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Microfluidic platform for controlling the differentiation of embryoid body

Wai-To Fung*, Ali Beyzavi§, Patrick Abgrall §, Nam-Trung Nguyen** and Hoi-Yeung Li***

Embryonic stem (ES) cells are pluripotent cells, which can differentiate into any cell types. This cell type has often been implicated as an eminent source of renewable cells for tissue regeneration and cellular replacement therapies. Studies on manipulation of the various differentiation pathways have been at the forefront of research. There are many ways in which ES cells can be differentiated. One of the most common techniques is to initiate the development of embryoid bodies (EBs) by in vitro aggregation of ES cells. Thereafter, EBs can be induced to undergo differentiation into various cell lineages. In this report, we present a microfluidic platform using biocompatible materials, which is suitable for culturing EB. The platform is based on a Y-channel device with two inlets for two different culturing media. An EB is located across both streams. Using the laminar characteristics at low Reynolds number and high Peclet numbers, we have induced cell differentiation on a half of the EB while maintaining the other half in un-induced stages. The results prove the potential of using microfluidic technology for manipulation of EBs and ES cells in tissue engineering.

Keywords: embryoid bodies, differentiation, microfluidics, laminar flow, diffusion

Introduction

Embryonic stem (ES) cells are derived from the inner cell mass of the blastocyst of a developing embryo. They are pluripotent cells which could virtually give rise to any cell type including neural cells, cardiomyocytes and hepatocytes through cellular differentiation. ES cells are self-renewable and for that reason, they have often been implicated as an eminent source of renewable cells for tissue regeneration and cellular replacement therapies. Hence, it is not surprising that the control and manipulation of the various differentiation pathways in ES cells have been at the forefront of research.

There are many ways to differentiate ES cells. One of the most common technique is to initiate the development of embryoid bodies (EBs) by in vitro aggregation of ES cells

Thereafter, EBs can be induced to undergo differentiation into the various cell lineages. To date, no attempt was ever made to sub-differentiate a single EB into more than one lineage. This is mainly compounded by the limitations of conventional culturing techniques, which do not permit an EB to be induced by more than one differentiation media at the same time.

Microfabricated culturing systems can circumvent this limitation and permit the control of the differentiation process through manipulating the environment of the culture in both time and space, which is almost impossible to achieve using conventional culturing protocols. The behaviour of fluids at the microscale is different from that in macroscale. For microfluidic systems some gripping and counterintuitive properties appear. When two fluids streams come together in a microchannel, a parallel laminar flow can be generated without turbulence. The Reynolds number, which represents the ratio between inertial force and viscous force, is in microscale often less then unity. At these low Reynolds numbers, the Peclet number, which represents the ratio between advection and diffusion, are still in the order of 1000. Thus a microchannel would allow two liquids to follow side by side without major intermixing. This unique property of liquid in microchannel enables possibilities and new tools for biological research.

With recent advance on design and fabrication of microfluidic platform, microfluidic platform has been applied in research areas including stem cell and developmental biology. Manipulation of microenvironments, such as temperature, pH, mechanical stimuli and growth factor gradient can be achieved by microfluidic platform, which will be useful in identifying factors involved in stem cell proliferation and differentiation. Interestingly, a gradient-generating microfluidic platform has been applied to study the effects of a continuous gradient of epidermal growth factor, fibroblast growth factor 2, and platelet-derived growth factor on the growth and differentiation of neural stem cells.

Interestingly, colony formation of murine embryonic stem cells can be influenced by different flow rates generated by microfluidic platform although the mechanism behind the flow rate dependent effects remains uncertain. Furthermore, temperature gradient created by microfluidic platform revealed the importance of temperature on development of Drosophila embryo. Cell fusion between mouse embryonic stem cells and mouse embryonic fibroblasts was...
Microfluidic platform has been shown to facilitate individual gene expression profiling on neural stem cells. To further improve the capability of microfluidic platform, 3D microfluidic cell culture systems have been developed, which offers a similar in vitro 3D microenvironment as in vitro. Clearly, microfluidic platform has become a new tool for stem cell and developmental biology research from 2D to 3D.

