-rich solution, such as human urine.

Keywords: Multiphysics model; Urea-sensitive hydrogel; Urease; Bioelectrochemistry

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1. Introduction

According to Mayo clinic the chronic kidney disease (CKD) is characterized by the gradual loss of kidney functions over time (Ficalora, 2013). As the CKD progresses to the advanced-stage, a dangerous level of urea and electrolytes build-up in the human system, which increases the risk of cardiovascular, hyperlipidemia, anemia and metabolic bone diseases (Thomas et al., 2008). Therefore, the urea and electrolytes levels in the human blood and urine are measured for CKD patients during medical evaluation, since it provides access to the state of health of the kidney. As such, there were great attempt to develop the urea biosensors to estimate the blood urea nitrogen (BUN) levels for CKD patients by means of integrating urease into the hydrogel (Dezhong et al., 1996; Kovács et al., 2003; Liu et al., 1996). However, it is evident from the open literature that the performance of the hydrogel in response to the external stimulus of urea is still poorly understood. Therefore, in this paper, a multiphysics model is developed to investigate the responsive behavior of the hydrogel subject to the external stimulus of urea at different environmental conditions.

It is found in the open literature that the urea-sensitive hydrogel-based biosensors were synthesized using the pH- or temperature-sensitive hydrogels, i.e. poly(vinyl alcohol) (Jha et al., 2008; Marchenko et al., 2015), polyaniline (Buron et al., 2014), poly(o-phenylenediamine) (Chirizzi and Malitesta, 2011), poly(carbamoylsulfonate) (Eggenstein et al., 1999), and poly(3-cyclohexyl thiophene) (Pandey et al., 2000). Moreover, most researchers opted for gamma radiation (Hamdy et al., 2008), covalent binding (Chen and Chiu, 1999, 2000; Chen et al., 1994; Miyata et al., 1997), or physical entrapment (Hamilton and Breslin, 2014) for the amalgamation of the urease with the polymer network chains for the synthesis of the urea-sensitive hydrogel.
As such, it is reported that the integration of the urease with the polymer network chains improved the mechanical properties of the hydrogel and stability of the urease, which may then prolong the life shelf of the biosensors (Novick and Dordick, 1998; Teixeira et al., 2012).

As mentioned above, despite the extensive experimental investigations carried out on the urea-sensitive hydrogel, very few efforts were made via theoretical modeling and numerical simulation for the examination of the hydrogels. From the open literature, it is found that most of the published mathematical models were unable to capture the coupled biochemo-electro-mechanical behavior of the hydrogel. For example, the published mathematical models were inadequate in predicting the volumetric behavior of the hydrogels in response to the urea, where the simulated diffusion and reaction of the reactive solutes were associated with the urea hydrolysis and not the concomitant mechanical behavior of the hydrogels (Blaedel and Kissel, 1975; Chen and Chiu, 2000), including Moynihan et al. (1989) and Chowdhury and Bhattacharya (1997), where they modelled the kinetic parameters of the urea-sensitive hydrogel via the mass balance and Michaelis-Menten equations.

It is known that the mathematical correlation presented by Ogawa and Kokufuta may be the first work to attempt to predict the mechanical deformation of the hydrogels in response to urea (Ogawa and Kokufuta, 2002). The mathematical correlation was obtained from the experimental pH-responsive equilibrium swelling of the hydrogel, where the correlation was limited to the swelling at pH 4~7 immersed in 5 mM maleate-buffer solution of 35°C. Therefore, the correlation was unable to estimate the equilibrium swelling ratio of the hydrogel at different environmental conditions other than the prescribed conditions. It is also noted that the influence of environmental pH on the activity of the urease was accounted for in their reaction-diffusion
model, while the effect of pH coupled with temperature on the performance of the urease was excluded. Moreover, the effect of the fixed charge groups in the hydrogels was also neglected by all the published models mentioned above on the distribution pattern of the concentration of the reactive mobile ions. In order to overcome these limitations of the published mathematical models, a multi-effect-coupling urea-stimulus (MECurea) model is proposed in this paper to investigate the performance of the hydrogels in response to urea.

