

Inhibition Properties of Arylsulfatase and β -Glucuronidase by Hydrogen Peroxide, Hypochlorite, and Peracetic Acid

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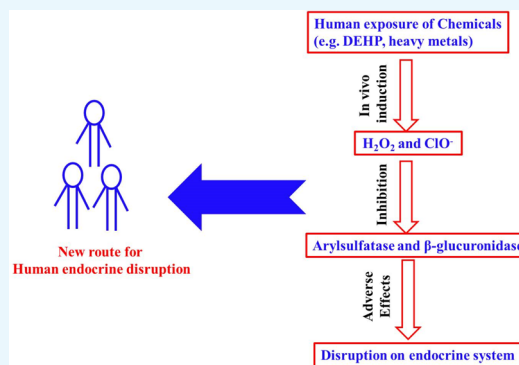
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ABSTRACT: Arylsulfatase and β -glucuronidase are two important enzymes in humans, which play an important role in the dynamic equilibrium of steroidal estrogens. This work probably for the first time reported that hydrogen peroxide (H_2O_2), hypochlorite, and peracetic acid (PAA) could effectively inhibit the activities of arylsulfatase and/or β -glucuronidase. The 50% of inhibitions (IC_{50}) of H_2O_2 , hypochlorite, and PAA on arylsulfatase were found to be 142.90 ± 9.00 , 91.83 ± 10.01 , and $43.46 \pm 2.92 \mu M$, respectively. The corresponding IC_{50} values of hypochlorite and PAA on β -glucuronidase were 704.90 ± 41.40 and $23.26 \pm 0.82 \mu M$, whereas H_2O_2 showed no inhibition on β -glucuronidase. The inhibitions of arylsulfatase and/or β -glucuronidase by these three chemicals were pH-dependent. It was further revealed that the inhibitions of hypochlorite on both arylsulfatase and β -glucuronidase were irreversible. On the contrary, the inhibitions by H_2O_2 and PAA were reversible. In addition, the inhibition by H_2O_2 was competitive and that by PAA was noncompetitive. In general, H_2O_2 and hypochlorite can be endogenously produced in humans, which suggested that the two compounds are potential endocrine disruption compounds (EDCs) as they can cause endocrine disruption via the inhibition of arylsulfatase and β -glucuronidase. This work further indicated that any agent that can induce the production of H_2O_2 or hypochlorite in humans is a potential EDC, which explains why some EDCs with very weak or no estrogenic potency can cause endocrine disruption, which is confirmed in epidemiological studies.



INTRODUCTION

Arylsulfatase and β -glucuronidase have been widely present in many organisms including humans, which are able to catalyze the hydrolysis of sulfates/glucuronides.^{1–4} The important physiological functions of the two enzymes are to maintain the dynamic equilibrium of natural estrogens in human body. Normally, only about 2–3% of natural estrogens are free estrogens in human, whereas the rest are in sulfate or glucuronide form.^{5,6} The free natural estrogens play important roles in stimulating growth, blood flow, water retention in sexual organs, neuro- and vasoprotection, and reduction of bone loss, whereas the conjugated estrogens act as reservoirs.^{5,7} Natural free estrogens in humans are accurately regulated, and once the equilibrium is disrupted, it may trigger a severe outcome. For example, humans with a significantly higher urinary arylsulfatase/ β -glucuronidase activity have higher free estrogens in their body, which have been thought to be the main reason for the development of many cancers, including breast cancer, stomach cancer, ovarian cancer, colonic cancer, and so forth.^{8–11} Therefore, the inhibition of arylsulfatase/ β -glucuronidase has been explored as one of the effective strategies for such cancer treatment, among which STX64 has been applied in clinical trials.^{7,12}

Arylsulfatase and β -glucuronidase have been reported to be two extreme enzymes, which can endure high concentrations of mercury dichloride, sodium azide, ethanol, and ethylenediaminetetraacetic acid.¹³ H_2O_2 , hypochlorite, and peracetic acid (PAA) have been commonly used as sanitizers for various medical and nonmedical applications.^{14–17} Among these, H_2O_2 and hypochlorite are also important compounds which can be endogenously produced by cellular enzymatic reactions in humans and are important signal biomarkers in humans.^{18–21} This work is the first to report that H_2O_2 , hypochlorite, and PAA can effectively inhibit arylsulfatase and/or β -glucuronidase, which may indicate important physiological functions.