In most of the previous experimental investigations on EB differentiation, EBs were only subjected to a single differentiation condition. Here we demonstrated that by independently cultivating the two halves of an EB in two separate media resulting from laminar co-flow in a microchannel, cell differentiation can be induced on a half of the EB while the other half of the EB maintained in un-induced stages can be achieved.

### Materials and Methods

**Fig. 1** Schematic diagram illustrating the components of the microfluidic device (A). Photographs of the microfluidic chip (B) and the assembled microfluidic device (including holders, microfluidic chip and tubings) (C).

**Mouse ES cell culture**

E14 Mouse ES (mES) cells were cultured in dishes coated with 0.1% w/v gelatin (Sigma) and maintained in a complete medium consisting of Dulbecco’s Modified Eagle’s Medium (DMEM; Gibco) containing 20% fetal bovine serum (FBS; HyClone), 0.1mM 2-mercaptoethanol (Sigma), 1% MEM non-essential amino acids (Gibco), 100 U/ml penicillin (Invitrogen), 100 U/ml streptomycin (Invitrogen) and 1000 U/ml ESGRO-LIF (Chemicon), in a humidified incubator (37°C in an atmosphere of 5% CO₂), with the medium changed every 2 days. To induce EB formation, mES cells were dissociated into a single-cell suspension by 0.05% trypsin and 0.04% EDTA in PBS and plated onto non-adherent bacterial culture dishes and maintained in the complete DMEM medium without LIF for 3 days on a 3D shaking platform in a humidified incubator (37°C in an atmosphere of 5% CO₂). After two days incubation, the EB attached and spread onto the tissue culture dishes, and then medium was changed into either differentiation medium [complete L15 medium (Gibco) added with 10μM retinoic acid (RA; Sigma)], or fresh complete L15 medium in a humidified incubator (37°C) with medium changed every 2 days. After 5 days incubation, the EB was then collected and lysed with lysis buffer (PIERCE). 20μg of EB lysate (5-10 EBs) was used in each lane of western blot analysis.

**Western blot analysis**

Mouse anti-PCNA (Santa Cruz), mouse anti-neurofilament 68kDa (NF68) (Abcam), and mouse anti-GAPDH (Santa Cruz) antibodies were used in these experiments. Total proteins were extracted with M-PER mammalian protein extraction reagent (Thermo Scientific). 20μg of EB lysate (5-10 EBs) was loaded on each lane and then separated by 12% SDS-PAGE and transferred onto a nitrocellulose membrane (Whatman). The membranes were incubated with antibodies to specific protein followed by incubation with secondary antibodies: polyclonal goat anti-mouse immunoglobulins/HRP (Dako), polyclonal mouse anti-goat immunoglobulins/HRP (Dako) or polyclonal rabbit anti-goat immunoglobulins/HRP (Dako), and developed with chemiluminescence reagent (Thermo Scientific).

**Fabrication of the PMMA/PDMS microfluidic platform**

A Y-shape channel was cut on a rectangular (76.2x25.4mm) piece of polymethylmethacrylate (PMMA). A CO₂ laser machining system (Universal M-300 Laser Machining Platform, Universal Laser Systems Inc., Arizona, USA) was used for cutting the 1 mm-thick PMMA sheet (Goodfellow, England). Two layers of polydimethylsiloxane (PDMS) at the top and at the bottom of the Y shape channel were used for sealing of the device (Figure 1A). The PDMS sheet with the thickness of 0.5mm was prepared by spin coating the PDMS mixture (Dow Corning Corporation, USA) at 500 rpm for 30 seconds. The prepolymer and the curing agent of PDMS were mixed with a weight ratio of 10:1. The thin PDMS layer was then cured in an oven for 4 hours at 90°C. Holes were cut in the top layer of PMMA and PDMS to provide inlet and outlet for the media. The microfluidic chip consisted of two PDMA layers (1mm thick), two layers of PDMS (0.5mm thick) and one glass slide. Thus, the internal dimension (width and
Determination of the minimum flow rate of the medium