2. Formulation of the MECurea model

In order to formulate the model, a few components are prescribed delicately, given as follows: (a) the movement of mobile solutes between the hydrogel and surrounding solution; (b) the enzymatic hydrolysis reaction of urea within the hydrogel; (c) the interaction between the mobile ions and fixed charge in the hydrogel; (d) the swelling forces arising from the biochemical reactions and (f) the swelling behavior of the hydrogels, as illustrated in Fig. 1.

The present MECurea model is formulated based on the following assumptions:

(i) The hydrogel is maintained at the isothermal condition, such that the rate of reaction $r_k$ is constant.

(ii) The urease is immobilized and distributed uniformly within the hydrogel, where the activity of the urease is also constant.

(iii) The optimum temperature $T_{op}$ of urease is equal to the temperature at the midpoint of transition between active and inactive forms $T_{eq}$, such that $\Delta H_{eq} \approx 2\Delta G_{cat}$, where $\Delta H_{eq}$ is the enthalpy change associated with the conversion of an active to an inactive enzyme, and $\Delta G_{cat}$ is the activation energy of the catalytic reaction (Peterson et al., 2004).
(iv) The hydrogel has a macroporous nature such that the diffusivity coefficients $D_k$ of the mobile solutes are the same in both the hydrogels and surrounding solution.

(v) The solution flow is very slow and the convection term is neglectable, where the urea hydrolysis in the dilute buffer solution is also neglected.

### 2.1 Conservation of mass

In the MECurea model, at least six mobile ions and one molecule are considered in the present system, namely the hydrogen ion ($H^+$), hydroxide ion ($OH^-$), ammonium ion ($NH_4^+$), bicarbonate ion ($HCO_3^-$), cation and anion species, and urea. The Nernst-Planck equation is used to characterize the fluxes of the mobile solutes, given as follows (Li et al., 2009)

$$\nabla \cdot \left( C^{-1} D_k \left[ \nabla C_k + \frac{z_k F}{RT} C_k \nabla \Phi \right] \right) = v_k r_k \left( k = H^+, OH^-, ..., N \right)$$

where $C^{-1}$ the inverse of the right Cauchy-Green tensor and $C^{-1} = F^{-1} F^T$. $D_k$, $C_k$, $z_k$, $v_k$ and $r_k$ indicate the diffusivity tensor (m$^2$/s), concentration (mM), valence number, stoichiometric coefficient in the chemical reaction, and rate of chemical reaction for the $k$th mobile species. $R, T, F$, and $\Phi$ denote the universal gas constant (8.314 J/mol K), absolute temperature (K), Faraday constant (96487 C/mol), and electrostatic potential of the hydrogel. For the non-ionic solute where the charge $z_k = 0$, the Nernst-Planck Eq. (1) is reduced to Fick’s first law of diffusion.

### 2.2 Conservation of momentum

The bioelectrochemistry reaction within the hydrogel gives rise to the swelling pressure, resulting in the configurational changes or swelling deformation of the urea-sensitive hydrogel. It is known that the swelling of the hydrogel progresses, until the swelling forces are balanced by
the restrictive forces exerted by the elastic structure of the polymer network chains to achieve the equilibrium hydration state. In order to formulate this phenomenon, the law of conservation of linear momentum is employed, given as follows

$$\nabla \cdot \mathbf{P} = 0$$ (2)

where $\mathbf{P}$ is the Piola stress.

2.3 Free energy imbalance inequality for constitutive equation

In order to develop the equations of state for the hydrogel subject to the isothermal condition, the local free energy imbalance inequality is written as follows (Drozdov and deClaville Christiansen, 2015c)

$$\dot{W} - (u_1 + u_2 + u_3) \leq 0$$ (3)

where $\dot{W}$ is temporal increase of the Helmholtz free energy density of the hydrogel, while $u_1, u_2$ and $u_3$ denote the rate of work per volume expanded into the system by the mechanical, electric and chemical fields.