MATERIALS AND METHODS

Enzymes and Chemicals. Potassium 4-nitrophenyl sulfate (*p*NPS, purity > 98%), 4-nitrophenyl- β -D-glucuronide (*p*NPG,

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purity > 98%), *p*-nitrophenol (*p*NP, spectrophotometric grade), arylsulfatase ($\geq 10,000$ units/g solid) from *Helix pomatia* (type H-1, catalog number: S9626), and β -glucuronidase ($\geq 100,000$ units/g solid) from *H. pomatia* (type H-1, catalog number: G0751) were purchased from Sigma-Aldrich (Shanghai, China). H_2O_2 (30%) was purchased from Guangzhou Chemical Reagent Factory (Guangzhou, China). NaClO (available chlorine, 8.0%, w/w) was purchased from ANPEL (Shanghai, China). The NaClO concentration was calibrated with the standard method of GB/T 19106.²² PAA (13%, w/w) including solutions I and II was purchased from GHTECH (Shantou, China). Before using, fresh PAA was obtained by mixing the solutions I and II in the proportion of 1:1 (v/v), and it was used within 2 days. This PAA concentration was calibrated according to the standard method of GB/T 19104.²³ The other reagents not listed were purchased from Aladdin (Shanghai, China).

Stock solutions of arylsulfatase and β -glucuronidase were prepared at the concentration of 10 U/mL by dissolving the enzyme powder into 0.5 M Tris-HCl buffer (pH 7.0). The *p*NPS stock solution was prepared with 0.5 M acetate buffer (pH 5.8), whereas the *p*NPG stock solution was prepared with 0.5 M phosphate buffer (pH 7.0), both of which were at concentrations of 5 mM. All solutions were prepared with reagent-grade chemicals and ultrapure water (18.2 M Ω /cm).

Enzyme Activity. The activity of arylsulfatase/ β -glucuronidase was measured according to Zhang et al.¹³ The arylsulfatase/ β -glucuronidase enzyme activity (U) was defined as the absorbance equivalent of 1 μmol *p*NP produced per hour per milliliter of enzyme solution at 37 °C. For the convenience of comparison, relative activity is used, and the maximal enzyme activity is arbitrarily set to 100%. The limit of detection (LOD) of the spectrophotometric method was calculated according to three times of the standard deviation, which was obtained from nine repeated determinations of the lowest *p*NP concentration used for the standard calibration curve.^{24,25} The LOD of this method for *p*NP was 0.22 μM , which is equal to 0.22 U/L for arylsulfatase and β -glucuronidase.

The Michaelis–Menten constant (K_m) and maximal reaction rate (V_{max}) of the two enzymes, arylsulfatase and β -glucuronidase, were obtained by fixing the enzyme concentration in the reaction and then analyzing the initial speed at several different substrate concentrations. The concentrations of the substrate in the reaction mixture were 0.02, 0.04, 0.08, 0.1, 0.2, 0.4, 0.8, 1.0, and 1.2 mM, respectively. The rest of the steps were the same as those for measuring the enzyme activity.

K_m and V_{max} were expressed by the Michaelis equation,²⁶ that is

$$v_0 = \frac{V_{\text{max}} \times [s]}{K_m + [s]} \quad (1)$$

K_m and V_{max} are calculated by the Lineweaver–Burk equation,²⁶ that is

$$\frac{1}{V} = \frac{K_m}{V_{\text{max}}} \frac{1}{[s]} + \frac{1}{V_{\text{max}}} \quad (2)$$

where V_0 is the production rate of the substrate, K_m is called the Michaelis constant, V_{max} is the reaction rate when the enzyme is saturated with the substrate, and $[s]$ is the concentration of the substrate.

Inhibitory Assays. Inhibition experiments on arylsulfatase or β -glucuronidase by different inhibitors were performed with different concentrations, and the inhibitor solution was freshly

prepared before use. For the arylsulfatase inhibition experiment, the reaction mixture solution (3.5 mL) contained 2 mL of 0.1 M acetate buffer (pH 5.8), 0.5 mL of dilute arylsulfatase enzyme, 0.5 mL of 5 mM *p*NPS, and 0.5 mL of water as the control or 0.5 mL of the inhibitor (H_2O_2 , NaClO, or PAA) with different concentrations. The addition concentrations of H_2O_2 and NaClO in the reaction mixture were 1, 5, 10, 50, 100, 500, 1000, 5000, and 10,000 μM , respectively, whereas the concentrations of PAA were 1, 5, 10, 50, 100, 500, and 1000 μM , respectively. The reaction mixture solution was mixed and incubated for 1 h at 37 °C. *p*NP deconjugated from *p*NPS was measured at a wavelength of 400 nm after the reaction was stopped by adding 1.5 mL of 0.5 M NaOH. Each experiment was performed in triplicate.