To optimize the parallel laminar flow with minimum consumption of the culturing medium, different flow rates were examined. It was found that all the tested flow rates could maintain a parallel laminar flow without intermixing between the fluorescence labelled medium and the non-labelled medium (Figure 2A and B). Hence, a flow rate of 50 μl/min was chosen in the subsequent experiments to minimize the consumption of the culturing medium.

Biocompatibility of PDMS for the growth and differentiation of embryoid bodies

The microfluidic chip was assembled as shown in Figure 1 to test the ability of PDMS to support the growth and differentiation of the embryoid body. A three-day EB (Figure 3Aa) was transferred by a pipette tip into the microchannel, and the whole microfluidic chip was immersed in the complete DMEM in a 100mm tissue culture dish. The EB was incubated at 37°C for another 2 days (Figure 3Ab). When the EB was attached onto the PDMS layer of the microfluidic chip, another two layers of PDMS and PMMA were placed on the PMMA layer as illustrated in Figure 1. A peristaltic pump was connected to the two inlets of the microchannel, which provided a continuous flow of either complete L-15 medium (Gibco), to maintain the normal growth of EB, or complete L15 medium with 10μM RA, to induce cell differentiation of EB, at a flow rate of 50μl/min, respectively. The microfluidic device was then incubated in the live imaging system (Carl Zeiss). The growth of EB was monitored by time-lapse microscopy (Carl Zeiss) (Figure 3B). After 5 day of incubation at 37°C, the microfluidic device was disassembled and the EB in the microchannel was analyzed by immunofluorescence staining using antibodies against Ki67 as proliferation marker and NF160 as cell differentiation marker. It was found that there was strong Ki67 staining on the EB incubated with complete L15 medium, suggesting that growth of EB was not affected by PDMS. Most importantly, addition of RA induced cell differentiation of EB in the microfluidic channel as suggested by the positive immunostaining of NF160 (Figure 3C). Taken together, PDMS has no adverse effect on the growth and differentiation of EB.

To investigate the effects of RA on changes in rate of cell proliferation and induction of cell differentiation during EB development, western blot analysis was performed. Our results showed that the level of PCNA, the proliferation marker, in EBs cultured in complete L15 medium was higher than those in complete L15 medium with 10μM RA after 5 days of incubation (Figure 3D and 3E). The two-tailed P value...
null
A red fluorescence dye was added into one of the media to further confirm that the laminar flow was not disturbed during the incubation. The fluorescence intensity across the channel was measured (data not shown). Neurofilament (NF160) was only found in one half of the EB, while the other half has strong staining of Ki67 (Figure 4B and Electronic Supplementary Information Figure 1). Consistently, we found that the expression of NF68 was present in the EB supplied with complete L15 medium with 10 μM RA and the EB under the laminar flow of complete L15 medium and complete L15 medium with 10 μM RA (Figure 4C and 4D). Most importantly, the level of PCNA for the EBs cultivated under the laminar flow of complete L15 medium and complete L15 medium with 10 μM RA (L15/L15+RA) is significantly higher than those cultivated in complete L15 medium (L15/L15) and complete L15 medium with 10 μM RA (L15+RA/L15+RA) (Figure 4C and 4D) (The two-tailed P value equals 0.0014 By conventional criteria, this difference is considered to be statistically significant), suggesting portion of the EB remains un-induced and proliferative.

Conclusions

In conclusion, we have fabricated a microfluidic device and cultured EB derived from mouse ES cells in this device. Furthermore, we have applied the microfluidic device in EB differentiation, resulting in differentiated and un-induced cells in different areas of the same EB simultaneously. Specific differentiation into different specialized cell lineages on the same EB can be achieved by applying microfluidic technology.

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References