The free energy density of an idealized ionic hydrogel is defined as a sum of four components (Drozdov and deClaville Christiansen, 2015b): (i) the free energy density of stretching of the polymer network chains $W_1$, (ii) the free energy density of mixing the polymer and solvent $W_2$, (iii) the free energy density due to entropy mixing of mobile ions and solvent $W_3$, and (iv) the free energy density of the electric field formed by mobile ions and bound charges $W_4$

$$W = W_1 + W_2 + W_3 + W_4$$ (4)

The free energy density of elastic stretching of the polymer network chains is given as (Hong et al., 2008)
\[ W_i = \frac{1}{2} N k_B T \left[ \text{tr}(F^T F) - 2 \ln(\det F) - 3 \right] \]  

(5)

where \( N k_B T \) is the ground-state shear modulus \( G \), \( N \) is the number of polymer chains per volume of the dry polymer \((2.43 \times 10^{-23} \text{ m}^3)\), and \( k_B \) is the Boltzmann constant \((1.381 \times 10^{-23} \text{ J/K})\). Based on the theory of multiplicative decomposition of the deformation gradient (Chester, 2015), the right Cauchy-Green tensor is written as \( C = (\lambda_s)^2 C_e \), where \( \lambda_s \) is the swelling stretch and \( C_e \) is the elastic right Cauchy-Green tensor. As such, \( W_1 \) is recast to

\[ W_1 = \frac{1}{2} N k_B T \left[ (\lambda_s)^2 \text{tr} (C_e) - 2 \ln(\det F) - 3 \right] \]  

(6)

The free energy density of mixing between the polymer and solvent is taken as (Chester, 2015; Drozdov and deClaville Christiansen, 2015a)

\[ W_2 = R T C \left( \ln(C \nu) - \ln(1 + C \nu) + \frac{\chi}{1 + C \nu} \right) \]  

(7)

where \( C \) is the concentration of the solvent, \( \nu \) is the volume of a mole of fluid molecules and \( \chi \) is the Flory-Huggins interaction parameter.

The free energy density due to the entropy of mixing free energy of the solvent and mobile ions is given as (Doi, 2013)

\[ W_3 = R T \sum_k C_k (\ln C_k - \ln C - 1) \]  

(8)

The free energy density of the electric field formed by the mobile ions and fixed charge groups is given as follows (Hong et al., 2008)

\[ W_4 = \frac{1}{2 \varepsilon_r \varepsilon_0 J} (H \cdot C H) \]  

(9)
where \( J = (\lambda_s)^3 \), \( H \) is the dielectric displacement, and \( \varepsilon, \varepsilon_0 \) is the relative dielectric constant. It is understood that the volume change of the hydrogel is due to swelling and \( W_4 \) is rewritten as follows (Chester, 2015)

\[
W_4 = \frac{1}{2\varepsilon, \varepsilon_0 (1 + C \nu)^{1/3}} (\mathbf{H} \cdot \mathbf{C} \cdot \mathbf{H})
\]  

(10)

The mechanical power per volume is given by (Belytschko et al., 2013; Gurtin et al., 2010)

\[ u_1 = J \sigma : \mathbf{L} \]  

(11)

where \( \sigma \) is the symmetric Cauchy stress tensor and \( \mathbf{L} \) is the velocity \( u \), is given as (Chester, 2015; Gurtin et al., 2010)

\[ u_1 = (\mathbf{P} \mathbf{F}^T_s + J_p \mathbf{F}_s^{-T}) : \dot{\mathbf{F}}_e - \dot{\mathbf{J}}_s, \mathbf{p} \]  

(12)

where \( \mathbf{F}_s \) is the deformation gradient for the swelling of the hydrogel, while \( \mathbf{F}_e \) is the deformation gradient for the elastic deformation of the hydrogel.

The rate of work done by electric field per volume is written as follows (Drozdov and deClaville Christiansen, 2015c)

\[ u_2 = \mathbf{E} \cdot \mathbf{H} - F \Phi \sum_k z_k \dot{C}_k \]  

(13)

The last power term is produced due to the transport of solvent and mobile ions per volume given as (Nardinocchi et al., 2011)

\[ u_3 = - \left( \mu \nabla \cdot \mathbf{J} + \sum_k \mu_k \nabla \cdot \mathbf{J}_k \right) - \left( J \nabla \mu + \sum_k J_k \nabla \mu_k \right) \]  