For β -glucuronidase inhibition experiment, the reaction mixture solution (1.2 mL) contained 0.6 mL of 0.1 M phosphate buffer (pH 7.0), 0.2 mL of diluted β -glucuronidase enzyme, 0.2 mL of 5 mM *p*NPG, and 0.2 mL of water as the control or 0.2 mL of H_2O_2 , NaClO, or PAA. The addition concentrations of H_2O_2 in the reaction mixture were 1, 5, 10, 50, 100, 500, 1000, 5000, and 10,000 μM , respectively, whereas the concentrations of NaClO and PAA were the same with 1, 5, 10, 50, 100, 500, and 1000 μM , respectively. The reaction mixture solution was mixed and incubated for 1 h at 37 °C. *p*NP deconjugated from *p*NPG was measured at a wavelength of 400 nm after the reaction was stopped by adding 0.8 mL of 0.5 M NaOH. Each experiment was performed in triplicate.

To check the inhibition differences at different pH conditions, different buffer solutions were used as listed: 0.1 M acetate buffer (pH 5.0–5.8), 0.1 M citric acid/0.1 M sodium citrate (pH 5.0–6.0), 0.5 M Tris-HCl (pH 7.0–8.0), and 0.1 M phosphate buffer (pH 6.0–8.0).

In order to explore the types of enzyme inhibition by different inhibitors, we used four different concentrations of inhibitors to perform inhibition experiments at different enzyme concentrations. The enzyme activities in the reaction solution were 0.01, 0.02, 0.03, 0.04, 0.05 and 0.06 U/mL, respectively. For arylsulfatase, the reaction concentrations of H_2O_2 were 0, 50, 150, and 500 μM , and the reaction concentrations of NaClO were 0, 50, 100, and 200 μM , whereas the reaction concentrations of PAA were 0, 10, 50, and 100 μM . For β -glucuronidase, the reaction concentrations of NaClO were 0, 500, 1000, and 2000 μM , whereas the reaction concentrations of PAA were 0, 50, 100, and 500 μM . The other conditions were consistent with the enzyme inhibition experiments.

Inhibition percentage was calculated as shown in eq 3

$$I(\%) = 100 - 100 \times (A/C) \quad (3)$$

where I is the inhibition percentage of enzyme activity by an inhibitor, A is the measured enzyme activity with an inhibitor at different concentrations, and C is the measured activity without the addition of an inhibitor. Inhibition dose–response curves were plotted with GraphPad Prism 8 software, and IC_{50} was simultaneously obtained, which means the half inhibition concentration of an inhibitor.

RESULTS AND DISCUSSION

Inhibition Dose–Response Curves. Figures 1 and 2 show the inhibition dose–response curves of H_2O_2 , hypochlorite, and PAA on arylsulfatase and β -glucuronidase. It is evident that hypochlorite and PAA could effectively inhibit both arylsulfatase and β -glucuronidase, whereas H_2O_2 was only effective for inhibiting arylsulfatase. The respective IC_{50} values of H_2O_2 ,

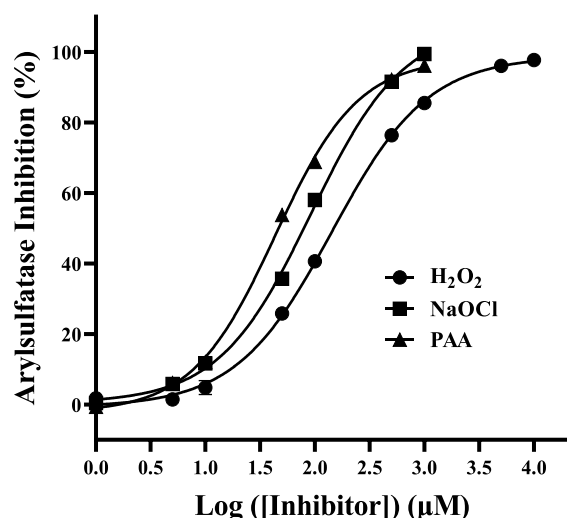


Figure 1. Dose–response curves of H_2O_2 , NaOCl , and PAA on arylsulfatase.