(14)

where \( \mu \) and \( \mu_k \) are the chemical potentials of solvent and mobile ion \( k \) and \( \mathbf{J} \) and \( \mathbf{J}_k \) are the fluxes of solvent and mobile ion \( k \). The free energy imbalance inequality equation for characterizing the responsive behavior of the urea-sensitive hydrogels is given as follows

\[ \Delta F \geq \sum_k \beta_k \Delta \mu_k + \int \mathbf{u}_1 \cdot \mathbf{d} \mathbf{v} + \int \mathbf{u}_2 \cdot \mathbf{d} \mathbf{v} + \int \mathbf{u}_3 \cdot \mathbf{d} \mathbf{v} \]
\[
\frac{1}{(1 + C\nu)^{1/3}} \left( Nk_BT \left( 1 + C\nu \right) F_e + \frac{1}{\varepsilon} (F_e (H \otimes H)) - (1 + C\nu)^{4/3} \left( \sigma F_e^{-T} + p F_e^{-T} \right) \right) : \dot{F}_e
\]

\[
+ \dot{C} \left( \mu + Nk_BT \left[ \frac{\nu}{(1 + C\nu)^{1/3}} \left( \frac{1}{3} \text{tr}(C_e) - \frac{1}{(1 + C\nu)^{2/3}} \right) \right] + RT \left( \ln \frac{C\nu}{1 + C\nu} + \frac{1}{1 + C\nu^2} + \frac{\chi}{(1 + C\nu^2)^2} \right) \right)
\]

\[
- \dot{C} \left( - p\nu + RT \sum_k \left( \frac{C_k}{C} \right) + \frac{1}{6\varepsilon_0} \frac{\nu}{(1 + C\nu)^{4/3}} (H \cdot C_e \cdot H) \right) + \dot{C}_k \left( RT \ln \frac{C_k}{C} + z_k F\Phi - \mu_k \right)
\]

\[
+ \left( J \nabla \mu + \sum_k J_k \nabla \mu_k \right) \leq 0
\]

2.4 Biochemistry for constitutive equation

To implement Eq. (1), the classical rate of chemical reaction \( r_k \) that characterizes the reaction of urea in the present of urease is given as follows (Fogler, 2010)

\[
r_k = k_{cat} \frac{C_{E_{cat}} C_{\text{urea}}}{K_M}
\]

where \( C_{E_{cat}} \) and \( C_{\text{urea}} \) are the concentrations of the active enzyme and urea, respectively. \( k_{cat} \) is the constant catalytic rate of the enzyme and \( K_M \) is the Michaelis constant. It is found that Eq. (16) excludes the effect of urease denaturation. As such, in the present of denaturation of urease due to temperature, Eq. (16) is rewritten as (Peterson et al., 2007)

\[
r_k = V_{\text{max}} \frac{C_{\text{urea}}}{C_{\text{urea}} + K_M K_{EQ}}
\]

where \( V_{\text{max}} \) is the maximum velocity that can be achieved by a given quantity of an enzyme, and

\[
V_{\text{max}} = \frac{k_{cat} C_{E_{cat}}}{1 + K_{EQ}} \quad \text{(Menichetti et al., 2015)}
\]

\( K_{EQ} = \exp \left[ \frac{\Delta H_{eq}}{R} \left( \frac{1}{T_{eq}} - \frac{1}{T} \right) \right] \) is the equilibrium constant between active and inactive urease, and \( C_{E_{0}} \) is the initial concentration of the urease in the system.
The denaturation of urease due to pH is characterized by (Tijskens et al., 2001)

(a) $C_{E_{act}} + C_{OH^-} \Leftrightarrow C_{EOH_{inact}} \rightarrow CX$

(b) $C_{E_{act}} + C_{H^+} \Leftrightarrow C_{EH_{inact}} \rightarrow CX$

where $C_{EH_{inact}}$ and $C_{EOH_{inact}}$ are the concentration of the protonated and hydroxylated ureases, which are completely inactive but not denature, and $CX$ is the concentration of the denatured urease (Fidaleo and Lavecchia, 2003; Tijskens et al., 2001). $K_{EOH}$ and $K_{EH}$ are the equilibrium constant for the hydroxylation and protonation of the urease, and $K_w$ is the water dissociation constant. As such, the concentration difference of total and denatured enzymes is given as

$$C_{E_0} - CX = C_{E_{act}} \left(1 + \frac{K_w}{K_{EOH}C_{H^+}} + \frac{K_{H^+}}{K_{EH}} \right)$$

Finally, the rate of chemical reaction $r_k$ is further recast for the effect of temperature coupled with pH on the activity of the urease into the following

$$r_k = V_{max} \frac{C_{urea}}{K_m} \left( \frac{C_{urea}}{K_m} + K_{eq} \right) \left(1 + \frac{K_{EOH}C_{H^+}}{K_wC_{H^+}} + \frac{K_{EH}}{C_{H^+}} \right)$$

2.5 Electrical field for constitutive equation

In order to consider the coupled effect of the electrical potential $\Phi$ of the hydrogel and the movement of ionic solutes $k$, the Poisson equation is incorporated into the MECurea model by (Li et al., 2009)

$$\nabla \cdot (\varepsilon \varepsilon_0 J C^{-1} \nabla \Phi) = -F \left( \sum_k z_k C_k + z_f C_f \right)$$
where fixed charge concentration $C_f$ of an anionic hydrogel is given as (Grimshaw et al., 1990)

$$C_f = \frac{C_f^0 K}{H (K + C_H^0)}$$

where $C_f^0$ is the initial fixed charge concentration at the hydrogel dry state, $H$ is the ratio of final to the initial volume of the hydrogel, and $K$ is the dissociation constant of the fixed charge groups.

2.6 Mechanical field for constitutive equation

From Eq. (15), when the system is in mechanical equilibrium the term in the first bracket vanishes and the Piola stress $P$ is given as follows (Hong et al., 2008)

$$P = -(1 + C\nu)^{2/3} p F_e^{-T} + Nk_BT (1 + C\nu)^{1/3} F_e + (1 + C\nu)^{2/3} F_e^{1/3} \frac{1}{\varepsilon} (H \otimes H)$$

where the Piola stress $P$ is balanced by the osmotic pressure $p$, elastic polymeric network chains stress, and Maxwell stress. At the equilibrium state, the Maxwell stress is neglectable (Drozdov and deClaville Christiansen, 2015c), such that the Piola stress $P$ is reduced to

$$P = -(\lambda_s^2 p F_e^{-T} + Nk_BT (\lambda_s) F_e$$

Again from Eq. (15), when the system is at steady-state, the chemical potentials of the interior and exterior solutions are given as follows (Illeperuma et al., 2013)

$$\mu = RT \sum \frac{C_k}{C} - RT \left( \ln \frac{C\nu}{(1 + C\nu)} - \frac{1}{(1 + C\nu)^2} + \frac{\chi}{(1 + C\nu)^2} \right) - Nk_BT \left[ \frac{\nu}{(1 + C\nu)^{1/3}} \left( \frac{1}{3} \text{tr}(C_e) - \frac{1}{(1 + C\nu)^{2/3}} \right) \right] - \nu p$$

$$\bar{\mu} = RT \sum \frac{\bar{C}_k}{\bar{C}}$$

(27)
where the overbar indicates the parameters in the surrounding solution.

By adopting the Donnan equilibrium, where the chemical potentials in the interior and exterior solutions coincide over the hydrogel-solution interface, namely \( \mu = \bar{\mu} \), the osmotic pressure \( p \) is given as (Drozdov and deClaville Christiansen, 2015b)

\[
p = RT \sum_k \left( \frac{C_k}{J-1} - \bar{C}_k \right) - \frac{RT}{\nu} \left( \ln \frac{C_v}{(1+C_v)} - \frac{1}{(1+C_v)} + \frac{\chi}{(1+C_v)^2} \right) \\
- Nk_B T \left[ \frac{1}{(1+C_v)^{1/3}} \left( \frac{1}{3} \text{tr}(C_e) - \frac{1}{(1+C_v)^{2/3}} \right) \right]
\]

where the first term on the right-hand side arises due to the imbalance distribution of ionic concentrations between the hydrogel and surrounding solution, the second term is due to the mixing of the solvent and hydrogel, and the third term is due to the elastic stretching of the polymeric network chains. It is noted that the osmotic pressure \( p \) here is consistent with the published results in the open literature (Duda et al., 2010; Marcombe et al., 2010).