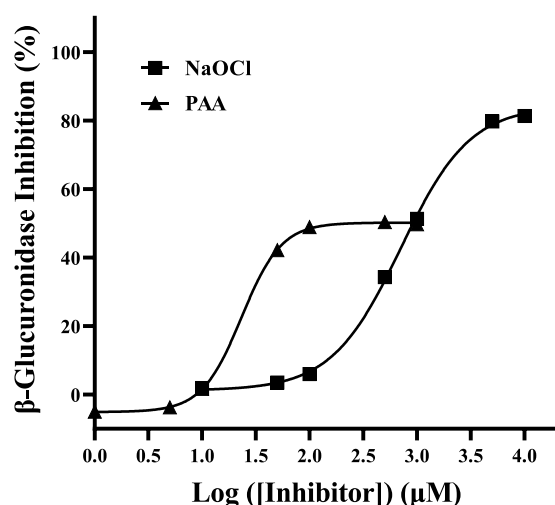


Figure 2. Dose–response curves of NaOCl and PAA on β -glucuronidase. Data for H_2O_2 are not drawn as it showed no inhibition on β -glucuronidase.

hypochlorite, and PAA on arylsulfatase were determined to be 142.90 ± 9.00 , 91.83 ± 10.01 , and $43.46 \pm 2.92 \mu\text{M}$, whereas they were 704.90 ± 41.40 and $23.26 \pm 0.82 \mu\text{M}$ for the inhibition of β -glucuronidase for hypochlorite and PAA. These clearly suggested that PAA had the strongest inhibitory effect on arylsulfatase, followed by hypochlorite, with H_2O_2 being the least. Meanwhile, the inhibitory effect of PAA on β -glucuronidase was much stronger than that of hypochlorite. Interestingly, the inhibitory effect of hypochlorite on arylsulfatase was significantly stronger than that on β -glucuronidase. In contrast, the inhibitory effect of PAA on β -glucuronidase appeared to be stronger than that on arylsulfatase. However, it should be noted that the maximum inhibition degree of PAA on β -glucuronidase was only about 60%.

Effect of pH on Enzyme Inhibition. As H_2O_2 and PAA belong to weak acids and hypochlorite is a weak alkali, it is reasonable to consider that their dissociation degrees are all pH-dependent and subsequently affect their inhibitory powers to arylsulfatase and β -glucuronidase. On the other hand, it has been known that both arylsulfatase and β -glucuronidase could be

completely inhibited at a high pH above 11 or below 3.¹³ Thus, in this study, the pH studied was controlled in the range of 5–8. As can be seen in Figure 3, there were no significant differences

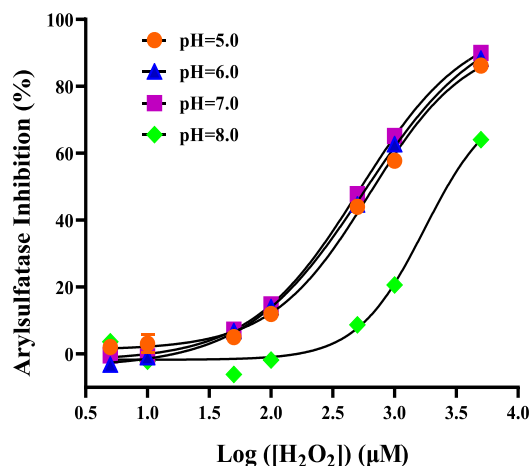


Figure 3. Dose–response curves of H_2O_2 on arylsulfatase under different pH conditions.

in the inhibition of arylsulfatase by H_2O_2 in the pH range of 5–7, whereas reduced inhibition was observed under the weak alkaline condition of pH 8.

Under acidic conditions, hypochlorite can react with hydrogen ions by forming hypochlorous acid (HOCl), which may have different inhibition powers to the two enzymes (data are not shown). It appears from Figure 4A that the inhibition of hypochlorite on arylsulfatase was much stronger at pH 5.0 than at pH 5.8. Moreover, its IC_{50} at pH 5.0 (i.e., $7.64 \pm 0.61 \mu\text{M}$) was 12 times lower than that at pH 5.8 (i.e., $91.83 \pm 10.01 \mu\text{M}$). A similar trend was observed in Figure 4B for β -glucuronidase. These seemed to suggest that the acidic condition was making hypochlorite exhibit more inhibitory potential toward arylsulfatase and β -glucuronidase. The inhibitory effects of PAA on arylsulfatase at different pH are shown in Figure 5. It seemed that inhibition tended to decrease with increasing pH (Figure 5B), for example, nearly no inhibition was observed at pH 8 with a PAA concentration below 1 mM (data are not shown). Similar to arylsulfatase, the inhibitory effect of PAA on β -glucuronidase was insignificant at pH 5 and 6 (Figure 5B).