2.7 Boundary conditions

In this section, to investigate the responsive behavior of the cylindrical-shaped urea-sensitive hydrogel immersed in the flow of urea-buffer solution, one-dimensional steady-state numerical simulations of the MECurea model are conducted. The hydrogel is taken as symmetrical and has a large length/radial ratio property, where the deformation of the urea-sensitive hydrogel is taken at the radial direction and restricted in the axial direction, such that the deformation of the hydrogel is a one-dimensional problem. (Deen, 1998). The computation domain includes both the hydrogel and buffer solution, while the momentum Eq. (2) only covers the hydrogel domain.

The Neumann type boundary conditions are imposed at the center of the cylindrical hydrogel to ensure the continuity in the hydrogel, given as follows
\[
\frac{\partial C_k}{\partial X} = 0, \quad \frac{\partial \Phi}{\partial X} = 0 \quad (k = H^+, OH^-, ..., N) \quad \text{at} \quad X = 0
\]  

(29)

while the Dirichlet boundary conditions along the solution

\[
C_k = \bar{C}_k, \quad \Phi = 0 \quad (k = H^+, OH^-, ..., N)
\]  

(30)

In order to consolidate the mechanical equilibrium equation with Eq. (2), Eq. (25) is further elaborated to the following form

\[
P = -\left(\lambda_s\right)^2 p(1) + Nk_B T\left(1 + \frac{\partial u_s}{\partial X}\right)
\]  

(31)

where the radial swelling stretch \( \lambda_s \) is given as

\[
\lambda_s = \frac{\partial x_s}{\partial X} = 1 + \frac{\partial u_s}{\partial X} = \frac{L_{gel}}{L_{gel}^o}
\]  

(32)

where \( u_s \) is the swelling displacement and \( L_{gel} \) is the diameter of the gel.

By Eq. (31), the Piola stress \( P \) is further rewritten as follows

\[
P = -\left(1 + \frac{\partial u_s}{\partial X}\right)^2 p(1) + Nk_B T\left(1 + \frac{\partial u_s}{\partial X}\right)
\]  

(33)

and by substituting Eq. (33) into the momentum Eq. (2) we obtain

\[
\frac{\partial}{\partial X}\left[-\left(1 + \frac{\partial u_s}{\partial X}\right)^2 p + Nk_B T\left(1 + \frac{\partial u_s}{\partial X}\right)\right] = 0
\]  

(34)

Finally, the swelling stretch \( \lambda_s \) between the hydrogel-solution interfaces is expressed as

\[
\lambda_s = \frac{Nk_B T}{p}, \quad X = L_{gel}
\]  

(35)

In order to simulate the responsive behavior of the hydrogel, the commercial finite-element solver, COMSOL Multiphysics 5.1 is employed for the numerical solution of the MECurea model due to the ability in efficiently handling the coupled governing equations with desired
computational accuracy. As such, the fixed charge density Eq. (23) is first solved to obtain the concentration of the fixed charge \( C_f \) according to the given boundary conditions and input parameter. By the computed \( C_f \), the Nernst-Planck equation Eq. (1) coupled with the Poisson equation Eq. (22) are solved numerically for the converged solution of mobile species concentration \( C_k \) and electric potential \( \Phi \). The converged solution of mobile species concentration \( C_k \) is then employed into the nonlinear mechanical equilibrium equation Eq. (35) for the corresponding swelling stretch \( \lambda_s \). Lastly, the stretching of the hydrogel causes the redistribution of the mobile ions and fixed charge groups within the hydrogel, such that a new iteration is started until all the independent variables reach convergence, including \( C_k \), \( \Phi \), \( C_f \) and \( \lambda_s \).

3. Result and discussions

3.1 Validation of the MECurea model

To validate the MECurea model, the equilibrium swelling behavior of urea-sensitive hydrogels is examined by comparing the present numerical result with the experimental data, as shown in Fig. 2, in which a good agreement is achieved. In the experimental work conducted by Ogawa and Kokufuta (Ogawa and Kokufuta, 2002), the urea-sensitive p(NIPAM)-based hydrogels with diameter of 290 \( \mu \)m was immersed in the flow of maleate-buffer of 5 mM, containing the urea of \( c_{\text{urea}} = 1 \) mM, with pH = 4, at the temperature of 35 °C, such that the urease concentration \( C_{E_0} \) of 0.2 to 8.0 mg/mL immobilized in the hydrogel. Several input are required by the MECurea model for the numerical simulation of the hydrogel, such as \( L = 2500 \mu \)m, \( z_k = -1 \), \( C_{f_0} = 0.775 \) mmol/g, \( K = 10^{-5.57} \), while the remaining inputs are given in Table 1.
Fig. 2 indicates that the hydrogel swells drastically less at a higher concentration of immobilized urease $C_{E_0}$ (i.e. more than 0.5 mg/mL), such that the entrapped urease improves the mechanical properties of the polymeric network chains, resulting in a stronger hydrogel and limit the swelling of the urea-sensitive hydrogel. For the hydrogels with a smaller concentration of the immobilized urease $C_{E_0}$ (i.e. less than 0.5 mg/mL), the present numerical results usually overestimate the experimental equilibrium swelling behavior as compared to the published numerical results, however the errors of the present numerical simulations are usually within the experimental error of ±13 μm, as seen in Fig. 2. Based on the comparison in Fig. 2, it is concluded that the numerical result of the MECurea model agrees well with the experimental findings.

3.2 The performance of the hydrogel in response to the urea cue

A case study through the numerical simulation of the MECurea model is conducted for the response of the anionic urea-sensitive hydrogel with $z_f = -1$ subject to the urea at different environmental temperatures. The material properties required by the MECurea model are given as $L = 250 \mu$m, $L_{gel}^0 = 50 \mu$m, $C_j^0 = 1800$ mM, $K = 10^{-20}$, and the other input parameters are tabulated in Table 1. For the present simulation, the hydrogel is immersed in the phosphate-buffered saline (PBS) solution ($\overline{C_{Na^+}} = 142$ mM and $\overline{C_{Cl^-}} = 103$ mM), where the urea concentration $\overline{C_{urea}}$ ranging from 0 to 2000 mM at pH=7.40 is taken as the input with the solution temperature of 34 ~ 40 °C with 0.5 mg/mL urease concentration $C_{E_0}$ immobilized in the hydrogel.
Fig. 3 is plotted to illustrate the responsive deformation of the urea-sensitive hydrogel subject to the change of urea concentration $\bar{C}_{\text{urea}}$ in the buffer solution at different environmental temperatures. It is found that the hydrogel collapses with the increase of the urea concentration $\bar{C}_{\text{urea}}$, where the hydrogel further collapses with the increase of the surrounding temperature. Furthermore, the responsive behavior of the hydrogel is divided into three stages, as shown in Fig. 3: (1) the hydrogel shows no noticeable change of swelling ratio with the increment of the environmental urea concentration $\bar{C}_{\text{urea}}$ smaller than 1 mM, (2) the hydrogel starts to collapse, when the environmental urea concentration $\bar{C}_{\text{urea}}$ is larger than 1 mM, and (3) the hydrogel begins to swell again, when the environmental urea concentration $\bar{C}_{\text{urea}}$ is larger than 400 mM. As such, the mechanical sensitivity of the hydrogel falls in the range of urea concentration $\bar{C}_{\text{urea}}$ 1 to 100 mM, since the hydrogel collapses drastically at this range of urea concentration $\bar{C}_{\text{urea}}$, as shown in Fig. 3.

Fig. 4 is plotted to visualize the distributive electrical potential within the hydrogel and surrounding solution subject to different concentrations of urea $\bar{C}_{\text{urea}}$. At $\bar{C}_{\text{urea}} = 100$ and 200 mM, where the fixed charge groups are mainly ionized, a constant electrical potential is found within the hydrogel, while the electrical potential $\Phi$ is equal to zero in the external solution since there is electroneutrality state in the total concentrations of all the mobile ionic solutes. To our best knowledge, this is the first example of biomaterial which exhibits the electrical potential gradient in response to the urea cue above 1000 mM, as shown in Fig. 4.