Inhibition Mechanisms. To explore the possible inhibition mechanisms of arylsulfatase and β -glucuronidase by H_2O_2 , hypochlorite, and PAA, the relationship between enzyme activity and individual inhibitor concentration is presented in Figure 6. It was found that the inhibitions of H_2O_2 and PAA on arylsulfatase was reversible, whereas the inhibition of hypochlorite on arylsulfatase was irreversible. Similar to β -glucuronidase, the inhibition of PAA on β -glucuronidase was reversible, whereas the inhibition of hypochlorite was irreversible. The K_m values of arylsulfatase and β -glucuronidase were 0.217 and 0.736 mM, respectively. To further determine the inhibition kinetics, the Lineweaver–Burk plot was plotted in Figure 7. In Figure 7A, compared with the control group, the experimental group K_m increased and V_{max} did not change, whereas in Figure 7B,C, K_m did not change but V_{max} decreased, showing that the inhibition on arylsulfatase by H_2O_2 was competitive, whereas the inhibitions of PAA on arylsulfatase and β -glucuronidase were noncompetitive.

Significance of This Work. Endocrine-disrupting compounds (EDCs) are, by definition, natural or synthetic agents

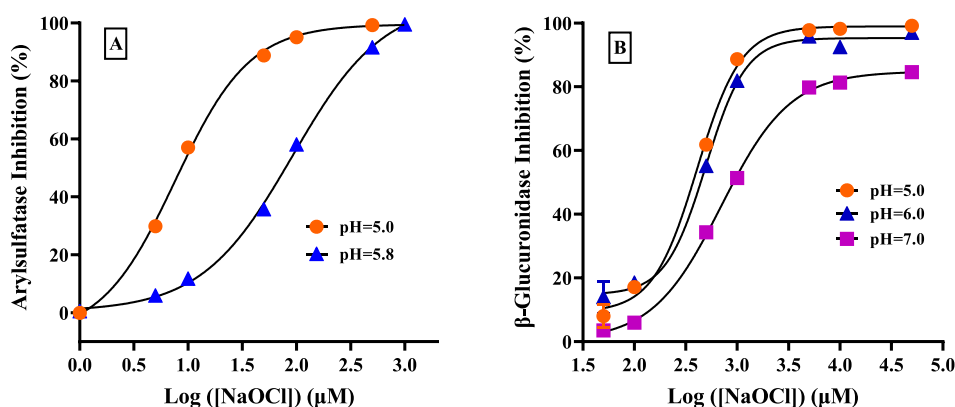


Figure 4. Dose–response curves of NaClO on arylsulfatase (A) and β -glucuronidase (B) under different pH conditions.

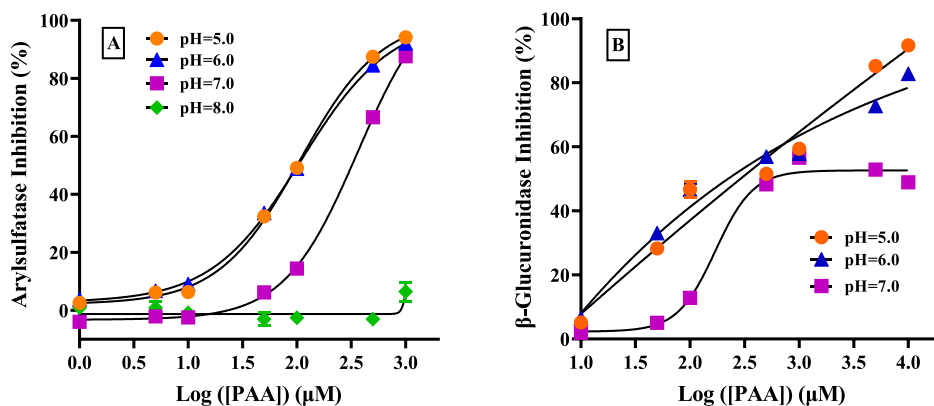


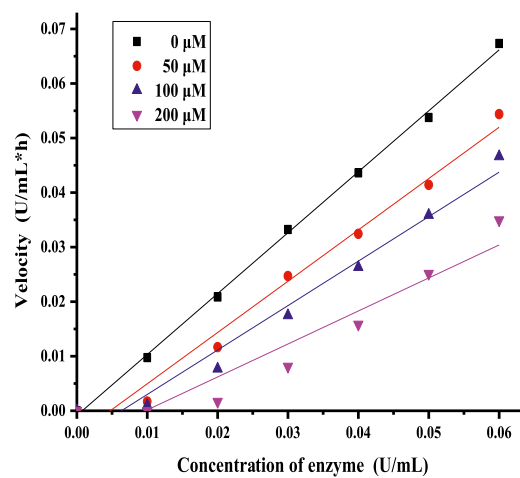
Figure 5. Dose–response curves of PAA on arylsulfatase (A) and β -glucuronidase (B) under different pH conditions.

that can mimic, enhance, or inhibit the action of endogenous hormones that are responsible for maintaining homeostasis and controlling normal development.^{27,28,59} The most commonly investigated EDCs are those that can mimic the effects of steroid hormones, which often show different estrogenic potencies that can be determined with different bioassays.^{29–31} With the definition of estrogenic potency, it is convenient to identify which are strong EDCs and which are weak. The potential adverse effect of each EDC depends on both its estrogenic potency and concentration; thus, the concept of estrogen equivalence has been widely adopted.^{32–34,60} This above criterion well describes the potential adverse effect of one individual EDC at any given concentration. However, many EDCs cannot be well explained with the above conception. For example, di-2-(ethyl hexyl) phthalate, well known as DEHP, the estrogenic potency of which is as low as 10^{-7} , is suggested as an extremely weak EDC.³² Considering its possible human exposure level to normal population, DEHP as an EDC unlikely poses potential adverse effects to humans. Nevertheless, epidemiological studies have shown that DEHP can pose different adverse effects on humans, including reproductive, developmental, and cardiovascular systems.^{35–38} Another example is cadmium, which possesses no estrogenic potency at all, but it is a well-known EDC.^{39–41}

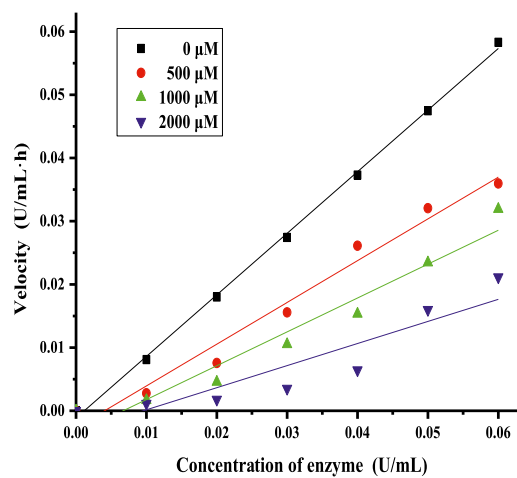
Many studies have shown that both DEHP and cadmium could induce the production of H_2O_2 in humans,^{42–46} whereas the existence of H_2O_2 can further induce hypochlorite.⁴⁷ Based on the fact that H_2O_2 and hypochlorite can effectively inhibit arylsulfatase and/or β -glucuronidase, this may suggest that DEHP and cadmium can disturb the human endocrine system

via the production of H_2O_2 and hypochlorite. This may well explain why DEHP and cadmium with very weak or no estrogenic potency can act as two EDCs that can pose adverse effects on humans. Not limited to DEHP and cadmium, any agent that can induce the production of H_2O_2 and hypochlorite in humans or animals is a potential EDC.

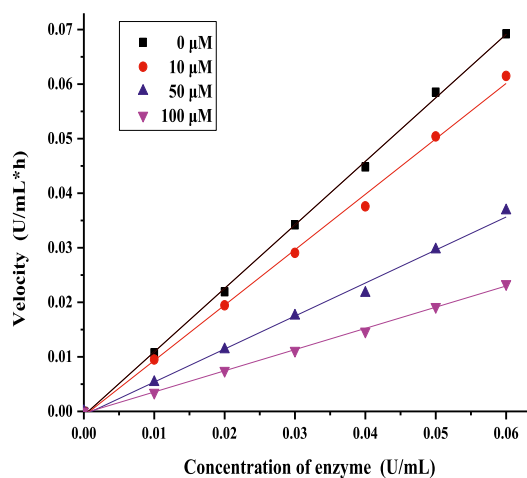
Considering the importance of H_2O_2 and hypochlorite in human body, studies relating their human blood concentration were summarized. As shown in Table 1, the reported concentrations of H_2O_2 in human blood samples varied greatly, which ranged from 0.1 to 6050 μM . Among these, some exceeded the IC_{50} value that is reported in this work for arylsulfatase and β -glucuronidase, which clearly suggests that H_2O_2 in human body can act as an EDC via inhibition of arylsulfatase. Compared to H_2O_2 , hypochlorite showed a much stronger inhibition to arylsulfatase, simultaneously showing inhibition to β -glucuronidase, which are all pH-dependent. As hypochlorite can also be endogenously produced through peroxidation of chloride ion by the catalysis of the enzyme myeloperoxidase in leukocytes including macrophages, monocytes, and neutrophils, the adverse effect of any potential EDC, via induction of H_2O_2 and hypochlorite, is likely enhanced when the production of hypochlorite is at a more favorable condition. It should be noted that because of the instability of H_2O_2 in human blood as well as the matrix interferences, not all the concentration data summarized in Table 1 are correct.¹⁹ However, it is still believable that H_2O_2 and hypochlorite in humans can act as two special EDCs via the inhibition of arylsulfatase and/or β -glucuronidase, which further suggests that any agent that can induce the production of H_2O_2 or