Fig. 5 illustrates the distribution profile of the pH in both the hydrogel and surrounding solution, when immersed in the urea-buffer solution of $\bar{C}_{\text{urea}} = 0.1, 1.0, 10, \text{ and } 100 \text{ mM}$. It is found that the solution in the hydrogel becomes more acidic as the concentration of urea $\bar{C}_{\text{urea}}$
increases, where a further increase of the concentration of urea $C_{\text{area}}$ (i.e. above 1.0mM) results in the pH gradient in the hydrogel. It has been reported in the open literature that the enzyme-induced pH gradient behavior has been employed in designing the biofuel cell, where the pH gradient is used to generate the potential difference in the biofuel cell system (Giroud et al., 2011). As such, the present urea-sensitive hydrogel is a promising material in the field of biofuel cell as it exhibits both the enzyme-induced pH and electrical potential gradients behavior, when subject to the urea concentration $C_{\text{area}}$ higher than 1000 mM, as seen in Fig. 5. Therefore, the urea-sensitive hydrogel could provide an innovative means for electricity generation from the human urine, which creates a new and affordable way of generating electrical power.

4. Conclusions

It is found that the urea-sensitive hydrogel is a promising material in the field of biofuel-cell as it exhibits both the enzyme-induced pH and electrical potential gradients behaviors in response to the urea, where it could provide a means for electricity generation from human urine. In future work, the MECurea model can be utilized to get a greater insight into the fundamental mechanism of the hydrogel in response to the urea cue.

Acknowledgement

The authors gratefully acknowledge the financial support from Nanyang Technological University through the project (No: M4081151.050) and NTU Research Scholarships.
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**Table 1. Input data for the numerical simulation of the MECurea model.**

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Diffusion coefficients $D_{H^+}$ (Plawsky, 2014)</td>
<td>$9.31 \times 10^{-9}$ m$^2$/s</td>
</tr>
<tr>
<td>Diffusion coefficients $D_{OH^-}$ (Plawsky, 2014)</td>
<td>$5.24 \times 10^{-9}$ m$^2$/s</td>
</tr>
<tr>
<td>Diffusion coefficients $D_{NH_3}$ (Plawsky, 2014)</td>
<td>$1.98 \times 10^{-9}$ m$^2$/s</td>
</tr>
<tr>
<td>Diffusion coefficients $D_{HCO_3^-}$ (Plawsky, 2014)</td>
<td>$1.18 \times 10^{-9}$ m$^2$/s</td>
</tr>
<tr>
<td>Flory interaction parameter $\chi$ (Bae et al., 1990)</td>
<td>$0.5090-0.9975$</td>
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<tr>
<td>Permittivity $\varepsilon_r \varepsilon_0$ (water)</td>
<td>$7.0832 \times 10^{-10}$ A s/V.m</td>
</tr>
<tr>
<td>Michaelis constant $K_M$ (Ogawa and Kokufuta, 2002)</td>
<td>$2.82$ mol/m$^3$</td>
</tr>
<tr>
<td>Reaction rate constant $k_{cat}$ (Chen and Chiu, 1999)</td>
<td>$1.5 \times 10^3$ 1/s</td>
</tr>
<tr>
<td>Deactivation rate constant $k_d$ (Chen and Chiu, 1999)</td>
<td>$2.18 \times 10^2$ 1/s</td>
</tr>
<tr>
<td>Urease equilibrium temperature $T_{eq}$ (Krajewska et al., 1990)</td>
<td>298.15 K</td>
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<tr>
<td>Urease optimum pH</td>
<td>7.4</td>
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<tr>
<td>Equilibrium constant $K_W$</td>
<td>$10^{-14}$</td>
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<tr>
<td>Equilibrium constant $K_H$ (Fidaleo and Lavecchia, 2003)</td>
<td>$7.57 \times 10^{-4}$ mol/ m$^3$</td>
</tr>
<tr>
<td>Enthalpy change for transition from the active to inactive enzyme $\Delta H_{eq}$ (Chen and Chiu, 2000; Peterson et al., 2007)</td>
<td>89594 J/mol</td>
</tr>
</tbody>
</table>
Fig. 1. Schematic diagram of the urea-sensitive hydrogel system.
Fig. 2. Comparison of the equilibrium swelling ratio for the hydrogel between the simulation results by the MECurea model and Ogawa and Kokufuta’s correlation, and their experimental data (Ogawa and Kokufuta, 2002) at initial radius 145.0μm.
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Highlights

- A 3D model has been developed to characterize urea-sensitive hydrogel as biosensor.
- The model couples the effects of biochemical, electro and mechanical together.
- The electrical, chemical and mechanical performances of the hydrogel are investigated.
- The hydrogel may generate electricity via the enzyme-induced pH and electrical potential gradients.