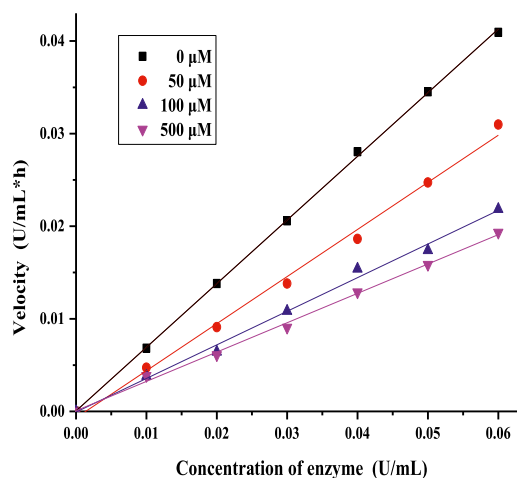
NaOCl — Arylsulfatase



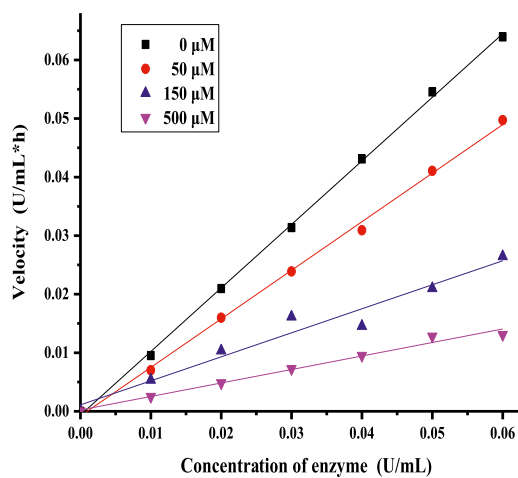
NaOCl — β-Glucuronidase



PAA — Arylsulfatase



PAA — β-Glucuronidase

Figure 6. Inhibitions of H₂O₂, NaClO, and PAA with different concentrations on arylsulfatase and/or β-glucuronidase.

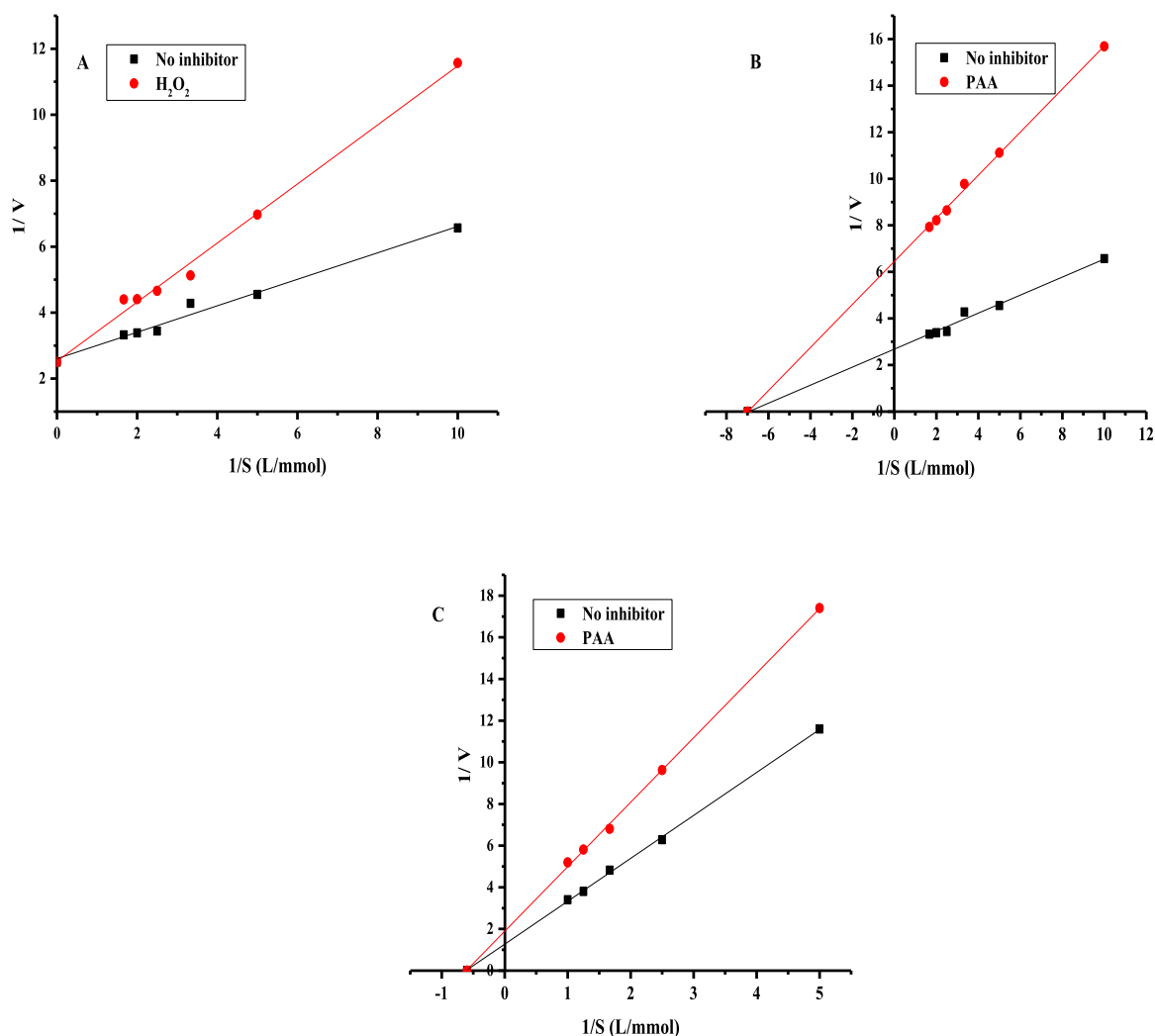


Figure 7. Lineweaver–Burk plots of the inhibition reaction of arylsulfatase in the presence of H_2O_2 (A) and PAA (B). Lineweaver–Burk plots of the inhibition reaction of β -glucuronidase in the presence of PAA (C).

Table 1. Hydrogen Peroxide in Humans Reported by Different Studies

number	objective	sample type	sample size	concentration (μM)	reference
1	man	plasma	1	4.825	48
2	man	plasma	1	5.5	49
3	men	plasma	17	5.85–7.15	50
4	men	plasma	50	2.14–3.15	51
5	men and women	plasma	236	0.61–6.79	52
6	children	plasma		1.4–2	53
7	men and women	plasma	60	2.5–6.2	54
8	men and women	plasma	30	21–113	55
9	men and women	plasma	53	30.5–50.3	56
10	pregnant women	plasma	31	50.1–66.9	57
11	men	blood	6	114–577 (288)	58
12	men	plasma	6	13–57 (34)	58

hypochlorite is a potential EDC. To the best of our knowledge, this is the first report that demonstrates that H_2O_2 , hypochlorite, and PAA can effectively inhibit arylsulfatase and/or β -

glucuronidase, which further suggests that some EDCs may act as endocrine disruptors via the induction of H_2O_2 and hypochlorite.

CONCLUSIONS

It was demonstrated for the first time that H_2O_2 , hypochlorite, and PAA could effectively inhibit arylsulfatase and/or β -glucuronidase. The following conclusions can be drawn.

- (1) Hypochlorite and PAA at the concentration level of μM could significantly inhibit both arylsulfatase and β -glucuronidase, whereas H_2O_2 was only effective for inhibiting arylsulfatase.
- (2) The inhibition of H_2O_2 and PAA to arylsulfatase and β -glucuronidase was found to be reversible, whereas the inhibition was found to be irreversible for hypochlorite.
- (3) H_2O_2 , hypochlorite, and PAA may behave as special endocrine disruptors through the inhibition of arylsulfatase and/or β -glucuronidase. This indeed led to an extended family of known endocrine disruptors.

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Notes

The authors declare no competing financial interest.

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